



4300 B Street, Suite 600  
Anchorage, AK 99503-5922 USA  
907-563-3322 Fax 907-563-3320

## After-Action Report

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Date 15 May 2013

To Meseret Ghebreslassie, USACE

From Kevin Maher, Jacobs

CC Pat Roth, 611 CES/CEAR

**Subject After-Action Report on the Supplemental Remedial Investigation at Source Area SS67 (Final)**

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This After-Action Report describes July to August and November 2012 activities conducted to supplement the remedial investigation at Source Area SS67 on Eielson Air Force Base (AFB) under Hazardous, Toxic, or Radioactive Waste (HTRW) Contract No. W911KB-11-D-0005, Task Order 07. The general site vicinity is shown on Figure 1 (Attachment 1). Field activities were documented in logbooks and on field data collection forms (Attachments 2 and 3).

Site work during the 2012 field season was conducted at SS67 to address the data gaps identified in the *Record of Decision (ROD) Five-Year Review* (U.S. Air Force [USAF] 2008). This document presents the data collected based on the following ROD recommendations:

- Sample fish tissue from Garrison Slough, Piledriver Slough, Chatanika River, and the Chena River (occurred from July to August 2012)
- *Perform MULTI INCREMENT<sup>1</sup>* sediment sampling of Garrison Slough decision units (DU) (re-scheduled for summer 2013 due to weather conditions)

Site work was conducted in accordance with the *Draft-Final Quality Assurance Project Plan, Remedial Investigation, Source Area SS67* (USAF 2012c), and the *Final Supplement to Quality Assurance Project Plan, Remedial Investigation, Source Area SS67* (USAF 2012b). Together, these documents constitute the Work Plan for the tasks described in this After-Action Report.

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<sup>1</sup> *MULTI INCREMENT*<sup>®</sup> is a registered trademark of EnviroStat, Inc.

**Introduction**

Source Area SS67 is part of Garrison Slough, which generally runs south to north through Eielson AFB. SS67 is an area approximately 1,500 feet long where elevated concentrations of polychlorinated biphenyls (PCB) have been found. A ROD was completed for source area SS67 in 1995. The ROD identified that the most likely PCB exposure pathway for human health risk at Garrison Slough was the ingestion of fish (USAF 1995). The ROD Five-Year Review (USAF 2008) made recommendations that were then outlined in the *Phase I Remedial Investigation Management Plan, Source Area SS67, Eielson Air Force Base, Alaska* (USAF 2012a):

- Determine the magnitude and spatial extent of PCBs, pesticides, and mercury in sediments at SS67 and in other portions of Garrison Slough.
- Establish the current concentration of PCB Aroclor 1260 in fish tissue of Garrison Slough.
- Investigate the possibility of additional sources of PCB contamination in fish tissue by evaluating the PCB concentration in fish from other water bodies in the vicinity of Garrison Slough.
- Perform human health and ecological risk assessments.
- Evaluate the feasibility of additional remedial actions to reduce PCB concentrations in Garrison Slough sediments to levels that assist with meeting the final remedial action objective for fish tissue.

The following sections describe Work Plan deviations, field conditions, preliminary sediment sampling investigation, fish tissue investigation, and waste management activities.

**Work Plan Deviations**

Deviations from the Work Plan (USAF 2012b, 2012c) that occurred during field activities included sediment sampling completion and fish tissue holding times, which are discussed below.

Sediment Sampling Completion

Sediment sampling was initiated in November 2012 but was not completed due to safety concerns resulting from the winter conditions of the slough as described in the Field Conditions section of this report. *MULTI INCREMENT* sampling of Garrison Slough is scheduled to resume in summer 2013.

Fish Tissue Holding Times

Fish tissue samples were collected and stored in accordance with the *Quality Assurance Project Plan, Remedial Investigation, Source Area SS67 (USAF 2012c)*. However, tissue samples exceeded the prescribed holding times specified for mercury (28 days) and pesticides (14 days) with holding times from 171 to 181 days and 178 to 190 days, respectively. Data are still considered usable because the samples were kept frozen.

**Field Conditions**

In July and August 2012, ambient temperatures ranged from 33 to 63 degrees Fahrenheit (°F).

In November 2012, ambient temperatures ranged from 3 to -33 °F. Water moving through Garrison Slough at DU7, DU8, and DU9 ranged from 0.5 to 3.5 feet deep with patches of ice and/or snow bridging across the water surface (Attachment 4, Photograph 1). Sediment sub-sample positions were recorded using a real-time kinematic global positioning system receiver that provides sub-meter accuracy.

A Base Civil Engineer Work Clearance Request was completed for the proposed sampling locations prior to fieldwork.

**Preliminary Sediment Sampling Investigation**

*MULTI INCREMENT* sediment samples from DU7, DU8, and DU9 were intended to address the uncertainties of the magnitude and spatial extent of PCBs, pesticides, and mercury in sediments at SS67 and other portions of the Garrison Slough. The targeted DUs are shown on Figure 2 (Attachment 1).

Field activities were halted due to safety concerns caused by the extreme winter weather discussed in the Field Conditions sections of this report. Sediment samples and analytical results from the abbreviated field effort are discussed below. Remaining sampling is scheduled for summer 2013.

Sediment Sampling

For sediment sampling purposes, each DU was split into 30 increments perpendicular to the course of Garrison Slough (USAF 2012b, 2012c). Each increment was further split into three

segments running parallel to Garrison Slough, randomly numbered 1, 2, or 3. Segment samples, once collected, were combined based on the random numbering to generate primary, duplicate, and triplicate *MULTI INCREMENT* samples.

Initially, segment samples were to be gathered using a manual soil coring device operated by personnel in waders. Due to weather and environmental conditions, waders were abandoned and a small boat was used as a sampling platform. Six segment samples were acquired in November 2012 from two increments within DU7. Those individual segment samples, collected from segments 7-1-1, 7-1-2, 7-1-3, 7-2-1, 7-2-2, and 7-2-3, were composited into a single sample and characterized for waste disposal purposes. Sampling activities are shown in photograph 5 (Attachment 4).

Sediment Sample Analytical Results

The sample was analyzed for PCBs and metals by U.S. Environmental Protection Agency (EPA) analytical test Methods SW8082 (PCBs) and SW7471/6020 (metals). Analytical results were compared to the project action levels (PAL) defined in the Work Plan (USAF 2012b, 2012c). Two analytes from the sample exceeded PALs for PCBs (Aroclor 1260) and arsenic. The results for analytes that exceed the PALs are presented in Table 1. The complete analytical results and a Data Quality Assessment are presented in Attachment 5.

**Table 1  
Exceedance of Cleanup Levels in Sediment Sample 12EAFB-SS67-SO-W01**

Sample Identification	Method	Analyte	PAL (mg/kg) <sup>1</sup>	Results (mg/kg)
12EAFB-SS67-SO-W01	SW6020	Arsenic	3.9	140
12EAFB-SS67-SO-W01	SW8082	PCB (1260)	0.14	19 JS-

**Notes:**

<sup>1</sup> Project Action Limit or ADEC cleanup level if no project action limit specified.

JS- = Value is estimated due to low surrogate recovery

mg/kg = milligram per kilogram

**Fish Tissue Investigation**

Fish tissue sampling activities were conducted by EA Engineering, Science, and Technology, Inc. in July and August 2012. Sampling activities were designed to determine whether other PCB sources contributed to the PCB concentrations found in fish tissues by evaluating the fish from other water bodies in the vicinity of Garrison Slough (USAF 2008). Figure 3 shows

the approximate locations where fish tissue samples were collected (Attachment 1). Fish tissue sampling procedures, analytical results, and fish aging analysis are discussed below.

#### Fish Tissue Sampling

Eighty arctic grayling were caught from Garrison Slough, Piledriver Slough (which runs parallel to Garrison Slough to the southwest), Chatanika River, and Chena River. Fillets were collected and submitted to the laboratory in accordance with the Work Plan (USAF 2012c).

#### Fish Tissue Analytical Results

Fish tissue samples were analyzed for PCBs by EPA Method SW8082A, organochlorine pesticides by EPA Method 8081B, and mercury by EPA Method SW7471B. Analytical results were compared the PALs defined in the Work Plan (USAF 2012c). Results are summarized below and presented in Table 2. Complete analytical results and a Data Quality Assessment are provided in Attachment 5.

The PCBs Aroclor 1254 and Aroclor 1260 were detected in fish samples from the four water bodies. Aroclor 1254 was detected above the PALs in 42 fish tissue samples from the four water bodies. Detections above the PAL occurred in order of decreasing frequency in Garrison Slough, Piledriver Slough, Chena River, and Chatanika River. Aroclor 1260 was detected above the PAL in 52 fish tissue samples from the four water bodies. Detections above the PAL occurred in order of decreasing frequency in Garrison Slough, Piledriver Slough, Chena River, and Chatanika River. Review of the Aroclor 1254 detections indicated that the majority were based on incomplete chromatogram patterns and on peak ratios that were inconsistent with the calibration standards. The laboratory was contacted regarding Aroclor 1254 detections and a review of the initial findings was requested. Review of the initial findings confirmed the original interpretation.

Organochlorine pesticides were detected in samples from the four water bodies.

Dichlorodiphenyldichloroethane (DDD), dichlorodiphenyldichloroethene (DDE), and dichlorodiphenyltrichloroethane (DDT) were detected above PALs in order of decreasing frequency in Garrison and Piledriver Sloughs. The pesticide dieldrin was detected above the PAL in order of decreasing frequency in Garrison Slough and Chatanika River. Gamma-chlordane was detected above the PAL in Garrison Slough. Heptachlor epoxide was detected above PAL in order of decreasing frequency in Garrison and Piledriver Sloughs.

Mercury was detected above the PAL in 79 of 80 fish tissue samples, and is considered ubiquitous in the four water bodies.

**Table 2  
Summary of Fillet Fish Tissue Detections**

Analyte	PAL Exceedances <sup>1</sup>	PAL (mg/kg)	RAO Exceedances <sup>2</sup>	Sample Results (mg/kg)		
				Mean	Min.	Max.
<b>Garrison Slough (20 Samples Taken)</b>						
Lipids (%)	--	--	--	2.15	0.49	6.00
4,4'-DDD	20	0.013	--	0.1241	0.03	0.34
4,4'-DDE	19	0.0093	--	0.0456	0.01	0.13
4,4'-DDT	1	0.0093	--	0.013	0.013	0.013
Dieldrin	10	0.0002	--	0.0026	0.00022	0.0061
gamma-Chlordane	8	0.009	--	0.0256	0.013	0.051
Heptachlor epoxide	8	0.00035	--	0.0012	0.00083	0.002
Mercury	20	0.014	--	0.0358	0.018	0.059
Aroclor 1254	20	0.0016	--	0.0773	0.013	0.19
Aroclor 1260	20	0.00269	20	0.1709	0.067	0.36
<b>Piledriver Slough (20 Samples Taken)</b>						
Lipids (%)	--	--	--	1.06	0.11	3.00
4,4'-DDD	3	0.013	--	0.055	0.017	0.12
4,4'-DDE	2	0.0093	--	0.03	0.011	0.049
4,4'-DDT	1	0.0093	--	0.016	0.016	0.016
Heptachlor epoxide	1	0.00035	--	0.0021	0.0021	0.0021
Mercury	20	0.014	--	0.0557	0.027	0.13
Aroclor 1254	9	0.0016	--	0.0217	0.0028	0.12
Aroclor 1260	13	0.00269	13	0.0266	0.0027	0.23
<b>Chatanika River (20 Samples Taken)</b>						
Lipids (%)	--	--	--	1.34	0.14	3.30
Dieldrin	1	0.0002	--	0.0006	0.00058	0.00058
Mercury	20	0.014	--	0.09	0.046	0.22
Aroclor 1254	5	0.0016	--	0.0265	0.0036	0.089
Aroclor 1260	9	0.00269	9	0.0107	0.0028	0.02
<b>Chena River (20 Samples Taken)</b>						
Lipids (%)	--	--	--	1.17	0.08	6.6
Mercury	19	0.014	--	0.198	0.048	0.48
Aroclor 1254	8	0.0016	--	0.0116	0.0019	0.039
Aroclor 1260	10	0.00269	10	0.0107	0.0034	0.029

**Notes:**

<sup>1</sup> PAL = Project Action Limit, (2012c)

<sup>2</sup> RAO = Remedial Action Objective as described in the 2008 ROD Five Year Review for Aroclor 1260 (USAF 2008)

mg/kg = milligrams per kilogram

-- = Not Established

**Fish Aging Analysis**

Fish aging analysis was conducted according to methods and procedures described in the Work Plan (2012c). Results of the fish aging analysis are presented in *Results of Fish Aging Using Otolith Analysis* (EA Engineering, Science, and Technology Inc. 2013), and highlighted in this section. Arctic grayling from the four water bodies were mature adults between three and ten years of age. Statistics for each fish sample are displayed on Table 5 in Appendix 5. The mean statistics for each water body are tabulated below in Table 3.

**Table 3  
Mean Fish Aging Analysis Statistics**

<b>Water Body</b>	<b>Age (years)</b>	<b>Length (mm)</b>	<b>Weight (grams)</b>
Garrison Slough	4.5	304.6	251.8
Piledriver Slough	4.8	298.7	214.5
Chatanika River	5.4	315.0	259.7
Chena River	5.4	324.2	279.8

**Note:**

mm = millimeters

Overall, grayling growth rates in the four water bodies were comparable to other grayling growth rates observed in Alaska (Carlander 1969; Gryska 2004). Although PCBs are known to bioaccumulate in living tissue, no direct correlation between fish age and the concentration of Aroclors 1254 or 1260 was evident (Figure 4 in Attachment 1,). Complete results of the fish aging analysis are presented in Table 5 (Attachment 5).

**Waste Management**

Sediment sampling activities produced minimal sediment and investigation-derived waste (IDW). Sediment waste was composited and submitted for analytical characterization. IDW consisted of used personal protective equipment, disposable sampling supplies, and direct-push acetate core liners from the manual coring device. The IDW was placed into one Super Sack® (12EAFB-SS67-SS01) totaling 0.66 tons.

Waste was removed from temporary storage locations on Eielson AFB on 19 February 2013 along with waste generated by investigation activities for source area WP45/SS57.

Environmental and Logistics Management, Inc. (ELM) and was disposed of in the Columbia

Ridge Landfill in Arlington, Oregon. The associated waste manifest and certificate of disposal are included in Attachment 6.

## Conclusions

Sediment sampling of Garrison Slough DU7, DU8, and DU9 will be completed in summer 2013 along with investigations of the other DUs described in the Work Plan (USAF 2012c). The objective of fish sampling was to investigate the possibility of other sources of PCB contamination in fish tissue by evaluating the concentration of PCBs in fish from other water bodies in the vicinity of Garrison Slough. Tissue sample data collected and the performance of an aging analysis indicated that Garrison Slough contained the highest PCB results from the four water bodies, but that results also exceed the PAL in the other areas. Data also suggested that the 0.00266 mg/kg remedial action objective for PCB cleanup and the 0.0135 mg/kg PAL for mercury may be difficult to achieve at Garrison Slough.

## References

- Carlander, K.D. 1969. *Handbook of Freshwater Fishery Biology*, Volume I. Iowa State University Press, Ames, Iowa.
- EA Engineering, Science, and Technology Inc.. 2013 (February). *Results of Fish Aging Using Otolith Analysis, Eielson Air Force Base, Alaska*.
- Gryska A. D. 2004. *Abundance and Length and Age composition of Arctic Grayling in the Snake River, 2001*. Alaska Dept. of Fish and Game, Fishery Data Series No. 04-15, Anchorage.
- USAF (U.S. Air Force) 2012a (December). *Phase I Remedial Investigation Management Plan, Source Area SS67, Eielson Air Force Base, Alaska*. Final. Prepared by EA Engineering, Science, and Technology, Inc.
- USAF. 2012b (November). *Supplement to Quality Assurance Project Plan for Remedial Investigation Source Area SS67*. Final. Prepared by Jacobs Engineering Group Inc.
- USAF. 2012c (June). *Draft Final Quality Assurance Project Plan Remedial Investigation Source Area SS67*. Prepared by EA Engineering, Science, and Technology, Inc.
- USAF 2008 (September). *Five-Year Record of Decision Review Report*. Final.
- USAF 1995 (September). *Eielson Air Force Base Operable Units 3, 4, and 5 Record of Decision*.



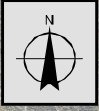
**Attachments**

- Attachment 1 Figures
- Attachment 2 Field Logbooks
- Attachment 3 Field Data Collection Forms
- Attachment 4 Photograph Log
- Attachment 5 Analytical Data and Data Quality Assessment
- Attachment 6 Waste Tracking

## **ATTACHMENT 1**

### **Figures**

- Figure 1 Site Location and Vicinity
- Figure 2 Sediment Sampling (Decision Units 7 through 9)
- Figure 3 Fish Tissue Sampling Water Bodies
- Figure 4 Fish Age and PCB Concentrations in Four Water Bodies





Main Entrance

Jacobs Field Office

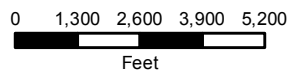
SS67 (Garrison Slough)

Airfield

**Legend**

-  Source Area
-  Eielson AFB Boundary

All Locations Are Approximate



WGS 1984 UTM Zone 6N  
Imagery: Provided by  
Eielson AFB GeoBase

**Site Location and Vicinity**

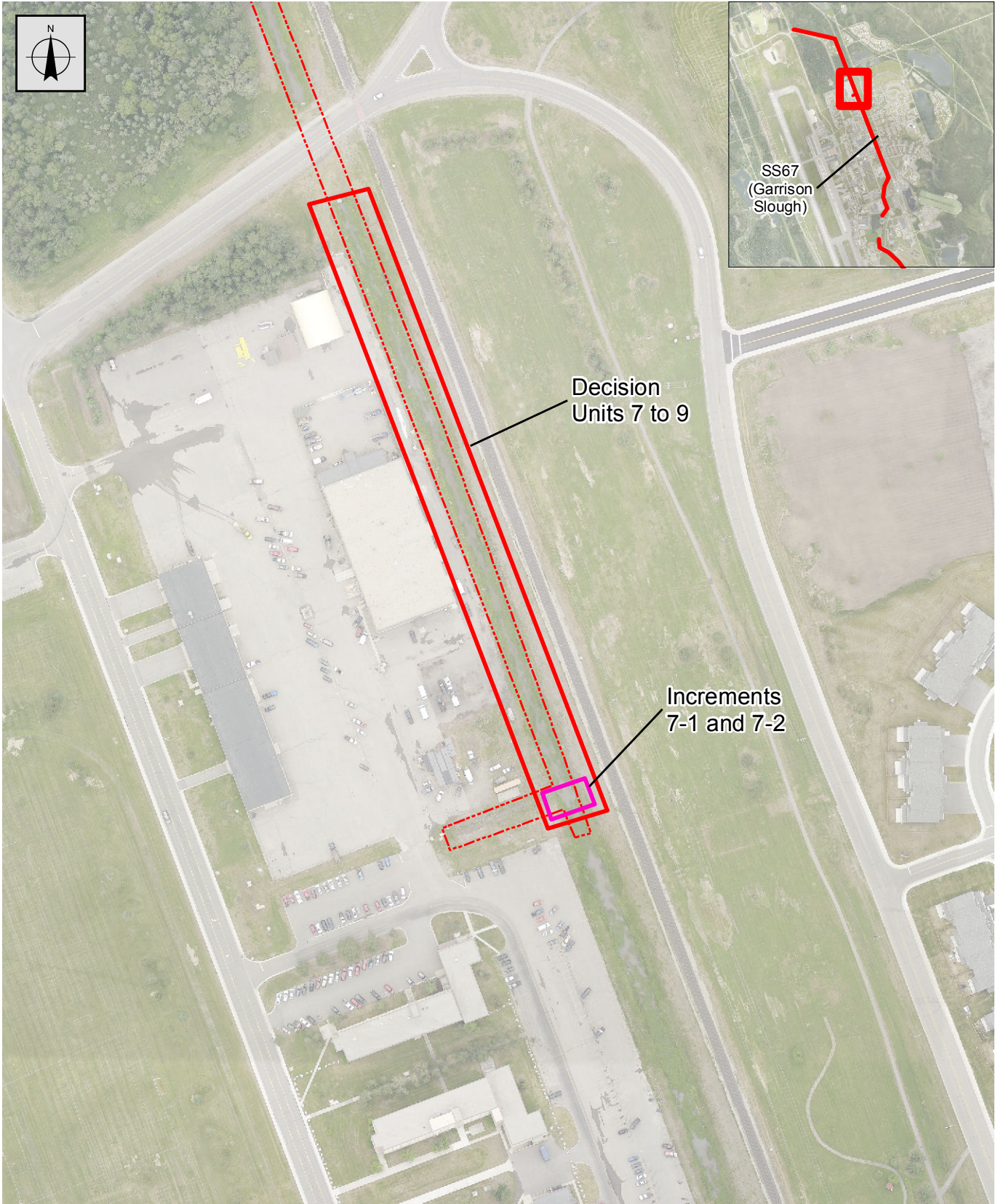
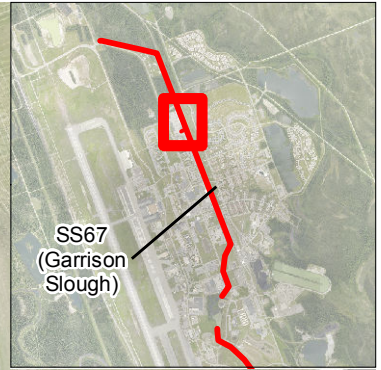
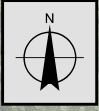
Eielson Air Force Base, Alaska



DATE:  
18 MAR 2013



PROJECT MANAGER:  
K. Maher

FIGURE NO.  
1



P:\AK\_Eielson\_AF\GIS\IMXD\13E1-SS67\_AA-RandFishTissue\Figure2\_DU7-9.mxd mckeynt

**Legend**

-  Source Area
-  Sediment Sub-sample Area (DU 7-1 and DU 7-2)

All Locations Are Approximate  
 0 60 120 180 240  
 Feet

WGS 1984 UTM Zone 6N  
 Imagery: 2012, 4 inch pixel

**Source Area SS67**  
**Sediment Sampling**  
**Decision Units 7 to 9 and Increments 7-1 and 7-2**  
 Eielson Air Force Base, Alaska

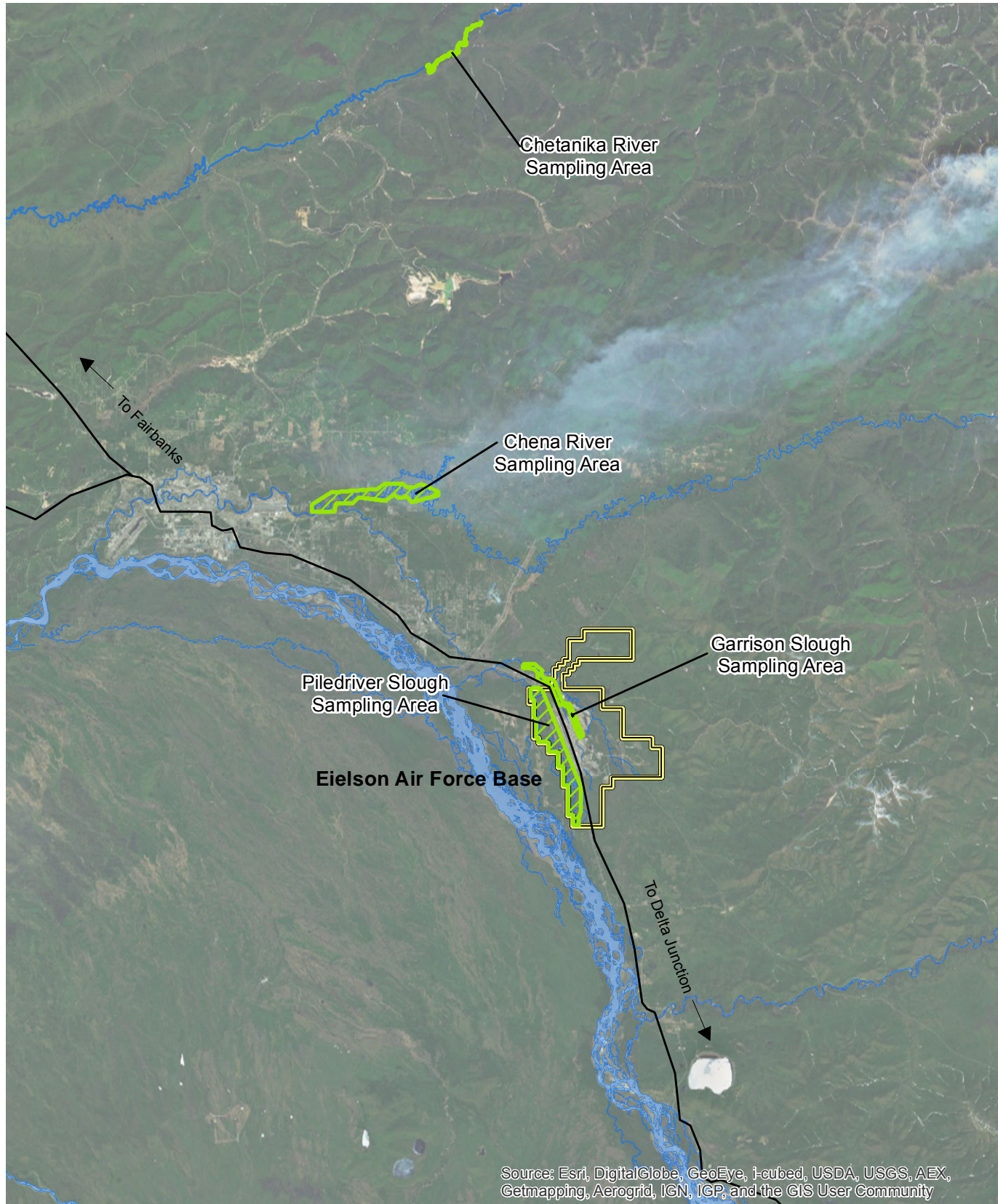
**JACOBS**

DATE:  
15 MAY 2013

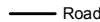


PROJECT MANAGER:  
K. Maher

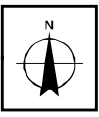
FIGURE NO.  
2

P:\AK\_Eielson\_AFB\GIS\IMXD\13E1-SS67\_AA\_Rand\Fish Tissue\Figure 3\_Fish Tissue Water Bodies.mxd felici

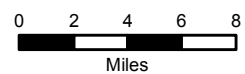


**Legend**

-  Road
-  Sampling Area
-  Eielson AFB Boundary



All Locations Are Approximate

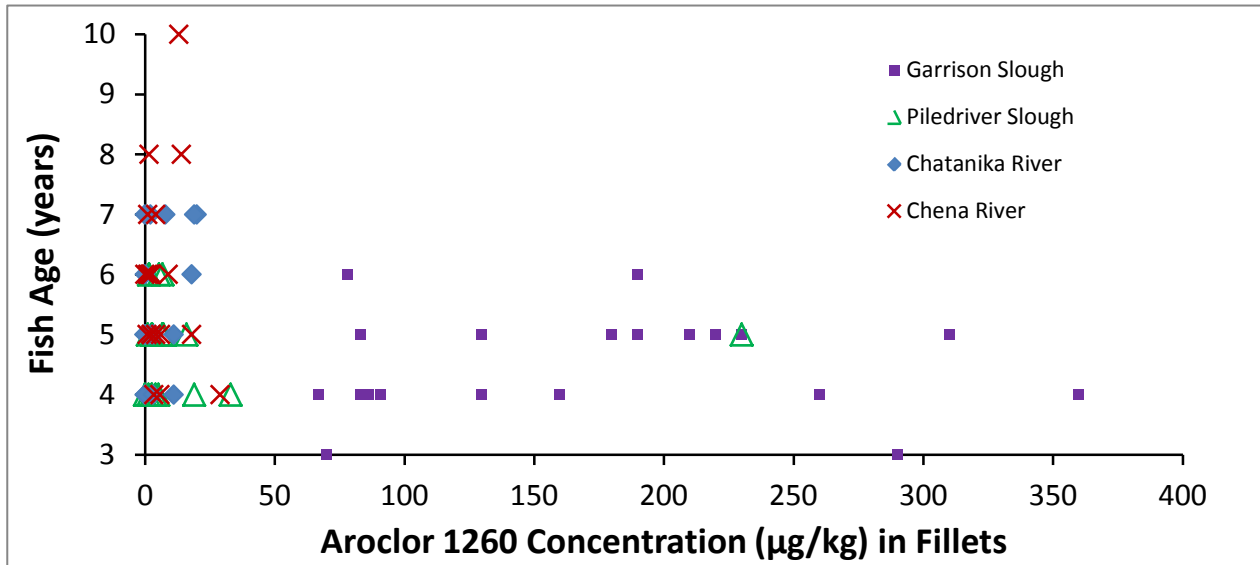
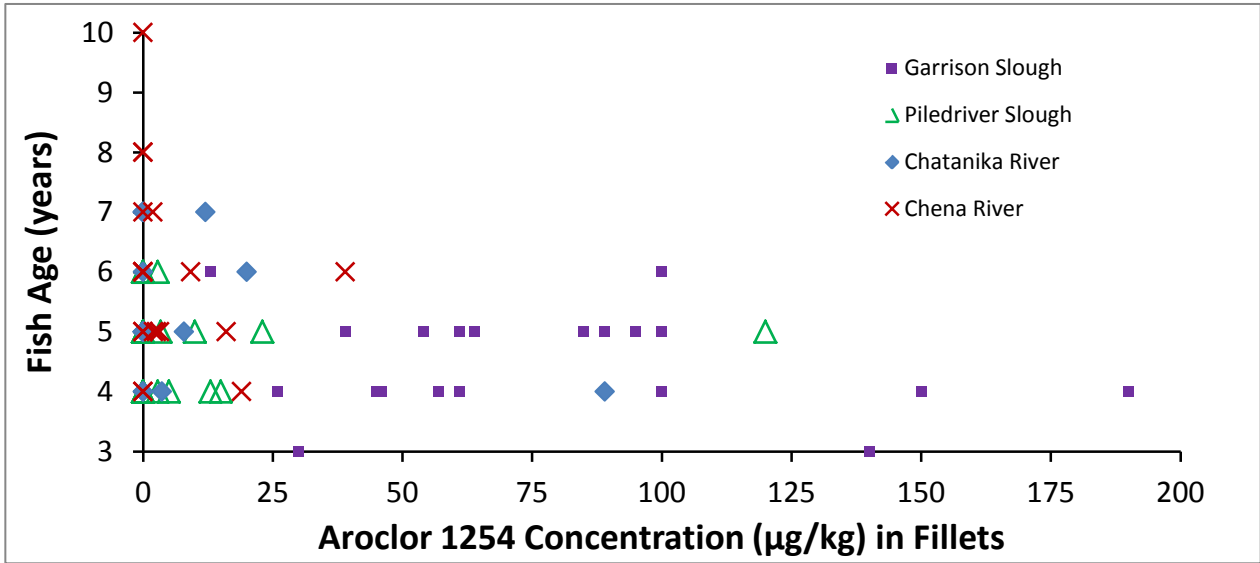


WGS 1984 UTM Zone 6N

**Source Area SS67  
Fish Tissue Sampling  
Water Bodies**

Eielson Air Force Base, Alaska

<b>JACOBS</b>	DATE:	PROJECT MANAGER:	FIGURE NO.
	18 MAR 2013	K. Maher	3



**Notes:**

Fish age was not related to PCB concentrations of Aroclor 1254 (top) or Aroclor 1260 (bottom) in fillet tissue samples at any water body.

**Fish Age and PCB Concentrations  
at the Four Water Bodies**

EIELSON AFB, ALASKA

**ATTACHMENT 2**  
**Field Logbooks**

Eielson Surface Sampling

~~Summary~~  
Field Observation  
Logbook #1

OSF4570



*R. L. ...*  
U.S. ENVIRONMENTAL  
PROTECTION AGENCY  
FIELD BOOK  
#550

C. Feil  
N. McKay  
R. Plate  
C. Jelle  
I. Peterson  
C. Peek

HTV-107-OSF4570-1104-0001





Location \_\_\_\_\_ Date \_\_\_\_\_

Project / Client \_\_\_\_\_

Project &amp; Emergency Contacts.

## PROJECT CONTACTS

Organization	Title	Name	Telephone
USACE	Contracting Officer's Representative	Scott Kendall	907-753-5661 (o)
USACE	Project Manager	Meseret Ghebreslassie	907-753-2670 (o)
AFCEC	Project Manager	Pat Roth	907-552-7893 (o)
Jacobs	USAF Program Coordinator	Kelly McGovern	907-751-3350 (o) 907-227-7833 (m)
	Project Manager	Kevin Maher	907-751-3429 (o) 907-632-8289 (m)
	Field Services Manager	Jon McVay	907-751-3395 (o) 907-230-5395 (m)
	Health and Safety Manager	Randall Jones	907-751-3332 (o) 931-607-3415 (m)
	Site Manager	David Saltee	907-451-0550 (o) 907-978-4291 (m)
	Site Safety and Health Officer	David Saltee	907-451-0550 (o) 907-978-4291 (m)
	Technical Lead	Katie Bloom	907-751-3360 (o) 907-382-3645 (m)
	Project Geologist	Dave Ward	907-751-3389 (o) 907-242-0775 (m)
	Lead Sampler	Ida Petersen	907-451-0550 (o) 907-687-0456 (m)
	Project Chemist	David Summerville	907-751-3342 (o) 907-632-8286 (m)

## EMERGENCY RESPONSE CONTACTS

<b>Medical and Fire Emergency</b> <ul style="list-style-type: none"> <li>State that you are on Eielson AFB. The Fairbanks dispatcher will transfer you to the Eielson AFB dispatcher.</li> <li>Provide your exact location.</li> <li>State your full name, the particulars of your emergency, type of first-aid being given, and your requirements for assistance. Be patient and deliberate in your communication to ensure clarity.</li> <li>Listen to, record, and follow further instructions.</li> </ul> DO NOT HANG UP until the dispatcher directs you to hang up.	911
Fairbanks Memorial Hospital	907-452-8181
Eielson AFB Police Department	911 or 907-377-1110 (base operator)
Eielson AFB Fire Department	911 or 907-377-1110 (base operator)
Poison Control Center	800-222-1222
Jacobs Emergency Medical Consultant (Dr. Baskous)	907-279-4953
<b>Spills and Toxins</b>	
National Response Center (Oil and Toxic Chemical Spills)	800-424-8802
Chemtel (Customer ID #JAC001)	800-424-9300
Poison Control Center	800-222-1222

Location SS67 Bldg 2230 Eielson Date 11/15/2012Project / Client SS67USTCCE

O&amp;CC Arrived on site

Setup to sample

Left  
Blank

6

Location Bldg 2230 Eielson AFB Date 11/15/2012  
 Project / Client SS67 USACE

1120	Colors.		
	Flagging Map	Prim, dup, triplicate	
	Yellow	Yellow	2
	Blue	Green	1
	Pink	Orange	3

1130 Collected subsample DU7-1-3  
 marked with pink flagging  
 Depth: 1.8ft Single tube, CW.

1140 Collected subsample DU7-1-1  
 marked with blue flagging  
 Depth: 2.0ft Single tube, CW.

1150 Collected subsample DU7-1-2  
 marked with pink flagging, Depth 1.2ft Single tube, CW.

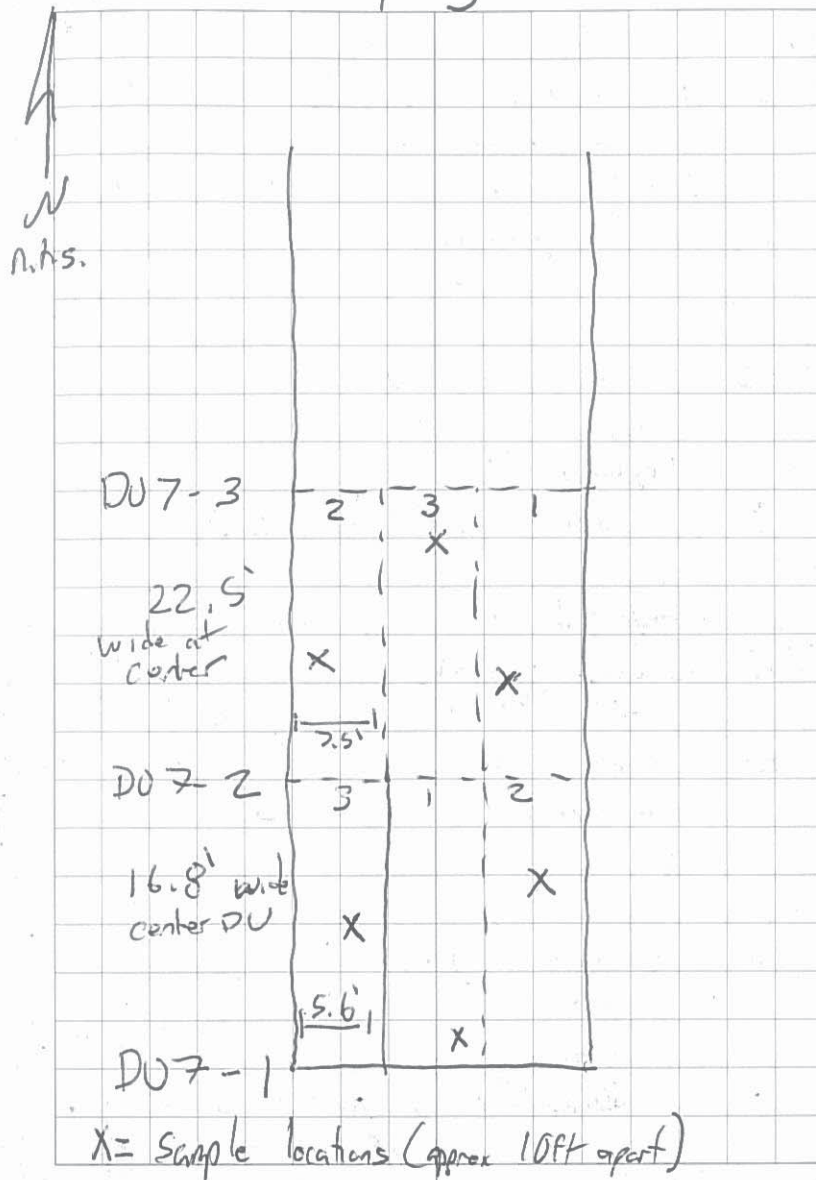
1200 Moved boat to next subsampled location  
 and sampling team took a warmup  
 break

1220 Resumed sampling activities.  
 - problems with coring device  
 freezing up.

1230 - thawed cutting shoe, went go back  
 on due to ice.

Location Bldg 2230 Eielson AFB Date 11/15/2012  
 Project / Client SS67 USACE

## DU 7 sampling



8

Location Bldg 2230 Eielson AFB Date 11/15/2012  
 Project / Client SS67 USACE

1233 Collected subsample DV7-2-2  
 marked with \_\_\_\_\_ flagging  
 Recovery: 0  
 Depth: 2 ft  
 Sampler: Single tube, C)

Red

1402 Collected subsample DV7-2-2  
 Flagging: Yellow  
 Depth: 2 ft  
 Recovery: 1.4 ft  
 Sampler: Single tube, C)

1425 Collected subsample DV7-2-3  
 Flagging: Pink  
 Depth: 2 ft  
 Recovery:  
 Sampler: Single tube, C)

1440 Collected subsample DV7-2-1  
 Flagging: Blue  
 Depth: 2 ft  
 Recovery:  
 Sampler: Single tube, C)

9

Location Bldg 2230 Eielson AFB Date 11/15/2012  
 Project / Client SS67 USACE

~~Collected subsample  
 Flagging:  
 Depth:  
 Recovery:  
 Sampler:~~ Did not collect

~~Collected subsample  
 Flagging:  
 Depth:  
 Recovery:  
 Sampler:~~ Did not collect

~~Collected subsample  
 Flagging:  
 Depth:  
 Recovery:  
 Sampler:~~ Did not collect

~~Charles D. Pell  
 11/15/2012~~

10

Location Bldg 2230 Eielson AFB Date 11/15/2012  
 Project / Client SSG7 USACE

1235 No recovery DU7-2-2  
 1238 Frozen H<sub>2</sub>O in tube, cannot reload a liner  
 1245 Took samples inside to throw  
 1255 Brake for Lunch  
 1340 Back from lunch  
 1400 Resumed sampling DU7-2-2  
 - icing issues  
 1425 Began sampling DU7-2-3  
 1440 Began sampling DU7-2-1

~~Christopher D Fall~~  
 11/15/2012

Location Bldg 2230 Eielson AFB Date 11/15/2012<sup>11</sup>  
 Project / Client SSG7 USACE

Did not  
 use phase

~~Christopher D Fall~~  
 11/15/2012

Location \_\_\_\_\_

Date

11/16/12

Project / Client \_\_\_\_\_

~~After~~ ADEC site visit on  
11/15 → questioning if it was  
worth money & safety to  
continue sampling in cold &  
ice

→ Air Force confers & decides  
to cancel sampling for  
now (USACE verbally confirms)

→ Samples from 7-1 & 7-2 will be  
saved until official letter comes  
saying to call off sampling

→ Team decons + clean up  
site for demob

*eg*

Location Eielson AFB, AK

Date

11/19/12

Project / Client

SS67

USACE

1300 Ida Petersen will <sup>send</sup> bring waste  
sample to Test America in Seattle  
via Gold Streak  
Composite sample from DV7, increments  
1 & 2 for waste characterization  
Add trash to 12EAFB-SS67-SS01

1330 Sample 12EAFB-SS67-SO-WO1  
1 8oz jar, Samplers: CJ; CF; IP  
SW7471/6020 (RCRA metals)  
SW8082 (PCBs)  
glass, amber, TAT=

*IP* 11/19/12

**ATTACHMENT 3**  
**Field Data Collection Forms**





## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

<b>Sampling Location (select):</b>	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input checked="" type="checkbox"/> Chatanika River
<b>GPS coordinate (UTM-WGS84):</b>	
<b>Date &amp; Time (24 hr)</b>	Date: 08/01/12                      Time: 1015
<b>Weather/ Temperature:</b>	clear 70's
<b>Tackle/Harvest Method (select):</b>	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
<b>Sample Collected By:</b>	SCHULTZ

### Morphometric Data

<b>Otolith Vial #:</b>	<del>CK-AG-02-OTOLITH</del> Not Collected.
<b>Scale Sample ID:</b>	CK-AG-02-SCALE
<b>Fresh Field Weight (g):</b>	193.5
<b>Length (cm):</b>	29.5
<b>Sex (select):</b>	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

<b>Fish Tissue Sample ID (fillet(s)):</b>	CK-AG-02-FILLET
<b>Fish Tissue Sample ID (remainder):</b>	CK-AG-02-WHOLE
<b>Number of Fillets:</b>	2
<b>Fillet Weight (g):</b>	78.0 g
<b>Remainder Weight (kg):</b>	115.5g

### Notes

Otolith not collected - couldn't locate

# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input checked="" type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: 08/01/12                      Time: 1049
Weather/ Temperature:	CLoud 70°
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	SCHULTZ

## Morphometric Data

Otolith Vial #:	<del>CR-AG-03-OTOLITH</del> Did not collect
Scale Sample ID:	CR-AG-03-SCALE
Fresh Field Weight (g):	174.1 g
Length (cm):	28.0
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CR-AG-03-FILLET
Fish Tissue Sample ID (remainder):	CR-AG-03-WHOLE
Number of Fillets:	2
Fillet Weight (kg): grams	76.1 g
Remainder Weight (kg):	98.0 g

## Notes

Fish too small to find otoliths. Did not collect.

## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input checked="" type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: 08/01/12                      Time: 1143
Weather/ Temperature:	CLEAR 70's
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	SCULTZ

### Morphometric Data

Otolith Vial #:	CK-AG-04-OTOLITH
Scale Sample ID:	CK-AG-04-SCALE
Fresh Field Weight (kg):	477.4
Length (cm):	39.5
Sex (select):	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CK-AG-04-FILLET
Fish Tissue Sample ID (remainder):	CK-AG-04-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	<del>101.0g</del> 209.3g
Remainder Weight (kg):	268.1g

### Notes

\*MS/MSD\*      Side A 101.0g                      Side B 108.3g

CONTACT LAB TO ENSURE THEY KNOW WE WANT A SAMPLE RESULT AND MS/MSD FOR THIS SAMPLE. IT IS THE LARGEST SAMPLE TO DATE.

## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input checked="" type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: 08/01/12                      Time: 1157
Weather/ Temperature:	clear 70's
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	SCHULTZ

### Morphometric Data

Otolith Vial #:	CK-AG-05-OTOLITH
Scale Sample ID:	CK-AG-05-SCALE
Fresh Field Weight (g):	349.4g
Length (cm):	34.5
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CK-AG-05-FILLET / CK-AG-05 <sup>21</sup> -FILLET (DUP)
Fish Tissue Sample ID (remainder):	CK-AG-05-WHOLE
Number of Fillets:	1 (2 <sup>ND</sup> FILLET WILL STAY AS DUP + BE LABELED)
Fillet Weight (kg):	83.7g / 81.0g for CK-AG-21                      CK-AG-21-F
Remainder Weight (kg):	265.7

### Notes

	<p>* THIS SAMPLE PROVIDES A SECOND FILLET FOR A DUPLICATE          - THE CONCENTRATION MUST BE ADDED BACK IN FOR WHOLE BODY CALCULATION</p>

# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input checked="" type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: 08/01/12                      Time: 1205
Weather/ Temperature:	Clear 70's
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	SCHULTZ

## Morphometric Data

Otolith Vial #:	CK-AG-06-OTOLITH
Scale Sample ID:	CK-AG-06-SCALE
Fresh Field Weight (kg):	183.9
Length (cm):	28.0
Sex (select):	<input type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CK-AG-06-FILLET
Fish Tissue Sample ID (remainder):	CK-AG-06-WHOLE
Number of Fillets:	2
Fillet Weight (g):	80.9 g
Remainder Weight (g):	103 g

## Notes


## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input checked="" type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: <u>08/01/12</u> Time: <del>1157</del> <u>1157</u>
Weather/ Temperature:	<u>clear 70's</u>
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	<u>SCHULTZ</u>

### Morphometric Data

Otolith Vial #:	<u>CK-AG-NA</u>
Scale Sample ID:	<u>NA</u>
Fresh Field Weight (kg):	<u>349.4</u>
Length (cm):	<u>34.5</u>
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	<u>CK-AG-21-Fillet</u>
Fish Tissue Sample ID (remainder):	<u>NA</u>
Number of Fillets:	<u>1</u>
Fillet Weight (kg):	<u>81.0g</u>
Remainder Weight (kg):	<u>NA</u>

### Notes

CK-AG-21-Fillet is the 2<sup>nd</sup> fillet of CK-AG-05 and serves as the duplicate. The concentrations detected must be mass normalized and added to with the fillet and whole concentrations of CK-AG-05







## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

<b>Sampling Location (select):</b>	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input checked="" type="checkbox"/> Chatanika River
<b>GPS coordinate (UTM-WGS84):</b>	
<b>Date &amp; Time (24 hr)</b>	Date: 08/01/12                      Time: 1412
<b>Weather/ Temperature:</b>	Partly Cloudy 70°
<b>Tackle/Harvest Method (select):</b>	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
<b>Sample Collected By:</b>	SCHULTZ

### Morphometric Data

<b>Otolith Vial #:</b>	CX-AG-09 - OTOLITH
<b>Scale Sample ID:</b>	CX-AG-09 - SCALE
<b>Fresh Field Weight (kg):</b>	171.7
<b>Length (cm):</b>	29.0
<b>Sex (select):</b>	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

<b>Fish Tissue Sample ID (fillet(s)):</b>	CX-AG-09-FILLET
<b>Fish Tissue Sample ID (remainder):</b>	CR-AG-09-WHOLE
<b>Number of Fillets:</b>	2
<b>Fillet Weight (kg):</b>	77.8 g
<b>Remainder Weight (kg):</b>	93.9 g

### Notes




## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input checked="" type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: 08/01/12                      Time: 1424
Weather/ Temperature:	Partly Cloudy 70°
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	SCITULTZ

### Morphometric Data

Otolith Vial #:	<del>CR-AG-11-OTOLITH</del> Broken
Scale Sample ID:	CR-AG-11-SCALE
Fresh Field Weight (kg):	222.9
Length (cm):	29.5
Sex (select):	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CR-AG-11-FILLET
Fish Tissue Sample ID (remainder):	CR-AG-11-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	<del>104.8</del> 104.8
Remainder Weight (kg):	118.1

### Notes

~~Otoliths Broken - not collected.~~ 201 BURN-TORN OBSERVED THAT OTOLITH RESIDUES INSIDE A COMPARTMENT THAT IS ~~OBSERVED~~ COVERED BY A THIN MEMBRANE OF TISSUE MAKING IT HARD DIFFICULT TO FIND. ONCE THIS WAS LUPRETTED, OTOLITHS WERE ABLE TO BE COLLECTED.

ADF&G Area Biologist: Audra Base-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input checked="" type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: 08/01/12      Time: 1426
Weather/ Temperature:	Partly Cloudy 70's
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	Voss

## Morphometric Data

Otolith Vial #:	<del>CR-AG-12-OTOLITH</del> not collected!
Scale Sample ID:	CR-AG-12-SCALE
Fresh Field Weight (kg):	237.0 g
Length (cm):	30.0
Sex (select):	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CR-AG-12-FILLET
Fish Tissue Sample ID (remainder):	CR-AG-12-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	100.9 g
Remainder Weight (kg):	136.1 g

## Notes

<p>could not locate otoliths - possibly cut through (P) - SEE OTOLITH EXPANSION OF CR-AG-11-OTOLITH</p>













# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

<b>Sampling Location (select):</b>	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input checked="" type="checkbox"/> Chatanika River
<b>GPS coordinate (UTM-WGS84):</b>	
<b>Date &amp; Time (24 hr) Sample Collected</b>	Date: 8/5/12                      Time: 958
<b>Weather/ Temperature:</b>	60 + Overcast
<b>Tackle/Harvest Method (select):</b>	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input checked="" type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
<b>Sample Collected By:</b>	Chris

## Morphometric Data

<b>Otolith Vial #:</b>	CK-AG-17-OTOLITH
<b>Scale Sample ID:</b>	CK-AG-17-SCALE
<b>Fresh Field Weight (kg):</b>	336.7
<b>Length (cm):</b>	35.5
<b>Sex (select):</b>	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

<b>Fish Tissue Sample ID (fillet(s)):</b>	CK-AG-17-FILLET
<b>Fish Tissue Sample ID (remainder):</b>	CK-AG-17-WHOLE
<b>Number of Fillets:</b>	1
<b>Fillet Weight (kg):</b>	<del>85.8</del> 85.8
<b>Remainder Weight (kg):</b>	

## Notes

Date Sample Shipped for Analysis:





## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

<b>Sampling Location (select):</b>	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input checked="" type="checkbox"/> Chatanika River
<b>GPS coordinate (UTM-WGS84):</b>	
<b>Date &amp; Time (24 hr) Sample Collected</b>	Date: 8/5/12                      Time: 1113
<b>Weather/ Temperature:</b>	
<b>Tackle/Harvest Method (select):</b>	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input checked="" type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
<b>Sample Collected By:</b>	Rich

### Morphometric Data

<b>Otolith Vial #:</b>	CK-AG-20-OTOLITH
<b>Scale Sample ID:</b>	CK-AG-20-SCALE
<b>Fresh Field Weight (kg):</b>	330.2
<b>Length (cm):</b>	34.0
<b>Sex (select):</b>	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

<b>Fish Tissue Sample ID (fillet(s)):</b>	CK-AG-20-FILLET
<b>Fish Tissue Sample ID (remainder):</b>	CK-AG-20-WHOLE
<b>Number of Fillets:</b>	1
<b>Fillet Weight (kg):</b>	74.0
<b>Remainder Weight (kg):</b>	

### Notes


**Date Sample Shipped for Analysis:**

ADF&G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: 08/03/12      Time: 1005
Weather/ Temperature:	Cloudy, Wind, 50°
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	CONNOR

### Morphometric Data

Otolith Vial #:	CR-AG-01-OTOLITH
Scale Sample ID:	CR-AG-01-SCALE
Fresh Field Weight (kg):	232.0g
Length (cm):	31.5
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CR-AG-01-FILLET
Fish Tissue Sample ID (remainder):	CR-AG-01-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	78.3g
Remainder Weight (kg):	153.7g

### Notes


ADF&G Area Biologist: Audra Base-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: 08/03/12      Time: 1045
Weather/ Temperature:	CLOUDY, WIND, 50°
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	CONNOR

## Morphometric Data

Otolith Vial #:	CR-AG-02-OTOLITH
Scale Sample ID:	CR-AG-02-SCALE
Fresh Field Weight (kg):	374.5g
Length (cm):	36.5
Sex (select):	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CR-AG-02-FILLET (MS/MSD)
Fish Tissue Sample ID (remainder):	CR-AG-02-WHOLE (MS/MSD)
Number of Fillets:	2
Fillet Weight (kg):	170.9g
Remainder Weight (kg):	203.6g

## Notes


ADF&G Area Biologist: Audra Base-(907) 459-7244

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EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: 08/03/12                      Time: 1100
Weather/ Temperature:	cloudy, wind, 50°
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	NEWBORN

### Morphometric Data

Otolith Vial #:	CR-AG-03-OTOLITH
Scale Sample ID:	CR-AG-03-SCALE
Fresh Field Weight (kg):	<del>144.2g</del> 143.9g
Length (cm):	26.0
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CR-AG-03-FILLET
Fish Tissue Sample ID (remainder):	CR-AG-03-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	72.1g
Remainder Weight (kg):	71.8g

### Notes


ADF&G Area Biologist: Audra Base-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

<b>Sampling Location (select):</b>	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
<b>GPS coordinate (UTM-WGS84):</b>	
<b>Date &amp; Time (24 hr)</b>	Date: 08/04/12                      Time: 1110
<b>Weather/ Temperature:</b>	cloudy, wind, 50°
<b>Tackle/Harvest Method (select):</b>	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
<b>Sample Collected By:</b>	V044

### Morphometric Data

<b>Otolith Vial #:</b>	CR-AG-04 - OTOLITH
<b>Scale Sample ID:</b>	CR-AG-04 - SCALE
<b>Fresh Field Weight (kg):</b>	298.5g
<b>Length (cm):</b>	34.0
<b>Sex (select):</b>	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

<b>Fish Tissue Sample ID (fillet(s)):</b>	CR-AG-04 - FILLET
<b>Fish Tissue Sample ID (remainder):</b>	CR-AG-04 - WHOLE
<b>Number of Fillets:</b>	1
<b>Fillet Weight (kg):</b>	73.2
<b>Remainder Weight (kg):</b>	225.3

### Notes




## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

<b>Sampling Location (select):</b>	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
<b>GPS coordinate (UTM-WGS84):</b>	
<b>Date &amp; Time (24 hr)</b>	Date: 08/03/12                      Time: 1113
<b>Weather/ Temperature:</b>	cloudy, wind, 50°
<b>Tackle/Harvest Method (select):</b>	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
<b>Sample Collected By:</b>	COMBURY

### Morphometric Data

<b>Otolith Vial #:</b>	CR-AG-05-OTOLITH
<b>Scale Sample ID:</b>	CR-AG-05-SCALE
<b>Fresh Field Weight (g):</b>	236.4g
<b>Length (cm):</b>	30.0
<b>Sex (select):</b>	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

<b>Fish Tissue Sample ID (fillet(s)):</b>	CR-AG-05-FILLET
<b>Fish Tissue Sample ID (remainder):</b>	CR-AG-05-WHOLE
<b>Number of Fillets:</b>	2
<b>Fillet Weight (g):</b>	104.9g
<b>Remainder Weight (g):</b>	131.5g

### Notes


## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

<b>Sampling Location (select):</b>	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
<b>GPS coordinate (UTM-WGS84):</b>	
<b>Date &amp; Time (24 hr)</b>	Date: <u>08/03/12</u> Time: <u>1335</u>
<b>Weather/ Temperature:</b>	
<b>Tackle/Harvest Method (select):</b>	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <del>Cast</del> Fly
<b>Sample Collected By:</b>	<u>Voss</u>

### Morphometric Data

<b>Otolith Vial #:</b>	<u>CR-AG-06 - OTOLITH</u>
<b>Scale Sample ID:</b>	<u>CR-AG-06 - SCALE</u>
<b>Fresh Field Weight (kg):</b>	<u>339.3 g</u>
<b>Length (cm):</b>	<u>33.5</u>
<b>Sex (select):</b>	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

<b>Fish Tissue Sample ID (fillet(s)):</b>	<u>CR-AG-06 - FILLET</u>
<b>Fish Tissue Sample ID (remainder):</b>	<u>CR-AG-06 - WHOLE</u>
<b>Number of Fillets:</b>	<u>1</u>
<b>Fillet Weight (kg):</b>	<u>75.2g</u>
<b>Remainder Weight (kg):</b>	<u>264.1g</u>

### Notes


# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr) Sample Collected	Date: 08/04/12                      Time: 1344
Weather/ Temperature:	cloudy, wind, 50 <sup>°</sup>
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	CONNOR

## Morphometric Data

Otolith Vial #:	CR-AG-07-OTOLITH
Scale Sample ID:	CR-AG-07-SCALE
Fresh Field Weight (kg):	230.9 g
Length (cm):	29.0
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CR-AG-07-FILLET
Fish Tissue Sample ID (remainder):	CR-AG-07-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	<del>103.5</del> 103.5 g
Remainder Weight (kg):	<del>127.4</del> 127.4 g

## Notes


Date Sample Shipped for Analysis:

ADF&G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760



## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr) Sample Collected	Date: 08/03/12      Time: 1450
Weather/ Temperature:	cloudy, wind, 50's
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	Conroy

## Morphometric Data

Otolith Vial #:	CR-AG-09-OTOLITH
Scale Sample ID:	CR-AG-09-SCALE
Fresh Field Weight (kg):	390.3g
Length (cm):	36.5
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CR-AG-09-FILLET      CR-AG-21-FILLET
Fish Tissue Sample ID (remainder):	CR-AG-09-WHOLE
Number of Fillets:	(2) 1) 72.1g      2) 80.9g
Fillet Weight (kg):	(1) 72.1g = CR-AG-09-FILLET      (2) 80.9g = CR-AG-21-FILLET
Remainder Weight (kg):	237.3g

## Notes

CR-AG-09 - PROVIDES THE FILLET DUPLICATES  
 CR-AG-21-FILLET - MUST BE ADDED IN TO  
 REMAINING WHOLE BODY CONCENTRATION.

Date Sample Shipped for Analysis:

ADF&G Area Biologist: Audra Brase-(907) 459-7244

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EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr) Sample Collected	Date: 08/03/12                      Time: 1450
Weather/ Temperature:	cc0007, windy, 50's
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	Conroy

## Morphometric Data

Otolith Vial #:	NA
Scale Sample ID:	NA
Fresh Field Weight (kg):	390.3g
Length (cm):	36.5
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CR-AG-21-FILLET
Fish Tissue Sample ID (remainder):	CR-AG-09-WHOLE
Number of Fillets:	2 - 80.9g + 72.1g (CR-AG-09-fillet)
Fillet Weight (kg):	(80.9g + 72.1g) = 153g
Remainder Weight (kg):	237.3g

## Notes

- DUPLICATE FOR CR-AG-09-FILLET

Date Sample Shipped for Analysis:



# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr) Sample Collected	Date: 08/03/12                      Time: 1515
Weather/ Temperature:	cloudy, wind, 50's
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	CONNOR

## Morphometric Data

Otolith Vial #:	CR-AG-11-OTOLITH
Scale Sample ID:	CR-AG-11-SCALE
Fresh Field Weight (kg):	257.8g
Length (cm):	33.0
Sex (select):	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CR-AG-11-FILLET
Fish Tissue Sample ID (remainder):	CR-AG-11-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	104.9g
Remainder Weight (kg):	152.9g

## Notes


Date Sample Shipped for Analysis:



# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr) Sample Collected	Date: 08/03/12                      Time: 1540
Weather/ Temperature:	Cloudy, Wind, 50°
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	Voss

## Morphometric Data

Otolith Vial #:	CR-A6-12-OTOLITH
Scale Sample ID:	CR-A6-12-SCALE
Fresh Field Weight (kg):	293.2g
Length (cm):	32.5
Sex (select):	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CR-A6-12-FILLET
Fish Tissue Sample ID (remainder):	CR-A6-12-REMAINDER
Number of Fillets:	1
Fillet Weight (kg):	70.4g
Remainder Weight (kg):	222.8g

## Notes


Date Sample Shipped for Analysis:

## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr) Sample Collected	Date: 06/ August / 2012      Time: 0920
Weather/ Temperature:	Sunny 60°, light breeze
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input checked="" type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	C. Schultz

### Morphometric Data

Otolith Vial #:	CR-AG-13-otolith
Scale Sample ID:	CR-AG-13-scale
Fresh Field Weight (kg):	333.5
Length (cm):	36.0
Sex (select):	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CR-AG-13-Fillet
Fish Tissue Sample ID (remainder):	CR-AG-13-whole
Number of Fillets:	1
Fillet Weight (kg):	72.3
Remainder Weight (kg):	261.2

### Notes


Date Sample Shipped for Analysis:

ADF&G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr) Sample Collected	Date: 06/August/2012                      Time: 0940
Weather/ Temperature:	Sunny 65°F, light breeze
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input checked="" type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	C. Schultz

### Morphometric Data

Otolith Vial #:	CR-AG-14-otolith
Scale Sample ID:	CR-AG-14-scale
Fresh Field Weight (kg):	326.3
Length (cm):	34.0
Sex (select):	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CR-AG-14-Fillet
Fish Tissue Sample ID (remainder):	CR-AG-14-whole
Number of Fillets:	1
Fillet Weight (kg):	77.5
Remainder Weight (kg):	248.8

### Notes


Date Sample Shipped for Analysis:

ADF&G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr) Sample Collected	Date: 06/August/2012                      Time: 1014
Weather/ Temperature:	Sunny 65°F, light breeze
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input checked="" type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	C. Schultz

### Morphometric Data

Otolith Vial #:	CR-AG-15-Otolith
Scale Sample ID:	CR-AG-15-Scale
Fresh Field Weight (kg):	242.6
Length (cm):	31.0
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CR-AG-15-Fillet
Fish Tissue Sample ID (remainder):	CR-AG-15-Whole
Number of Fillets:	2
Fillet Weight (kg):	92.9
Remainder Weight (kg):	149.7

### Notes


Date Sample Shipped for Analysis:

ADF&G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr) Sample Collected	Date: 06 August / 2012      Time: 1040
Weather/ Temperature:	SUNNY 65°F, breeze
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input checked="" type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	S. VOSS

## Morphometric Data

Otolith Vial #:	CR-AG-16 - Otolith
Scale Sample ID:	CR-AG-16 - scale
Fresh Field Weight (kg):	257.2
Length (cm):	31.5
Sex (select):	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CR-AG-16 - Fillet
Fish Tissue Sample ID (remainder):	<del>CR-AG-16</del> EBT CR-AG-16 - whole
Number of Fillets:	2
Fillet Weight (kg):	106.2
Remainder Weight (kg):	151.0

## Notes


Date Sample Shipped for Analysis:

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr) Sample Collected	Date: 06/August/2012                      Time: 1140
Weather/ Temperature:	70°F Sunny
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input checked="" type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	C. Schultz

## Morphometric Data

Otolith Vial #:	CR-AG-17-otolith
Scale Sample ID:	CR-AG-17-scale
Fresh Field Weight (kg):	254.2
Length (cm):	31.0
Sex (select):	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CR-AG-17-Fillet
Fish Tissue Sample ID (remainder):	CR-AG-17-whole
Number of Fillets:	2
Fillet Weight (kg):	110.6
Remainder Weight (kg):	143.6

## Notes

Date Sample Shipped for Analysis:

ADF&G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

<b>Sampling Location (select):</b>	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
<b>GPS coordinate (UTM-WGS84):</b>	
<b>Date &amp; Time (24 hr) Sample Collected</b>	Date: <i>06 August</i> Time: <i>1155</i>
<b>Weather/ Temperature:</b>	<i>70°F, Sunny</i>
<b>Tackle/Harvest Method (select):</b>	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input checked="" type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
<b>Sample Collected By:</b>	<i>S. Voss</i>

### Morphometric Data

<b>Otolith Vial #:</b>	<i>CR-AG-18-otolith</i>
<b>Scale Sample ID:</b>	<i>CR-AG-18-scale</i>
<b>Fresh Field Weight (kg):</b>	<i>255.6</i>
<b>Length (cm):</b>	<i>31.5</i>
<b>Sex (select):</b>	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

<b>Fish Tissue Sample ID (fillet(s)):</b>	<i>CR-AG-18-Fillet</i>
<b>Fish Tissue Sample ID (remainder):</b>	<i>CR-AG-18-whole</i>
<b>Number of Fillets:</b>	<i>2</i>
<b>Fillet Weight (kg):</b>	<i>82.4</i>
<b>Remainder Weight (kg):</b>	<i>173.2</i>

### Notes


<b>Date Sample Shipped for Analysis:</b>
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ADF&G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr) Sample Collected	Date: 06/August/2012      Time: 1210
Weather/ Temperature:	Sunny, 70°F.
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input checked="" type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	C. Schultz

## Morphometric Data

Otolith Vial #:	CR-AG-19 - otolith
Scale Sample ID:	CR-AG-19 - scale
Fresh Field Weight (kg):	306.3
Length (cm):	33.0
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CR-AG-19 - FILLET
Fish Tissue Sample ID (remainder):	CR-AG-19 - whole
Number of Fillets:	1
Fillet Weight (kg):	74.1
Remainder Weight (kg):	232.2

## Notes

Date Sample Shipped for Analysis:

ADF&G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760



# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input checked="" type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr) Sample Collected	Date: 06 / August / 2012      Time: 1215
Weather/ Temperature:	SUNNY, 75°F, light breeze.
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input checked="" type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	S. VOSS

## Morphometric Data

Otolith Vial #:	CR-AG-20-otolith
Scale Sample ID:	CR-AG-20-scale
Fresh Field Weight (kg):	178.7
Length (cm):	27.5
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	CR-AG-20-Fillet
Fish Tissue Sample ID (remainder):	CR-AG-20-whole
Number of Fillets:	2
Fillet Weight (kg):	70.3
Remainder Weight (kg):	108.4

## Notes

Date Sample Shipped for Analysis:

ADF&G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760



## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

<b>Sampling Location (select):</b>	<input checked="" type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River		
<b>GPS coordinate (UTM-WGS84):</b>	(At Culvert below Fish Gate) DU 17		
<b>Date &amp; Time (24 hr)</b>	Date: 7/31/12	Time: 1250	
<b>Weather/ Temperature:</b>	cloud 70°		
<b>Tackle/Harvest Method (select):</b>	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input checked="" type="checkbox"/> Rod & Reel : <input checked="" type="checkbox"/> Spin <input type="checkbox"/> Fly		
<b>Sample Collected By:</b>	Rich Connelly		

### Morphometric Data

<b>Otolith Vial #:</b>	GS-AG-01-OTOLITH		
<b>Scale Sample ID:</b>	GS-AG-01-SCALE		
<b>Fresh Field Weight (kg):</b> grams	255.8 grams		
<b>Length (cm):</b>	30 cm		
<b>Sex (select):</b>	<input checked="" type="checkbox"/> Male	<input type="checkbox"/> Female	<input type="checkbox"/> Unknown

### Sample Processing Information

<b>Fish Tissue Sample ID (fillet(s)):</b>	GS-AG-01-FILLET		
<b>Fish Tissue Sample ID (remainder):</b>	GS-AG-01-WHOLE		
<b>Number of Fillets:</b>	1		
<b>Fillet Weight (kg):</b> grams	67.5 grams		
<b>Remainder Weight (kg):</b>	188.3		

### Notes

DU 17

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ADF&G Area Biologist: Audra Base-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

<b>Sampling Location (select):</b>	<input checked="" type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River		
<b>GPS coordinate (UTM-WGS84):</b>	Road culvert of DU 18		
<b>Date &amp; Time (24 hr)</b>	Date: 7/31/12	Time: 1345	
<b>Weather/ Temperature:</b>	75° Sunny		
<b>Tackle/Harvest Method (select):</b>	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input checked="" type="checkbox"/> Rod & Reel : <input checked="" type="checkbox"/> Spin <input type="checkbox"/> Fly		
<b>Sample Collected By:</b>	Chris Schultz (CS)		

### Morphometric Data

<b>Otolith Vial #:</b>	<del>GS-AG-02-OTOLITH</del> GS-AG-02-OTOLITH		
<b>Scale Sample ID:</b>	GS-AG-02-OTOLITH SCALE		
<b>Fresh Field Weight (kg):</b>	204.5g		
<b>Length (cm):</b>	28.5		
<b>Sex (select):</b>	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown		

### Sample Processing Information

<b>Fish Tissue Sample ID (fillet(s)):</b>	GS-AG-02-FILLET		
<b>Fish Tissue Sample ID (remainder):</b>	GS-AG-02-WHOLE		
<b>Number of Fillets:</b>	2		
<b>Fillet Weight (kg):</b>	87.6 grams		
<b>Remainder Weight (kg):</b>	116.9		

### Notes

DU 18	From road/culvert		
	6502		

ADF&G Area Biologist: Audra Base-(907) 459-7244

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EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input checked="" type="checkbox"/> Garrison Slough	<input type="checkbox"/> Piledriver Slough	<input type="checkbox"/> Chena River	<input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	(Road culvert DU 18)			
Date & Time (24 hr)	Date: 7/31/12	Time: 1405		
Weather/ Temperature:	75 Sunny			
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing	<input type="checkbox"/> Backpack- Electrofishing	<input checked="" type="checkbox"/> Rod & Reel	<input checked="" type="checkbox"/> Spin <input type="checkbox"/> Fly
Sample Collected By:	Chris Schultz			

## Morphometric Data

Otolith Vial #:	GS-AG-03-OTOLITH
Scale Sample ID:	GS-AG-03-SCALE
Fresh Field Weight (kg):	339.3 grams
Length (cm):	33 cm
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	GS-AG-03-FILLET
Fish Tissue Sample ID (remainder):	GS-AG-03-WHOLE
Number of Fillets:	
Fillet Weight (kg):	970.5
Remainder Weight (kg):	268.8 grams

## Notes

DU 18  
 GS03

ADF&G Area Biologist: Audra Base-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

<b>Sampling Location (select):</b>	<input checked="" type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
<b>GPS coordinate (UTM-WGS84):</b>	
<b>Date &amp; Time (24 hr)</b>	Date: 7/31/2012                      Time: 1545
<b>Weather/ Temperature:</b>	75 Sunny
<b>Tackle/Harvest Method (select):</b>	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input checked="" type="checkbox"/> Spin <input type="checkbox"/> Fly
<b>Sample Collected By:</b>	JOE NEUBAUER

### Morphometric Data

<b>Otolith Vial #:</b>	GS-AG-04-OTOLITH
<b>Scale Sample ID:</b>	GS-AG-04-SCALE
<b>Fresh Field Weight (kg):</b>	276.4 grams
<b>Length (cm):</b>	33cm
<b>Sex (select):</b>	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

<b>Fish Tissue Sample ID (fillet(s)):</b>	GS-AG-04-FILLET
<b>Fish Tissue Sample ID (remainder):</b>	GS-AG-04-WHOLE
<b>Number of Fillets:</b>	1
<b>Fillet Weight (kg):</b>	71.5 grams
<b>Remainder Weight (kg):</b>	204.9

### Notes

- GS-04
- DU 17
33cm

ADF&G Area Biologist: Audra Base-(907) 459-7244

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Towing Service: A-1 Denali Towing- (907) 388-6760



# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input checked="" type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	(Road @ Fish Gate)
Date & Time (24 hr)	Date: 7/31/2012    Time: 1550
Weather/ Temperature:	75 Sunny
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input checked="" type="checkbox"/> Spin <input type="checkbox"/> Fly
Sample Collected By:	RICH CONNOR

## Morphometric Data

Otolith Vial #:	GS-AG-05-OTOLITH
Scale Sample ID:	GS-AG-05-SCALE
Fresh Field Weight (kg):	1.91
Length (cm):	28 cm
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	GS-AG-05-FILLET
Fish Tissue Sample ID (remainder):	GS-AG-05-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	92.1 grams
Remainder Weight (kg):	<del>88.9</del> 98.9

## Notes

- GS-05
- DU 17
- 28 cm

ADF&G Area Biologist: Audra Base-(907) 459-7244

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Towing Service: A-1 Denali Towing- (907) 388-6760



# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input checked="" type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	(DU 18 Round at Culvert)
Date & Time (24 hr)	Date: 7/31/12    Time: 1600
Weather/ Temperature:	
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input checked="" type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input type="checkbox"/> Fly
Sample Collected By:	Rich Connelly / Sam Voss

## Morphometric Data

Otolith Vial #:	GS-AG-06-OTOLITH
Scale Sample ID:	GS-AG-06-SCALE
Fresh Field Weight (kg):	312.7
Length (cm):	33.5
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	GS-AG-06-FILLET
Fish Tissue Sample ID (remainder):	GS-AG-06-WHOLE
Number of Fillets:	1
Fillet Weight (kg):	70 grams
Remainder Weight (kg):	242.7

## Notes

GS-06  
DU 18  
33.5 cm

ADF&G Area Biologist: Audra Base-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760



Field Sampling Form 2012 - Fish Tissue & Otolith Collection

Harvest & Sample Location Information

Sampling Location (select):	<input checked="" type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: 08/02/12                      Time: 0829
Weather/ Temperature:	Part Cloudy 70's
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel: <input type="checkbox"/> Spin <input type="checkbox"/> Fly
Sample Collected By:	SCHULTE

Morphometric Data

Otolith Vial #:	GS-AG-07-OTOLITH
Scale Sample ID:	GS-AG-07-SCALE
Fresh Field Weight (kg):	182.9g
Length (cm):	28.0cm
Sex (select):	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	GS-AG-07-FILLET
Fish Tissue Sample ID (remainder):	GS-AG-07-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	80.1g
Remainder Weight (kg):	102.8g

Notes






# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input checked="" type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: 08/02/12                      Time: 0830
Weather/ Temperature:	Partly Cloudy 70's
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input type="checkbox"/> Fly
Sample Collected By:	Conroy

## Morphometric Data

Otolith Vial #:	GS-AG-08-OTOLITH
Scale Sample ID:	GS-AG-08-SCALE
Fresh Field Weight (kg):	213.1 g
Length (cm):	29.5
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	GS-AG-08-FILLET
Fish Tissue Sample ID (remainder):	GS-AG-08-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	103.0
Remainder Weight (kg):	110.1

## Notes


ADF&G Area Biologist: Audra Base-(907) 459-7244  
Eielson NRM: Ron Gunderson- (907) 377-5182  
EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297  
Towing Service: A-1 Denali Towing- (907) 388-6760

## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

Sampling Location (select):	<input checked="" type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: 08/02/12      Time: 0840
Weather/ Temperature:	Partly Cloudy 70's
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	SCHULTZ

### Morphometric Data

Otolith Vial #:	GS-AG-09-OTOLITH
Scale Sample ID:	GS-AG-09-SCALE
Fresh Field Weight (kg):	268.5g
Length (cm):	30.5
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	GS-AG-09-FILLET / GS-AG-21-FILLET <sup>(DUP)</sup>
Fish Tissue Sample ID (remainder):	GS-AG-09-WHOLE
Number of Fillets:	1st <sup>①</sup> 2nd FILLET IS DUP GS-AG-21-FILLET
Fillet Weight (kg):	1st 71.8g      2nd 72.8g
Remainder Weight (kg):	196.7 AFTER 1st FILLET

### Notes


ADF&G Area Biologist: Audra Base-(907) 459-7244

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EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

Sampling Location (select):	<input checked="" type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River		
GPS coordinate (UTM-WGS84):			
Date & Time (24 hr)	Date: 08/02/12	Time: 042	
Weather/ Temperature:	Partly Cloudy 70's		
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input type="checkbox"/> Fly		
Sample Collected By:	SCHULTZ		

### Morphometric Data

Otolith Vial #:	GS-AG-10-OTOLITH		
Scale Sample ID:	GS-AG-10-SCALE		
Fresh Field Weight (kg):	352.5		
Length (cm):	35.0		
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown		

### Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	GS-AG-10-FILLET (MS/MSD)		
Fish Tissue Sample ID (remainder):	GS-AG-10-WHOLE (MS/MSD)		
Number of Fillets:	2		
Fillet Weight (kg):	<del>71.8</del> <sup>RD</sup> 179.4g		
Remainder Weight (kg):	<del>179.4g</del> <sup>RD</sup> 173.1g		

### Notes

WILL BE USED AS MS/MSD + SAMPLE GS-AG-10

ADF&G Area Biologist: Audra Base-(907) 459-7244

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Towing Service: A-1 Denali Towing- (907) 388-6760

# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input checked="" type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: 08/02/12                      Time: 843
Weather/ Temperature:	PARTLY CLOUDY 70's
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input type="checkbox"/> Fly
Sample Collected By:	CONNORS

## Morphometric Data

Otolith Vial #:	GS-AG-11-OTOLITH
Scale Sample ID:	GS-AG-11-SCALE
Fresh Field Weight (kg):	235.6g
Length (cm):	30.0cm
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	GS-AG-11-FILLET
Fish Tissue Sample ID (remainder):	GS-AG-11-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	108.0
Remainder Weight (kg):	127.6

## Notes

~~CONNORS~~ ~~CONNORS~~ ~~CONNORS~~ (circled)

ADF&G Area Biologist: Audra Base-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

<b>Sampling Location (select):</b>	<input checked="" type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
<b>GPS coordinate (UTM-WGS84):</b>	
<b>Date &amp; Time (24 hr)</b>	Date: 08/02/12                      Time: 855
<b>Weather/ Temperature:</b>	Partly Cloudy 70's
<b>Tackle/Harvest Method (select):</b>	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
<b>Sample Collected By:</b>	SCHULTZ

### Morphometric Data

<b>Otolith Vial #:</b>	GS-AG-12-OTOLITH
<b>Scale Sample ID:</b>	GS-AG-12-SCALE
<b>Fresh Field Weight (kg):</b>	295.1g
<b>Length (cm):</b>	32.0
<b>Sex (select):</b>	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

<b>Fish Tissue Sample ID (fillet(s)):</b>	GS-AG-12-FILLET
<b>Fish Tissue Sample ID (remainder):</b>	GS-AG-12-WHOLE
<b>Number of Fillets:</b>	1
<b>Fillet Weight (kg):</b>	73.5g
<b>Remainder Weight (kg):</b>	221.6g

### Notes

- INJURY ALONG LEFT SIDE APPX 1cm MARK, POSSIBLY FROM BIRD (COMMONLY OBSERVED)

ADF&G Area Biologist: Audra Base-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760



## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

<b>Sampling Location (select):</b>	<input checked="" type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
<b>GPS coordinate (UTM-WGS84):</b>	
<b>Date &amp; Time (24 hr)</b>	Date: <u>08/02/12</u> Time:
<b>Weather/ Temperature:</b>	<u>Partly Cloudy 70's</u>
<b>Tackle/Harvest Method (select):</b>	<input type="checkbox"/> Boat- Electrofishing <input checked="" type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input type="checkbox"/> Fly
<b>Sample Collected By:</b>	<u>Electrosysteme Company, Voss</u>

### Morphometric Data

<b>Otolith Vial #:</b>	<u>GS-AG-14-OTOLITH</u>
<b>Scale Sample ID:</b>	<u>GS-AG-14-SCALE</u>
<b>Fresh Field Weight (kg):</b>	<u>204.9g</u>
<b>Length (cm):</b>	<u>27.5</u>
<b>Sex (select):</b>	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

<b>Fish Tissue Sample ID (fillet(s)):</b>	<u>GS-AG-14-FILLET</u>
<b>Fish Tissue Sample ID (remainder):</b>	<u>GS-AG-14-WHOLE</u>
<b>Number of Fillets:</b>	<u>2</u>
<b>Fillet Weight (kg):</b>	<u>78.0</u>
<b>Remainder Weight (kg):</b>	

### Notes


## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

Sampling Location (select):	<input checked="" type="checkbox"/> Garrison Slough	<input type="checkbox"/> Piledriver Slough	<input type="checkbox"/> Chena River	<input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):				
Date & Time (24 hr)	Date: 08/02/12	Time: 840		
Weather/ Temperature:	Partly Cloudy			
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing	<input type="checkbox"/> Backpack- Electrofishing	<input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly	
Sample Collected By:	SCHULTZ			

### Morphometric Data

Otolith Vial #:	NA
Scale Sample ID:	NA
Fresh Field Weight (kg):	268.5g
Length (cm):	30.5
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	GS-AG-21-FILLET
Fish Tissue Sample ID (remainder):	GS-AG-09-WHOLE
Number of Fillets:	2nd FILLET - 72.8g - 1 FILLET IN SAMPLER
Fillet Weight (kg):	72.8g
Remainder Weight (kg):	$268.5g - (71.8g + 72.8g) = 123.9g$

### Notes

- GS-AG-21-FILLET WILL BECOME AS DUPLICATE AND IS SECOND FILLET OF GS-AG-09.

ADF&G Area Biologist: Audra Base-(907) 459-7244

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EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760



## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

Sampling Location (select):	<input checked="" type="checkbox"/> Harrison Slough <input checked="" type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input checked="" type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: 08/02 Time: 9:55
Weather/ Temperature:	Partly Cloudy 40°
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input checked="" type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input type="checkbox"/> Fly
Sample Collected By:	Connelly Voss

### Morphometric Data

Otolith Vial #:	GS-AG-15 OTOLITH
Scale Sample ID:	GS-AG-15-SCALE
Fresh Field Weight (g):	287.6g
Length (cm):	31.0
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	GS-AG-15-FILLET
Fish Tissue Sample ID (remainder):	GS-AG-15-WHOLE
Number of Fillets:	1
Fillet Weight (kg):	79.7 g
Remainder Weight (kg):	207.9 g

### Notes


ADF&G Area Biologist: Audra Base-(907) 459-7244

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Towing Service: A-1 Denali Towing- (907) 388-6760





EA Engineering,  
Science, and  
Technology, Inc.

### Field Sampling Form 2012 - Fish Tissue & Otolith Collection

#### Harvest & Sample Location Information

Sampling Location (select):	<input checked="" type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr) Sample Collected	Date: <i>8/5/12</i> Time: <i>1426</i>
Weather/ Temperature:	
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input checked="" type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	<i>Chris Schultz</i>

#### Morphometric Data

Otolith Vial #:	<i>GS-AG-17-OTOLITH</i>
Scale Sample ID:	<i>GS-AG-17-scale</i>
Fresh Field Weight (kg):	<i>175.5</i>
Length (cm):	<i>27.5</i>
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

#### Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	<i>GS-AG-17-Fillet</i>
Fish Tissue Sample ID (remainder):	<i>GS-AG-17-whole</i>
Number of Fillets:	<i>2</i>
Fillet Weight (kg):	<i>77.5</i>
Remainder Weight (kg):	

#### Notes

*DU 17 below fish gate*

Date Sample Shipped for Analysis:

ADF&G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

**Field Sampling Form 2012 - Fish Tissue & Otolith Collection****Harvest & Sample Location Information**

Sampling Location (select):	<input checked="" type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr) Sample Collected	Date: 8/5/12                      Time: 1436
Weather/ Temperature:	
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input checked="" type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	Evana Burt-Toland

**Morphometric Data**

Otolith Vial #:	GS-AG-18-OTOLITH
Scale Sample ID:	GS-AG-18-scale
Fresh Field Weight (kg):	192.1
Length (cm):	29.0
Sex (select):	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

**Sample Processing Information**

Fish Tissue Sample ID (fillet(s)):	GS-AG-18-Fillet
Fish Tissue Sample ID (remainder):	GS-AG-18-whole
Number of Fillets:	2
Fillet Weight (kg):	80.2
Remainder Weight (kg):	

**Notes**

Date Sample Shipped for Analysis:

ADF&amp;G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

<b>Sampling Location (select):</b>	<input checked="" type="checkbox"/> Garrison Slough <input type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
<b>GPS coordinate (UTM-WGS84):</b>	
<b>Date &amp; Time (24 hr) Sample Collected</b>	Date: 8/5/12    Time: 1500
<b>Weather/ Temperature:</b>	
<b>Tackle/Harvest Method (select):</b>	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input checked="" type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
<b>Sample Collected By:</b>	Evana Burt-Toland

### Morphometric Data

<b>Otolith Vial #:</b>	GS-AG-19-OTOLITH
<b>Scale Sample ID:</b>	GS-AG-19-scale
<b>Fresh Field Weight (kg):</b>	244.6
<b>Length (cm):</b>	30.5
<b>Sex (select):</b>	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

<b>Fish Tissue Sample ID (fillet(s)):</b>	GS-AG-19-Fillet
<b>Fish Tissue Sample ID (remainder):</b>	GS-AG-19-whole
<b>Number of Fillets:</b>	2
<b>Fillet Weight (kg):</b>	93.8
<b>Remainder Weight (kg):</b>	

### Notes

DU17 below fish gate

**Date Sample Shipped for Analysis:**

**Field Sampling Form 2012 - Fish Tissue & Otolith Collection****Harvest & Sample Location Information**Sampling Location (select):  Garrison Slough     Piledriver Slough     Chena River     Chatanika River

GPS coordinate (UTM-WGS84):

Date &amp; Time (24 hr) Sample Collected

Date: 8/5/12

Time: 1520

Weather/ Temperature:

70 Sunny

Tackle/Harvest Method (select):

 Boat- Electrofishing Backpack- Electrofishing Dip net Rod & Reel :  Spin  Fly

Sample Collected By:

Chris Schultz

**Morphometric Data**

Otolith Vial #:

GS-AG-20-OTOLITH

Scale Sample ID:

GS-AG-20-SCALE

Fresh Field Weight (kg):

368.1g

Length (cm):

35.0

Sex (select):

 Male Female Unknown**Sample Processing Information**

Fish Tissue Sample ID (fillet(s)):

GS-AG-20-FILLET

Fish Tissue Sample ID (remainder):

GS-AG-20-WHOLE

Number of Fillets:

1

Fillet Weight (kg):

92.9

Remainder Weight (kg):

~~92.9~~**Notes**

DU 17 below fish gate

Date Sample Shipped for Analysis:

ADF&amp;G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

<b>Sampling Location (select):</b>	<input type="checkbox"/> Garrison Slough <input checked="" type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
<b>GPS coordinate (UTM-WGS84):</b>	
<b>Date &amp; Time (24 hr)</b>	Date: 08/02/12                      Time: 1240
<b>Weather/ Temperature:</b>	Partly Cloudy 70°
<b>Tackle/Harvest Method (select):</b>	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
<b>Sample Collected By:</b>	Voss

### Morphometric Data

<b>Otolith Vial #:</b>	PS-AG-01-OTOLITH
<b>Scale Sample ID:</b>	PS-AG-01-SCALE
<b>Fresh Field Weight (kg):</b>	361.1g
<b>Length (cm):</b>	36.0
<b>Sex (select):</b>	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

<b>Fish Tissue Sample ID (fillet(s)):</b>	PS-AG-01-FILLET - DUP PS-AG-21-FILLET
<b>Fish Tissue Sample ID (remainder):</b>	PS-AG-01-REMAINDER - <del>DUP PS-AG-21-FILLET</del>
<b>Number of Fillets:</b>	1st 1 FOR <del>PS-AG-01-FILLET</del> 2nd 95.3g
<b>Fillet Weight (kg):</b>	89.5g                      95.3g
<b>Remainder Weight (kg):</b>	= 361.1g - (89.5g + 95.3g) = 176.2

### Notes

<b>Notes:</b>	- PS-AG-01-FILLET - HAS 2ND FILLET AS DUPLICATE LABLED PS-AG-21-FILLET
<b>Notes:</b>	
<b>Notes:</b>	
<b>Notes:</b>	



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## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough	<input checked="" type="checkbox"/> Piledriver Slough	<input type="checkbox"/> Chena River	<input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):				
Date & Time (24 hr)	Date: 08/02/12	Time: 1245		
Weather/ Temperature:	Partly Cloudy 70°			
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing	<input type="checkbox"/> Backpack- Electrofishing		
	<input type="checkbox"/> Dip net	<input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly		
Sample Collected By:	Voss			

### Morphometric Data

Otolith Vial #:	PS-AG-02-OTOLITH
Scale Sample ID:	PS-AG-02-SCALE
Fresh Field Weight (kg):	164.3g
Length (cm):	28.5
Sex (select):	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	PS-AG-02-FILLET
Fish Tissue Sample ID (remainder):	PS-AG-02-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	73.0g
Remainder Weight (kg):	91.3g

### Notes


ADF&G Area Biologist: Audra Base-(907) 459-7244

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Towing Service: A-1 Denali Towing- (907) 388-6760





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## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough	<input checked="" type="checkbox"/> Piledriver Slough	<input type="checkbox"/> Chena River	<input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):				
Date & Time (24 hr)	Date: 08/03/12	Time: 1327		
Weather/ Temperature:	CLOUDY, WIND, 50'S			
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing	<input type="checkbox"/> Backpack- Electrofishing		
	<input type="checkbox"/> Dip net	<input type="checkbox"/> Rod & Reel :	<input type="checkbox"/> Spin	<input checked="" type="checkbox"/> Fly
Sample Collected By:	SCHULTZ			

### Morphometric Data

Otolith Vial #:	PS-AG-03-OTOLITH		
Scale Sample ID:	PS-AG-03-SCALE		
Fresh Field Weight (kg):	201.5g		
Length (cm):	28.5		
Sex (select):	<input checked="" type="checkbox"/> Male	<input type="checkbox"/> Female	<input type="checkbox"/> Unknown

### Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	PS-AG-03-FILLET
Fish Tissue Sample ID (remainder):	PS-AG-03-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	78.4g
Remainder Weight (kg):	123.1g

### Notes

ADF&G Area Biologist: Audra Base-(907) 459-7244

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Towing Service: A-1 Denali Towing- (907) 388-6760

## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

<b>Sampling Location (select):</b>	<input type="checkbox"/> Garrison Slough <input checked="" type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
<b>GPS coordinate (UTM-WGS84):</b>	
<b>Date &amp; Time (24 hr)</b>	Date: 08/03/12                      Time: 1400
<b>Weather/ Temperature:</b>	CLOUDY, WIND, 50°
<b>Tackle/Harvest Method (select):</b>	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
<b>Sample Collected By:</b>	SCHULTZ

### Morphometric Data

<b>Otolith Vial #:</b>	PS-AG-04-OTOLITH
<b>Scale Sample ID:</b>	PS-AG-04-SCALE
<b>Fresh Field Weight (kg):</b>	226.4g
<b>Length (cm):</b>	140
<b>Sex (select):</b>	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

<b>Fish Tissue Sample ID (fillet(s)):</b>	PS-AG-04-FILLET
<b>Fish Tissue Sample ID (remainder):</b>	PS-AG-04-WHOLE
<b>Number of Fillets:</b>	2
<b>Fillet Weight (kg):</b>	97.5g
<b>Remainder Weight (kg):</b>	128.9g

### Notes


# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input checked="" type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: 08/03/12      Time: 1405
Weather/ Temperature:	CLOUDY, WIND, 50°
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	SCAVUTZ

## Morphometric Data

Otolith Vial #:	PS-AG-05-OTOLITH
Scale Sample ID:	PS-AG-05-SCALE
Fresh Field Weight (kg):	166.3g
Length (cm):	29.0
Sex (select):	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	PS-AG-05-FILLET
Fish Tissue Sample ID (remainder):	PS-AG-05-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	70.4g
Remainder Weight (kg):	95.9g

## Notes


ADF&G Area Biologist: Audra Base-(907) 459-7244

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Towing Service: A-1 Denali Towing- (907) 388-6760

# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input checked="" type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: 08/03/12 Time: 1420
Weather/ Temperature:	CLOUDY, WIND 50"
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	Schultz

## Morphometric Data

Otolith Vial #:	PS-AG-06-OTOLITH
Scale Sample ID:	PS-AG-06-SCALE
Fresh Field Weight (kg):	251.7
Length (cm):	32.5
Sex (select):	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	PS-AG-06-FILLET
Fish Tissue Sample ID (remainder):	PS-AG-06-WHOLE
Number of Fillets:	1
Fillet Weight (kg):	67.4g
Remainder Weight (kg):	184.3g

## Notes


ADF&amp;G Area Biologist: Audra Base-(907) 459-7244

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Towing Service: A-1 Denali Towing- (907) 388-6760

# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input checked="" type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: 08/04/12 <sup>03</sup> Time: 1733
Weather/ Temperature:	CLOUDY, WIND, 50°
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	SCHULTZ

## Morphometric Data

Otolith Vial #:	PS-AG-07-OTOLITH
Scale Sample ID:	PS-AG-07-SCALE
Fresh Field Weight (kg):	184.4g
Length (cm):	28.5
Sex (select):	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	PG-AG-07-FILLET
Fish Tissue Sample ID (remainder):	PS-AG-07-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	72.2g
Remainder Weight (kg):	112.2 g

## Notes


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Towing Service: A-1 Denali Towing- (907) 388-6760

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough	<input checked="" type="checkbox"/> Piledriver Slough	<input type="checkbox"/> Chena River	<input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):				
Date & Time (24 hr) Sample Collected	Date: 08/04/12		Time: 1017	
Weather/ Temperature:	CLOUDY, 60's			
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing	<input type="checkbox"/> Backpack- Electrofishing		
	<input type="checkbox"/> Dip net	<input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin	<input checked="" type="checkbox"/> Fly	
Sample Collected By:	CUNNOY			

## Morphometric Data

Otolith Vial #:	PS-AG-08-OTOLITH
Scale Sample ID:	PS-AG-08-SCALE
Fresh Field Weight (kg):	200.1 g
Length (cm):	30.5
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	PS-AG-08-FILLET
Fish Tissue Sample ID (remainder):	PS-AG-08-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	75.4 g
Remainder Weight (kg):	124.7 g

## Notes

Date Sample Shipped for Analysis:

ADF&G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

## Field Sampling Form 2012 - Fish Tissue & Otolith Collection

### Harvest & Sample Location Information

<b>Sampling Location (select):</b>	<input type="checkbox"/> Garrison Slough <input checked="" type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
<b>GPS coordinate (UTM-WGS84):</b>	
<b>Date &amp; Time (24 hr) Sample Collected</b>	Date: 09/04/2012                      Time: 1022
<b>Weather/ Temperature:</b>	CLOUDY, 60'S
<b>Tackle/Harvest Method (select):</b>	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
<b>Sample Collected By:</b>	VOSS

### Morphometric Data

<b>Otolith Vial #:</b>	PS-AG-09-OTOLITH
<b>Scale Sample ID:</b>	PS-AG-09-SCALE
<b>Fresh Field Weight (kg):</b>	276.5g
<b>Length (cm):</b>	33.5
<b>Sex (select):</b>	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

### Sample Processing Information

<b>Fish Tissue Sample ID (fillet(s)):</b>	PS-AG-09-FILLET MS/MSD
<b>Fish Tissue Sample ID (remainder):</b>	PS-AG-09-WHOLE MS/MSD
<b>Number of Fillets:</b>	2
<b>Fillet Weight (kg):</b>	117g
<b>Remainder Weight (kg):</b>	159.5g

### Notes

MS/MSD - SELECTED THE LARGEST
PONDING FISH FROM PILEDRIWER -

**Date Sample Shipped for Analysis:**

ADF&G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input checked="" type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr) Sample Collected	Date: 08/04/2012      Time: 1030
Weather/ Temperature:	Cloudy, 60's
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	SCHULTZ

## Morphometric Data

Otolith Vial #:	PS-AG-10-OTOLITH
Scale Sample ID:	PS-AG-10-SCALE
Fresh Field Weight (kg):	208.9g
Length (cm):	28.5
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	PS-AG-10-FILLET
Fish Tissue Sample ID (remainder):	PS-AG-10-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	94.5g
Remainder Weight (kg):	114.4g

## Notes


Date Sample Shipped for Analysis:



## Field Sampling Form 2012 - Fish Tissue &amp; Otolith Collection

## Harvest &amp; Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough	<input checked="" type="checkbox"/> Piledriver Slough	<input type="checkbox"/> Chena River	<input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):				
Date & Time (24 hr) Sample Collected	Date: 08/04/2012	Time: 1040		
Weather/ Temperature:	Cloudy 60's			
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing	<input type="checkbox"/> Backpack- Electrofishing	<input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin	<input checked="" type="checkbox"/> Fly
Sample Collected By:	COMBLY			

## Morphometric Data

Otolith Vial #:	PS-AG-11-OTOLITH		
Scale Sample ID:	PS-AG-11-SCALE		
Fresh Field Weight (kg):	238.3		
Length (cm):	32.0		
Sex (select):	<input type="checkbox"/> Male	<input checked="" type="checkbox"/> Female	<input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	PS-AG-11-FILLET
Fish Tissue Sample ID (remainder):	PS-AG-11-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	88.8g
Remainder Weight (kg):	149.5g

## Notes


Date Sample Shipped for Analysis:

ADF&amp;G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760



# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input checked="" type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr) Sample Collected	Date: 08/04/2012                      Time: 1151
Weather/ Temperature:	CLOUDY 60'S
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	SCHULTZ

## Morphometric Data

Otolith Vial #:	PS-AG-13-OTOLITH
Scale Sample ID:	PS-AG-13-SCALE
Fresh Field Weight (kg):	214.0
Length (cm):	29.0
Sex (select):	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	PS-AG-13-FILLET
Fish Tissue Sample ID (remainder):	PS-AG-13-WHOLE
Number of Fillets:	2
Fillet Weight (kg):	96.0g
Remainder Weight (kg):	118.0g

## Notes


Date Sample Shipped for Analysis:

## Field Sampling Form 2012 - Fish Tissue &amp; Otolith Collection

## Harvest &amp; Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input checked="" type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr) Sample Collected	Date: 8/7/12                      Time: 0930
Weather/ Temperature:	cloudy, rain / 52° F
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input checked="" type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	R. Connelly

## Morphometric Data

Otolith Vial #:	PS-A6-14-otolith
Scale Sample ID:	PS-A6-14-scale
Fresh Field Weight (kg):	167.9 g
Length (cm):	27.0
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	PS-A6-14-Fillet
Fish Tissue Sample ID (remainder):	PS-A6-14-whole
Number of Fillets:	2
Fillet Weight (kg):	74.3
Remainder Weight (kg):	43.6

## Notes


Date Sample Shipped for Analysis:

ADF&amp;G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

**Field Sampling Form 2012 - Fish Tissue & Otolith Collection**
**Harvest & Sample Location Information**

Sampling Location (select):	<input type="checkbox"/> Garrison Slough	<input checked="" type="checkbox"/> Piledriver Slough	<input type="checkbox"/> Chena River	<input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):				
Date & Time (24 hr) Sample Collected	Date: 8/7/12	Time: 1035		
Weather/ Temperature:	Cloudy, rain / 52°F			
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing	<input type="checkbox"/> Backpack- Electrofishing		
	<input type="checkbox"/> Dip net	<input checked="" type="checkbox"/> Rod & Reel	<input type="checkbox"/> Spin	<input checked="" type="checkbox"/> Fly
Sample Collected By:	S. Voss			

**Morphometric Data**

Otolith Vial #:	PS-A6-15-Otolith		
Scale Sample ID:	PS-A6-15-Scale		
Fresh Field Weight (kg):	218.9g		
Length (cm):	30.0		
Sex (select):	<input checked="" type="checkbox"/> Male	<input type="checkbox"/> Female	<input type="checkbox"/> Unknown

**Sample Processing Information**

Fish Tissue Sample ID (fillet(s)):	PS-A6-15-Fillet
Fish Tissue Sample ID (remainder):	PS-A6-15-Whole
Number of Fillets:	2
Fillet Weight (kg):	92.0
Remainder Weight (kg):	126.9

**Notes**

Date Sample Shipped for Analysis:

ADF&amp;G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760



# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input checked="" type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr) Sample Collected	Date: 8/8/12 Time: 1142
Weather/ Temperature:	
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input checked="" type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	E. Burt - Toland

## Morphometric Data

Otolith Vial #:	PS-AG-16 - OTOLITH
Scale Sample ID:	PS-AG-16 - Scale
Fresh Field Weight (kg):	164.7
Length (cm):	27.5
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	PS-AG-16 - FILLET
Fish Tissue Sample ID (remainder):	PS-AG-16 - WHOLE
Number of Fillets:	2
Fillet Weight (kg):	70.4
Remainder Weight (kg):	94.3

## Notes

Date Sample Shipped for Analysis:

ADF&G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760





# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough	<input checked="" type="checkbox"/> Piledriver Slough	<input type="checkbox"/> Chena River	<input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):				
Date & Time (24 hr) Sample Collected	Date: 8/8/12	Time: 1720		
Weather/ Temperature:				
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing	<input type="checkbox"/> Backpack- Electrofishing		
	<input type="checkbox"/> Dip net	<input checked="" type="checkbox"/> Rod & Reel	<input type="checkbox"/> Spin	<input checked="" type="checkbox"/> Fly
Sample Collected By:	C Schultz			

## Morphometric Data

Otolith Vial #:	PS-AG-18-OTOLITH
Scale Sample ID:	PS-AG-18-SCALE
Fresh Field Weight (kg):	206.7
Length (cm):	29.0
Sex (select):	<input type="checkbox"/> Male <input checked="" type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	PS-AG-18-FILLET
Fish Tissue Sample ID (remainder):	PS-AG-18-WHOLE
Number of Fillets:	
Fillet Weight (kg):	73.6
Remainder Weight (kg):	133.1

## Notes


Date Sample Shipped for Analysis:

ADF&G Area Biologist: Audra Brase-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760







# Field Sampling Form 2012 - Fish Tissue & Otolith Collection

## Harvest & Sample Location Information

Sampling Location (select):	<input type="checkbox"/> Garrison Slough <input checked="" type="checkbox"/> Piledriver Slough <input type="checkbox"/> Chena River <input type="checkbox"/> Chatanika River
GPS coordinate (UTM-WGS84):	
Date & Time (24 hr)	Date: 08/02/12 Time: 1240
Weather/ Temperature:	Partly Cloudy
Tackle/Harvest Method (select):	<input type="checkbox"/> Boat- Electrofishing <input type="checkbox"/> Backpack- Electrofishing <input type="checkbox"/> Dip net <input type="checkbox"/> Rod & Reel : <input type="checkbox"/> Spin <input checked="" type="checkbox"/> Fly
Sample Collected By:	Yoss

## Morphometric Data

Otolith Vial #:	NA
Scale Sample ID:	NA
Fresh Field Weight (kg):	361.1g
Length (cm):	36.0
Sex (select):	<input checked="" type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Unknown

## Sample Processing Information

Fish Tissue Sample ID (fillet(s)):	PS-AG-21-Fillet
Fish Tissue Sample ID (remainder):	PS-AG-01-WHOLE
Number of Fillets:	200 Fillets of PS-AG-01 (1st fillet = 89.5g)
Fillet Weight (kg):	95.3g
Remainder Weight (kg):	$= 361.1g - (95.3g + 89.5g) = 176.2g$

## Notes

PS-AG-21-Fillet is duplicate for PS-AG-01

ADF&G Area Biologist: Audra Base-(907) 459-7244

Eielson NRM: Ron Gunderson- (907) 377-5182

EA Contact Numbers: Fairbanks Office (907) 456-4751 and Kyle Waldron (907) 529-0297

Towing Service: A-1 Denali Towing- (907) 388-6760

**ATTACHMENT 4**  
**Photograph Log**

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**Photo No. 1** – 14 November 2012  
Typical condition of Garrison Slough. View facing northeast.



**Photo No. 2** – 14 November 2012  
Test sampling at drainage channel entrance to Garrison Slough. View facing northeast.

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**Photo No. 3 – 14 November 2012**  
Sample core retrieved using the single tube sampler. View facing northeast.



**Photo No. 4 – 14 November 2012**  
Test sample behind Bldg. 2230, no recovery. View facing east.

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**Photo No. 5** – 15 November 2012  
Penetrating ice for sub-sample acquisition from boat at DU-7-1. View facing west.



**Photo No. 6** – 15 November 2012  
Sub-sample acquisition using a fence post driver from boat at DU-7-2. View facing west.

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**Photo No. 7 – 14 November 2012**  
Establishing survey control adjacent to Garrison Slough DU-7-1. View facing southwest.



**ATTACHMENT 5**  
**Analytical Data and Data Quality Assessment**

## SS67 Site Investigation – Eielson Air Force Base

### Data Quality Assessment

#### INTRODUCTION

This Data Quality Assessment (DQA) was performed to assess the analytical data quality for the results described in this Supplemental Remedial Investigation at Source Area SS67 After-Action Report. Tissue sampling activities were conducted by EA Engineering, Science, and Technology Inc. in August 2012. TestAmerica Laboratories, Inc. (TestAmerica) of Pittsburg, Pennsylvania provided analytical support for tissue analysis. Sediment sampling was conducted by Jacobs Engineering Group Inc. (Jacobs) in November 2012. TestAmerica of Tacoma, Washington provided analytical support for sediment characterization.

This DQA contains the following exhibits:

- Exhibit A contains a sample summary table and the analytical data tables.
- Exhibit B contains tables of qualified data.
- Exhibit C includes the Alaska Department of Environmental Conservation (ADEC) Laboratory Data Review Checklists for each sample delivery group.
- Exhibit D contains the laboratory data packages in electronic format.

In addition to preparing this DQA, Jacobs performed a data quality review and completed the ADEC Laboratory Data Review Checklists (Exhibit C) for the records associated with the analytical data. The data review was performed in accordance with the data review and qualification criteria listed in Section 1.1 of the *Phase I Remedial Investigation Management Plan Source Area SS67 Work Plan* (United States Air Force [USAF] 2012a). Results were categorized as either “acceptable” or “estimated” using the data qualifier definitions defined in this DQA. A completeness check of the laboratory data was performed to verify that the data package and electronic files include all information requested.

#### Data Review and Qualification

All analytical data were reviewed by the Jacobs Project Chemist. This evaluation consisted of a review of chain-of-custody (CoC) and sample receipt records, laboratory case narratives, laboratory data (including analytical methodology), sample holding times, laboratory blanks, limits of detection (LOD), limits of quantitation (LOQ), surrogate recoveries, laboratory control sample (LCS) accuracy, and matrix spike (MS) accuracy and precision. Analytical results

## SS67 Site Investigation – Eielson Air Force Base

were evaluated against analytical methods (ADEC 2002; U.S. Environmental Protection Agency [EPA] 2007) and laboratory limits.

If a result or recovery fell outside the control limits, a qualifier code was applied to that datum. Qualification was not required in the following circumstances:

- Surrogate or MS recoveries were outside quality control (QC) limits, and the sample was diluted by a factor of five or greater.
- MS recoveries were outside QC limits, and the spiked concentration was less than twice that of the parent sample.
- An analyte was detected in the method blank, but there was no detection in the sample.
- MS or LCS recoveries exceeded upper control limits, and there was no detection in the sample(s).

Qualifiers applied to the analytical data set include the following:

- J The analyte was positively identified, but the associated result was less than the LOQ and greater than or equal to the LOD.
- B The analyte was detected in the method blank or the trip blank above the LOD, and the concentration in the sample did not exceed the blank concentration by a factor of five (factor of 10 for common laboratory contaminants acetone and methylene chloride).
- E The analytical result was nondetect and the LOQ is greater than the project action limit (PAL).
- JS- The result was an estimated value because at least one surrogate failed recovery criteria for that sample or at least two surrogates failed recovery criteria for the SW8270-SIM and SW-8260 methods.

Data may be rejected on the following grounds:

- Initial calibration (per compound) criteria not met
- Continuing calibration (per compound) not verified
- All nondetects with the continuing calibration recovery less than control limits
- Any compound with LCS recovery less than 10 percent
- Missed holding times greater than two times the method-specified holding time
- Surrogate recovery of less than 10 percent and a dilution factor of 5 or less
- Incorrect or inadequate preservation methods
- Cooler temperature reading greater than 12 degrees Celsius (°C)

Completeness is a quantitative evaluation indicating the percentage of the data that was considered usable (not rejected) for the intent of the project. The completeness goal is met

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when 95 percent of sample data is not rejected; 100 percent completeness was met for this project.

### **DATA QUALITY SUMMARY**

A review of the analytical results and associated QC samples found the overall quality of the project data to be acceptable. Complete details of the evaluation and associated samples are provided in the ADEC Laboratory Data Review Checklists (Exhibit C). All data was considered usable with the limitations discussed in this DQA and the ADEC laboratory data review checklists with regard to laboratory qualifiers defined above.

#### Sample Handling

Four coolers containing the fish samples were submitted to TestAmerica in Pittsburg. No significant issues were identified during review of the sample handling documentation. Cooler temperatures were recorded, but each temperature was not specified to the cooler from which it was measured. Three of the coolers were within  $4 \pm 2$  °C. One cooler temperature was measured at 1.9 °C. The sample results are unaffected by this shipping temperature. Upon arrival at the lab, the fish were stored in the freezer until analysis took place.

#### Holding Times

Samples significantly exceeded the holding times specified for each analysis. Due to budget constraints, EA Engineering was unable to have the samples analyzed after shipment to the laboratory in August 2013. Jacobs was contracted to continue the analysis and manage the data. In January 2013, the fish tissue samples were analyzed by the laboratory. All samples were more than 100 days past the specified holding times for pesticides and mercury. However, the usability of the tissue data should not be significantly affected as the samples had been kept frozen since the laboratory received them, and the analytes of interest (total mercury, pesticides, and polychlorinated biphenyls [PCB]) are non-volatile. Sample results were not qualified with JH to signal a hold time issue because the qualifier would affect all pesticide and mercury results.

#### Sample Matrix

With the exception of mercury and total lipids, all sample analyses required a minimum of a 5-fold dilution due to matrix interferences. There were multiple SW8082 and SW8081 MS and surrogate recoveries that were outside QC criteria due to dilution; however, as stated under Data Review and Qualification, qualification was not required if surrogate or MS were

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outside QC limits and the sample was diluted by a factor of five. These issues are discussed in the ADEC laboratory data review checklists (Exhibit C) and are most likely effects of matrix interference.

Aroclors 1254 and 1260 were reported in many samples. Although the Aroclor 1260 detections appeared to be of good quality, the majority of the 1254 detections were based on poor pattern matches and poor peak-to-peak ratio agreement with the reference standard. Jacobs asked the laboratory to review their Aroclor 1254 findings. Although the laboratory noted some of the same observations, they elected to confirm the original results.

### Reporting Limits

Laboratory LODs for nondetect sample results were evaluated against the PALs found in the *Quality Assurance Project Plan Remedial Investigation Source Area SS67 (USAF 2012c)*. Among 46 samples, 9 different analytes had LODs greater than the PAL. These results were qualified E and italicized in the analytical data tables (Exhibit A). The data was considered usable for the purpose of the project.

### Blanks

Method blank contamination was evaluated to the LOD and associated project samples within 5 times the method blank concentration were evaluated.

The method blanks for analytical batches 180-61868 and 180-62279 for methoxychlor had detections greater than the LOD. Associated project samples within 5 times the method blank concentration were qualified B. The impact is minimal since the sample results qualified are biased high but less than the PAL. Sample results qualified B can be found in Exhibit B.

### Surrogates

Several project sample surrogate recoveries were outside the limits specified in DoD Quality Systems Manual for Environmental Laboratories (QSM) (U.S. Department of Defense [DoD 2003]) or laboratory-established criteria, generally due to dilution. Surrogate recoveries were only evaluated if the sample was diluted by a factor of less than 5.

The SW8082 decachlorobiphenyl surrogate recovery for sample 12EAFB-SS67-SO-W01 was less than the control limits and is potentially biased low. All Aroclors for this sample were

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flagged JS-. Data quality is minimally affected because this sample was for waste characterization and Aroclor 1260 had a result of 19 milligrams per kilogram (mg/kg), which is significantly greater than the ADEC cleanup criteria of 1 mg/kg and significantly less than the Toxic Substances Control Act (TSCA) waste level of 50 mg/kg.

### CONCLUSION

In general, the overall quality of the project data was acceptable. Most Aroclor 1254 detections are not consistent with the reference standard and this analyte should be confirmed by another laboratory before inclusion as a site contaminant of concern. All data were considered usable for the purposes of this remedial investigation, with the limitations discussed in this DQA and the ADEC Laboratory Data Review Checklists (Exhibit C).

### REFERENCES

- ADEC (Alaska Department of Environmental Conservation). 2012 (April). Title 18 Alaska Administrative Code Section 75. *Oil and Other Hazardous Substances Pollution Control*.
- ADEC. 2002 (November). *Underground Storage Tanks Procedures Manual – Guidance for Remediation of Petroleum-Contaminated Soil and Water and Standard Sampling Procedures*.
- DoD (U.S. Department of Defense). 2003(June). *Quality Systems Manual for Environmental Laboratories*. Version 4.2.
- EPA (U.S. Environmental Protection Agency). 2007 (February). *Test Methods for Evaluating Solid Waste, Physical/Chemical Methods*. Final Update IV of the Third Edition. SW-846.
- USAF (U.S. Air Force) 2012a (December). *Phase I Remedial Investigation Management Plan, Source Area SS67, Eielson Air Force Base, Alaska*. Final. Prepared by Prepared by EA Engineering, Science, and Technology, Inc.
- USAF. 2012b (November). *Supplement to Quality Assurance Project Plan for Remedial Investigation Source Area SS67*. Final. Prepared by Jacobs Engineering Group Inc.
- USAF. 2012c (June). *Draft Final Quality Assurance Project Plan Remedial Investigation Source Area SS67*. Prepared by EA Engineering, Science, and Technology, Inc.

## SS67 Site Investigation – Eielson Air Force Base

### EXHIBITS

- Exhibit A Sample Summary Table & Analytical Results Tables
- Exhibit B Qualified Data Tables
- Exhibit C Alaska Department of Environmental Conservation Laboratory Data Review Checklists
- Exhibit D Laboratory Data Deliverables

**EXHIBIT A**  
**Sample Summary Table &**  
**Analytical Results Tables**



**Fish Collection Sample Summary**  
**Eielson AFB, Alaska**

Sample ID	Name of water body	Date	Fish collection method	Species	Life Stage	Length (mm)	Length method	Weight (g)	Sex	Age*	QC*	Aging method	Fish Tissue Analysis Results	Comments
GS-AG-01	Garrison Slough	7/31/2012	Angling	Arctic grayling	adult	300	total	256	Male	4		otolith	TBD	
GS-AG-02	Garrison Slough	7/31/2012	Angling	Arctic grayling	adult	285	total	205	Male	3		otolith	TBD	
GS-AG-03	Garrison Slough	7/31/2012	Angling	Arctic grayling	adult	330	total	339	Female	5 or 6**	5 or 6	otolith	TBD	Sample vial contained only one otolith – it was broken and unusable for age
GS-AG-04	Garrison Slough	7/31/2012	Angling	Arctic grayling	adult	330	total	276	Male	5 or 6**	5 or 6	scale	TBD	Otoliths were broken/shattered upon removal and discarded in field
GS-AG-05	Garrison Slough	7/31/2012	Angling	Arctic grayling	adult	280	total	191	Female	4		otolith	TBD	
GS-AG-06	Garrison Slough	7/31/2012	Backpack Electrofisher	Arctic grayling	adult	335	total	313	Female	5		otolith	TBD	
GS-AG-07	Garrison Slough	8/2/2012	Angling	Arctic grayling	adult	280	total	183	Male	4		otolith	TBD	
GS-AG-08	Garrison Slough	8/2/2012	Angling	Arctic grayling	adult	295	total	213	Female	4		otolith	TBD	
GS-AG-09	Garrison Slough	8/2/2012	Angling	Arctic grayling	adult	305	total	269	Female	4		otolith	TBD	
GS-AG-10	Garrison Slough	8/2/2012	Angling	Arctic grayling	adult	350	total	353	Female	5	5	otolith	TBD	
GS-AG-11	Garrison Slough	8/2/2012	Angling	Arctic grayling	adult	300	total	236	Female	5		otolith	TBD	
GS-AG-12	Garrison Slough	8/2/2012	Angling	Arctic grayling	adult	320	total	295	Female	6		otolith	TBD	
GS-AG-13	Garrison Slough	8/2/2012	Angling	Arctic grayling	adult	275	total	196	Male	4	4	otolith	TBD	
GS-AG-14	Garrison Slough	8/2/2012	Backpack Electrofisher	Arctic grayling	adult	275	total	205	Female	4		otolith	TBD	
GS-AG-15	Garrison Slough	8/2/2012	Backpack Electrofisher	Arctic grayling	adult	310	total	288	Female	5		otolith	TBD	
GS-AG-16	Garrison Slough	8/5/2012	Angling	Arctic grayling	adult	325	total	280	Female	5		otolith	TBD	
GS-AG-17	Garrison Slough	8/5/2012	Angling	Arctic grayling	adult	275	total	176	Female	3		otolith	TBD	
GS-AG-18	Garrison Slough	8/5/2012	Angling	Arctic grayling	adult	290	total	192	Male	4		otolith	TBD	
GS-AG-19	Garrison Slough	8/5/2012	Angling	Arctic grayling	adult	305	total	245	Female	5		otolith	TBD	
GS-AG-20	Garrison Slough	8/5/2012	Angling	Arctic grayling	adult	350	total	368	Male	6		otolith	TBD	

\* = Age determination conducted by Larry Bushing; Quality Control checks conducted by Ken Cummings

\*\* Age Estimated

\*\*\* Minimum Age Estimated

TBD= To Be Determined

**Fish Collection Sample Summary  
Eielson AFB, Alaska**

Sample ID	Name of water body	Date	Fish collection method	Species	Life Stage	Length (mm)	Length method	Weight (g)	Sex	Age*	QC*	Aging method	Fish Tissue Analysis Results	Comments
PS-AG-01	Piledriver Slough	8/2/2012	Angling	Arctic grayling	adult	360	total	361	Male	6		otolith	TBD	
PS-AG-02	Piledriver Slough	8/2/2012	Angling	Arctic grayling	adult	285	total	164	Male	4		otolith	TBD	
PS-AG-03	Piledriver Slough	8/3/2012	Angling	Arctic grayling	adult	285	total	202	Male	4		otolith	TBD	
PS-AG-04	Piledriver Slough	8/3/2012	Angling	Arctic grayling	adult	305	total	226	Male	5		otolith	TBD	
PS-AG-05	Piledriver Slough	8/3/2012	Angling	Arctic grayling	adult	290	total	166	Male	4		otolith	TBD	Otolith in poor condition, but was able to be aged
PS-AG-06	Piledriver Slough	8/3/2012	Angling	Arctic grayling	adult	325	total	252	Male	6		otolith	TBD	
PS-AG-07	Piledriver Slough	8/3/2012	Angling	Arctic grayling	adult	285	total	184	Male	5	5	otolith	TBD	
PS-AG-08	Piledriver Slough	8/4/2012	Angling	Arctic grayling	adult	305	total	200	Female	6 or 7	6 or 7	otolith	TBD	
PS-AG-09	Piledriver Slough	8/4/2012	Angling	Arctic grayling	adult	335	total	277	Male	6		otolith	TBD	
PS-AG-10	Piledriver Slough	8/4/2012	Angling	Arctic grayling	adult	285	total	209	Female	5	5	otolith	TBD	One of two otoliths was broken but both were found to be readable
PS-AG-11	Piledriver Slough	8/4/2012	Angling	Arctic grayling	adult	320	total	238	Female	5		otolith	TBD	
PS-AG-12	Piledriver Slough	8/4/2012	Angling	Arctic grayling	adult	285	total	175	Male	4 or 5		otolith	TBD	
PS-AG-13	Piledriver Slough	8/4/2012	Angling	Arctic grayling	adult	290	total	214	Male	5	5	otolith	TBD	
PS-AG-14	Piledriver Slough	8/7/2012	Angling	Arctic grayling	adult	270	total	168	Female	4		otolith	TBD	
PS-AG-15	Piledriver Slough	8/7/2012	Angling	Arctic grayling	adult	300	total	219	Male	5	5	otolith	TBD	
PS-AG-16	Piledriver Slough	8/8/2012	Angling	Arctic grayling	adult	275	total	165	Female	4	4	otolith	TBD	
PS-AG-17	Piledriver Slough	8/8/2012	Angling	Arctic grayling	adult	280	total	168	Male	4		otolith	TBD	
PS-AG-18	Piledriver Slough	8/8/2012	Angling	Arctic grayling	adult	290	total	207	Female	5		otolith	TBD	
PS-AG-19	Piledriver Slough	8/8/2012	Angling	Arctic grayling	adult	285	total	193	Male	4	4	otolith	TBD	
PS-AG-20	Piledriver Slough	8/8/2012	Angling	Arctic grayling	adult	320	total	258	Female	5		otolith	TBD	

\* = Age determination conducted by Larry Bushing; Quality Control checks conducted by Ken Cummings

\*\* Age Estimated

\*\*\* Minimum Age Estimated

TBD= To Be Determined

**Fish Collection Sample Summary  
Eielson AFB, Alaska**

Sample ID	Name of water body	Date	Fish collection method	Species	Life Stage	Length (mm)	Length method	Weight (g)	Sex	Age*	QC*	Aging method	Fish Tissue Analysis Results	Comments
CR-AG-01	Chena River	8/3/2012	Angling	Arctic grayling	adult	315	total	232	Female	6		otolith	TBD	
CR-AG-02	Chena River	8/3/2012	Angling	Arctic grayling	adult	375	total	365	Male	8 or 9	8 or 9	otolith	TBD	
CR-AG-03	Chena River	8/3/2012	Angling	Arctic grayling	adult	260	total	144	Female	4		otolith	TBD	
CR-AG-04	Chena River	8/3/2012	Angling	Arctic grayling	adult	340	total	299	Female	7		otolith	TBD	
CR-AG-05	Chena River	8/3/2012	Angling	Arctic grayling	adult	300	total	236	Female	5		otolith	TBD	
CR-AG-06	Chena River	8/3/2012	Angling	Arctic grayling	adult	335	total	339	Female	8 or 9	8 or 9	otolith	TBD	
CR-AG-07	Chena River	8/3/2012	Angling	Arctic grayling	adult	290	total	231	Female	4		otolith	TBD	
CR-AG-08	Chena River	8/3/2012	Angling	Arctic grayling	adult	350	total	331	Male	6		otolith	TBD	
CR-AG-09	Chena River	8/3/2012	Angling	Arctic grayling	adult	365	total	390	Female	10 or 11	10 or 11	otolith	TBD	
CR-AG-10	Chena River	8/3/2012	Angling	Arctic grayling	adult	320	total	290	Female	6	6	otolith	TBD	
CR-AG-11	Chena River	8/3/2012	Angling	Arctic grayling	adult	330	total	258	Male	6		otolith	TBD	
CR-AG-12	Chena River	8/3/2012	Angling	Arctic grayling	adult	325	total	293	Male	6		otolith	TBD	
CR-AG-13	Chena River	8/6/2012	Angling	Arctic grayling	adult	360	total	334	Male	7	7	otolith	TBD	
CR-AG-14	Chena River	8/6/2012	Angling	Arctic grayling	adult	340	total	326	Male	5		otolith	TBD	
CR-AG-15	Chena River	8/6/2012	Angling	Arctic grayling	adult	310	total	243	Female	5		otolith	TBD	
CR-AG-16	Chena River	8/6/2012	Angling	Arctic grayling	adult	315	total	257	Male	5		otolith	TBD	
CR-AG-17	Chena River	8/6/2012	Angling	Arctic grayling	adult	310	total	254	Male	5		otolith	TBD	
CR-AG-18	Chena River	8/6/2012	Angling	Arctic grayling	adult	315	total	256	Female	5		otolith	TBD	
CR-AG-19	Chena River	8/6/2012	Angling	Arctic grayling	adult	330	total	306	Female	6	6	otolith	TBD	
CR-AG-20	Chena River	8/6/2012	Angling	Arctic grayling	adult	275	total	179	Female	4		otolith	TBD	

\* = Age determination conducted by Larry Bushing; Quality Control checks conducted by Ken Cummings

\*\* Age Estimated

\*\*\* Minimum Age Estimated

TBD= To Be Determined

**Fish Collection Sample Summary**  
**Eielson AFB, Alaska**

Sample ID	Name of water body	Date	Fish collection method	Species	Life Stage	Length (mm)	Length method	Weight (g)	Sex	Age*	QC*	Aging method	Fish Tissue Analysis Results	Comments
CK-AG-01	Chatanika River	8/1/2012	Angling	Arctic grayling	adult	310	total	240	Male	5		otolith	TBD	
CK-AG-02	Chatanika River	8/1/2012	Angling	Arctic grayling	adult	295	total	194	Male	5		otolith	TBD	
CK-AG-03	Chatanika River	8/1/2012	Angling	Arctic grayling	adult	280	total	174	Female	5		otolith	TBD	
CK-AG-04	Chatanika River	8/1/2012	Angling	Arctic grayling	adult	395	total	477	Male	7***		otolith	TBD	Sample vial contained a single opaque otolith - unusable for age determination,
CK-AG-05	Chatanika River	8/1/2012	Angling	Arctic grayling	adult	345	total	349	Female	7		otolith	TBD	
CK-AG-06	Chatanika River	8/1/2012	Angling	Arctic grayling	adult	280	total	184	Unknown	5		otolith	TBD	
CK-AG-07	Chatanika River	8/1/2012	Angling	Arctic grayling	adult	350	total	333	Male	7		otolith	TBD	
CK-AG-08	Chatanika River	8/1/2012	Angling	Arctic grayling	adult	270	total	153	Female	4		otolith	TBD	
CK-AG-09	Chatanika River	8/1/2012	Angling	Arctic grayling	adult	290	total	172	Male	4		otolith	TBD	Sample vial contained a single otolith - Successfully cleaned and aged whole
CK-AG-10	Chatanika River	8/1/2012	Angling	Arctic grayling	adult	260	total	152	Male	4 or 5	4 or 5	otolith	TBD	Sample vial contained a single otolith - Successfully cleaned and aged whole
CK-AG-11	Chatanika River	8/1/2012	Angling	Arctic grayling	adult	295	total	223	Male	5		otolith	TBD	
CK-AG-12	Chatanika River	8/1/2012	Angling	Arctic grayling	adult	300	total	237	Male	5		otolith	TBD	Sample vial contained a single otolith - Successfully cleaned and aged whole
CK-AG-13	Chatanika River	8/1/2012	Angling	Arctic grayling	adult	315	total	261	Female	7		otolith	TBD	
CK-AG-14	Chatanika River	8/1/2012	Angling	Arctic grayling	adult	295	total	195	Male	4		otolith	TBD	
CK-AG-15	Chatanika River	8/1/2012	Angling	Arctic grayling	adult	295	total	203	Unknown	5		otolith	TBD	
CK-AG-16	Chatanika River	8/5/2012	Angling	Arctic grayling	adult	290	total	209	Female	5		otolith	TBD	
CK-AG-17	Chatanika River	8/5/2012	Angling	Arctic grayling	adult	355	total	337	Male	6		otolith	TBD	
CK-AG-18	Chatanika River	8/5/2012	Angling	Arctic grayling	adult	360	total	383	Female	6		otolith	TBD	
CK-AG-19	Chatanika River	8/5/2012	Angling	Arctic grayling	adult	290	total	190	Female	5		otolith	TBD	
CK-AG-20	Chatanika River	8/5/2012	Angling	Arctic grayling	adult	340	total	330	Male	7	7	otolith	TBD	

\* Age determination conducted by Larry Bushing; Quality Control checks conducted by Ken Cummings

\*\* Age Estimated

\*\*\* Minimum Age Estimated

TBD= To Be Determined

**Tissue Sample Summary  
Eielson AFB, Alaska**

Lab	SDG	CoC Number	Cooler Name	Location ID	Sample ID	Sample Date	Sample Time	Method	Matrix	Container Type/ Volume	QC	Sample Depth	Field Preservation	Sampler	TAT
STLP	180-13438-1	SS67-COC001	-	GSAG01-F	GS-AG-01-FILLET	31-Jul-12	1250	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG01-W	GS-AG-01-WHOLE	31-Jul-12	1250	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG02-G	GS-AG-02-FILLET	31-Jul-12	1345	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG02-W	GS-AG-02-WHOLE	31-Jul-12	1345	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG03-F	GS-AG-03-FILLET	31-Jul-12	1405	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG03-W	GS-AG-03-WHOLE	31-Jul-12	1405	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG04-F	GS-AG-04-FILLET	31-Jul-12	1545	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG04-W	GS-AG-04-WHOLE	31-Jul-12	1545	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG05-F	GS-AG-05-FILLET	31-Jul-12	1550	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG05-W	GS-AG-05-WHOLE	31-Jul-12	1550	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG06-F	GS-AG-06-FILLET	31-Jul-12	1600	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG06-W	GS-AG-06-WHOLE	31-Jul-12	1600	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG07-F	GS-AG-07-FILLET	02-Aug-12	0829	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG07-W	GS-AG-07-WHOLE	02-Aug-12	0829	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG08-F	GS-AG-08-FILLET	02-Aug-12	0930	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG08-W	GS-AG-08-WHOLE	02-Aug-12	0930	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG09-F	GS-AG-09-FILLET	02-Aug-12	0940	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG09-W	GS-AG-09-WHOLE	02-Aug-12	0940	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG10-F	GS-AG-10-FILLET	02-Aug-12	0842	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG10-W	GS-AG-10-WHOLE	02-Aug-12	0842	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG11-F	GS-AG-11-FILLET	02-Aug-12	0843	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG11-W	GS-AG-11-WHOLE	02-Aug-12	0843	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG12-F	GS-AG-12-FILLET	02-Aug-12	0855	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG12-W	GS-AG-12-WHOLE	02-Aug-12	0855	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG13-F	GS-AG-13-FILLET	02-Aug-12	0906	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG13-W	GS-AG-13-WHOLE	02-Aug-12	0906	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG14-F	GS-AG-14-FILLET	02-Aug-12	0937	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days

**Tissue Sample Summary**  
**Eielson AFB, Alaska**

Lab	SDG	CoC Number	Cooler Name	Location ID	Sample ID	Sample Date	Sample Time	Method	Matrix	Container Type/ Volume	QC	Sample Depth	Field Preservation	Sampler	TAT
STLP	180-13438-1	SS67-COC001	-	GSAG14-W	GS-AG-14-WHOLE	02-Aug-12	0937	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG15-F	GS-AG-15-FILLET	02-Aug-12	0955	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG15-W	GS-AG-15-WHOLE	02-Aug-12	0955	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG16-F	GS-AG-16-FILLET	05-Aug-12	1419	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG16-W	GS-AG-16-WHOLE	05-Aug-12	1419	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG17-F	GS-AG-17-FILLET	05-Aug-12	1426	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG17-W	GS-AG-17-WHOLE	05-Aug-12	1426	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG18-F	GS-AG-18-FILLET	05-Aug-12	1436	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG18-W	GS-AG-18-WHOLE	05-Aug-12	1436	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG19-F	GS-AG-19-FILLET	05-Aug-12	1500	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG19-W	GS-AG-19-WHOLE	05-Aug-12	1500	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG20-F	GS-AG-20-FILLET	05-Aug-12	1520	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG20-W	GS-AG-20-WHOLE	05-Aug-12	1520	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-13438-1	SS67-COC001	-	GSAG21-F	GS-AG-21-FILLET	02-Aug-12	0800	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG01-F	CR-AG-01-Fillet	03-Aug-12	1005	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG01-W	CR-AG-01-Whole	03-Aug-12	1005	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG02-F	CR-AG-02-Fillet	03-Aug-12	1045	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG02-W	CR-AG-02-Whole	03-Aug-12	1045	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG03-F	CR-AG-03-Fillet	03-Aug-12	1100	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG03-W	CR-AG-03-Whole	03-Aug-12	1100	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG04-F	CR-AG-04-Fillet	03-Aug-12	1110	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG04-W	CR-AG-04-Whole	03-Aug-12	1110	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG05-F	CR-AG-05-Fillet	03-Aug-12	1113	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG05-W	CR-AG-05-Whole	03-Aug-12	1113	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG06-F	CR-AG-06-Fillet	03-Aug-12	1335	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG06-W	CR-AG-06-Whole	03-Aug-12	1335	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG07-F	CR-AG-07-Fillet	03-Aug-12	1344	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days

**Tissue Sample Summary  
Eielson AFB, Alaska**

Lab	SDG	CoC Number	Cooler Name	Location ID	Sample ID	Sample Date	Sample Time	Method	Matrix	Container Type/ Volume	QC	Sample Depth	Field Preservation	Sampler	TAT
STLP	180-18147-1	SS67-COC001	-	CRAG07-W	CR-AG-07-Whole	03-Aug-12	1344	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG08-F	CR-AG-08-Fillet	03-Aug-12	1430	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG08-W	CR-AG-08-Whole	03-Aug-12	1430	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG09-F	CR-AG-09-Fillet	03-Aug-12	1450	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG09-W	CR-AG-09-Whole	03-Aug-12	1450	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG10-F	CR-AG-10-Fillet	03-Aug-12	1510	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG10-W	CR-AG-10-Whole	03-Aug-12	1510	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG11-F	CR-AG-11-Fillet	03-Aug-12	1515	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG11-W	CR-AG-11-Whole	03-Aug-12	1515	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG12-F	CR-AG-12-Fillet	03-Aug-12	1540	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG12-W	CR-AG-12-Whole	03-Aug-12	1540	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG13-F	CR-AG-13-Fillet	06-Aug-12	0920	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG13-W	CR-AG-13-Whole	06-Aug-12	0920	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG14-F	CR-AG-14-Fillet	06-Aug-12	0940	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG14-W	CR-AG-14-Whole	06-Aug-12	0940	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG15-F	CR-AG-15-Fillet	06-Aug-12	1014	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG15-W	CR-AG-15-Whole	06-Aug-12	1014	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG16-F	CR-AG-16-Fillet	06-Aug-12	1040	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG16-W	CR-AG-16-Whole	06-Aug-12	1040	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG17-F	CR-AG-17-Fillet	06-Aug-12	1140	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG17-W	CR-AG-17-Whole	06-Aug-12	1140	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG18-F	CR-AG-18-Fillet	06-Aug-12	1155	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG18-W	CR-AG-18-Whole	06-Aug-12	1155	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG19-F	CR-AG-19-Fillet	06-Aug-12	1210	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG19-W	CR-AG-19-Whole	06-Aug-12	1210	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG20-F	CR-AG-20-Fillet	06-Aug-12	1215	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18147-1	SS67-COC001	-	CRAG20-W	CR-AG-20-Whole	06-Aug-12	1215	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days

**Tissue Sample Summary  
Eielson AFB, Alaska**

Lab	SDG	CoC Number	Cooler Name	Location ID	Sample ID	Sample Date	Sample Time	Method	Matrix	Container Type/ Volume	QC	Sample Depth	Field Preservation	Sampler	TAT
STLP	180-18147-1	SS67-COC001	-	CRAG21-F	CR-AG-21-Fillet	03-Aug-12	0800	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG01-F	CK-AG-01-Fillet	01-Aug-12	0915	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG01-W	CK-AG-01-Whole	01-Aug-12	0915	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG02-F	CK-AG-02-Fillet	01-Aug-12	1015	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG02-W	CK-AG-02-Whole	01-Aug-12	1015	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG03-F	CK-AG-03-Fillet	01-Aug-12	1049	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG03-W	CK-AG-03-Whole	01-Aug-12	1049	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG04-F	CK-AG-04-Fillet	01-Aug-12	1143	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG04-W	CK-AG-04-Whole	01-Aug-12	1143	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG05-F	CK-AG-05-Fillet	01-Aug-12	1157	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG05-W	CK-AG-05-Whole	01-Aug-12	1157	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG06-F	CK-AG-06-Fillet	01-Aug-12	1205	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG06-W	CK-AG-06-Whole	01-Aug-12	1205	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG07-F	CK-AG-07-Fillet	01-Aug-12	1215	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG07-W	CK-AG-07-Whole	01-Aug-12	1215	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG08-F	CK-AG-08-Fillet	01-Aug-12	1215	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG08-W	CK-AG-08-Whole	01-Aug-12	1215	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG09-F	CK-AG-09-Fillet	01-Aug-12	1412	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG09-W	CK-AG-09-Whole	01-Aug-12	1412	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG10-F	CK-AG-10-Fillet	01-Aug-12	1418	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG10-W	CK-AG-10-Whole	01-Aug-12	1418	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG11-F	CK-AG-11-Fillet	01-Aug-12	1424	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG11-W	CK-AG-11-Whole	01-Aug-12	1424	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG12-F	CK-AG-12-Fillet	01-Aug-12	1426	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG12-W	CK-AG-12-Whole	01-Aug-12	1426	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG13-F	CK-AG-13-Fillet	01-Aug-12	1446	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG13-W	CK-AG-13-Whole	01-Aug-12	1446	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days



**Tissue Sample Summary**  
**Eielson AFB, Alaska**

Lab	SDG	CoC Number	Cooler Name	Location ID	Sample ID	Sample Date	Sample Time	Method	Matrix	Container Type/ Volume	QC	Sample Depth	Field Preservation	Sampler	TAT
STLP	180-18152-1	SS67-COC001	-	CKAG14-F	CK-AG-14-Fillet	01-Aug-12	1440	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG14-W	CK-AG-14-Whole	01-Aug-12	1440	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG15-F	CK-AG-15-Fillet	01-Aug-12	1500	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG15-W	CK-AG-15-Whole	01-Aug-12	1500	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG16-F	CK-AG-16-Fillet	05-Aug-12	0950	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG16-W	CK-AG-16-Whole	05-Aug-12	0950	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG17-F	CK-AG-17-Fillet	05-Aug-12	0958	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG17-W	CK-AG-17-Whole	05-Aug-12	0958	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG18-F	CK-AG-18-Fillet	05-Aug-12	1018	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG18-W	CK-AG-18-Whole	05-Aug-12	1018	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG19-F	CK-AG-19-Fillet	05-Aug-12	1030	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG19-W	CK-AG-19-Whole	05-Aug-12	1030	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG20-F	CK-AG-20-Fillet	05-Aug-12	1113	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG20-W	CK-AG-20-Whole	05-Aug-12	1113	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18152-1	SS67-COC001	-	CKAG21-F	CK-AG-21-Fillet	01-Aug-12	0800	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG01-F	PS-AG-01-Fillet	02-Aug-12	1240	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG01-W	PS-AG-01-Whole	02-Aug-12	1240	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG02-F	PS-AG-02-Fillet	03-Aug-12	1245	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG02-W	PS-AG-02-Whole	03-Aug-12	1245	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG03-F	PS-AG-03-Fillet	03-Aug-12	1327	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG03-W	PS-AG-03-Whole	03-Aug-12	1327	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG04-F	PS-AG-04-Fillet	03-Aug-12	1400	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG04-W	PS-AG-04-Whole	03-Aug-12	1400	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG05-F	PS-AG-05-Fillet	03-Aug-12	1405	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG05-W	PS-AG-05-Whole	03-Aug-12	1405	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG06-F	PS-AG-06-Fillet	03-Aug-12	1420	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG06-W	PS-AG-06-Whole	03-Aug-12	1420	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days

**Tissue Sample Summary  
Eielson AFB, Alaska**

Lab	SDG	CoC Number	Cooler Name	Location ID	Sample ID	Sample Date	Sample Time	Method	Matrix	Container Type/ Volume	QC	Sample Depth	Field Preservation	Sampler	TAT
STLP	180-18159-1	SS67-COC001	-	PSAG07-F	PS-AG-07-Fillet	03-Aug-12	1733	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG07-W	PS-AG-07-Whole	03-Aug-12	1733	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG08-F	PS-AG-08-Fillet	04-Aug-12	1017	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG08-W	PS-AG-08-Whole	04-Aug-12	1017	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG09-F	PS-AG-09-Fillet	04-Aug-12	1022	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG09-W	PS-AG-09-Whole	04-Aug-12	1022	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG10-F	PS-AG-10-Fillet	04-Aug-12	1030	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG10-W	PS-AG-10-Whole	04-Aug-12	1030	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG11-F	PS-AG-11-Fillet	04-Aug-12	1040	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG11-W	PS-AG-11-Whole	04-Aug-12	1040	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG12-F	PS-AG-12-Fillet	04-Aug-12	1106	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG12-W	PS-AG-12-Whole	04-Aug-12	1106	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG13-F	PS-AG-13-Fillet	04-Aug-12	1151	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG13-W	PS-AG-13-Whole	04-Aug-12	1151	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG14-F	PS-AG-14-Fillet	07-Aug-12	0930	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG14-W	PS-AG-14-Whole	07-Aug-12	0930	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG15-F	PS-AG-15-Fillet	07-Aug-12	1035	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG15-W	PS-AG-15-Whole	07-Aug-12	1035	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG16-F	PS-AG-16-Fillet	08-Aug-12	1142	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG16-W	PS-AG-16-Whole	08-Aug-12	1142	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG17-F	PS-AG-17-Fillet	08-Aug-12	1151	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG17-W	PS-AG-17-Whole	08-Aug-12	1151	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG18-F	PS-AG-18-Fillet	08-Aug-12	1220	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG18-W	PS-AG-18-Whole	08-Aug-12	1220	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG19-F	PS-AG-19-Fillet	08-Aug-12	1441	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG19-W	PS-AG-19-Whole	08-Aug-12	1441	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG20-F	PS-AG-20-Fillet	08-Aug-12	1506	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days

**Tissue Sample Summary**  
**Eielson AFB, Alaska**

Lab	SDG	CoC Number	Cooler Name	Location ID	Sample ID	Sample Date	Sample Time	Method	Matrix	Container Type/ Volume	QC	Sample Depth	Field Preservation	Sampler	TAT
STLP	180-18159-1	SS67-COC001	-	PSAG20-W	PS-AG-20-Whole	08-Aug-12	1506	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
STLP	180-18159-1	SS67-COC001	-	PSAG21-F	PS-AG-21-Fillet	01-Aug-12	0800	SW7471B, SW8082A, SW8081B, BDTL	TF	N/A	-	N/A	Frozen	EA Engineering	30 days
TA	580-36027-1	12EAFB-SS67-01	-	SS67-W01	12EAFB-SS67-SO-W01	11/15/2012	1030	SW8082A, SW7471B, SW6020	SE	Amber Glass/ 8 oz.	-	composite	4°C	Jacobs Engineering	14 days

**Notes:**

CoC = contaminant of concern

SDG = sample delivery group

STLP = TestAmerica, Pittsburgh

TA = TestAmerica, Tacoma

TAT = turnaround time

QC - none specified

**Sediment Analytical Results**  
**Eielson AFB, Alaska**

				Location ID	SS67-W01
				Sample ID	12EAFB-SS67-SO-W01
				Lab Sample ID	580-36027-1
				Date Collected	11/15/2012
				Matrix	SO
				Laboratory	TA
Method	Analyte	Units	PAL <sup>1</sup>		
D2216	Total Solids	PERCENT	-	62	
D2216	Percent Moisture	PERCENT	-	38	
SW6020	Arsenic	mg/kg	3.9	<b>140 [0.63]</b>	
SW6020	Barium	mg/kg	1100	250 [0.063]	
SW6020	Cadmium	mg/kg	5	0.35 [0.032]	
SW6020	Chromium	mg/kg	25	17 [0.24]	
SW6020	Lead	mg/kg	400	21 [0.032]	
SW6020	Selenium	mg/kg	3.4	0.54 [0.63]	
SW6020	Silver	mg/kg	11.2	0.059 [0.032]	
SW7471	Mercury	mg/kg	0.07	0.045 [0.013]	
SW8082	PCB-1016 (Aroclor 1016)	mg/kg	0.14	ND [0.37] JS-	
SW8082	PCB-1221 (Aroclor 1221)	mg/kg	0.14	ND [0.73] JS-	
SW8082	PCB-1232 (Aroclor 1232)	mg/kg	0.14	ND [0.73] JS-	
SW8082	PCB-1242 (Aroclor 1242)	mg/kg	0.14	ND [0.37] JS-	
SW8082	PCB-1248 (Aroclor 1248)	mg/kg	0.14	ND [0.37] JS-	
SW8082	PCB-1254 (Aroclor 1254)	mg/kg	0.06	ND [0.37] JS-	
SW8082	PCB-1260 (Aroclor 1260)	mg/kg	0.14	<b>19 [0.37] JS-</b>	

**Notes:**

<sup>1</sup> Project Action Levels (PAL) are defined in the Draft Final Quality Assurance Project Plan Remedial Investigation Source Area SS67. If an analyte did not have an associated PAL, ADEC Method 2 migration to groundwater cleanup criteria were used.

The action limit for lead is the ADEC Method 2 direct contact cleanup criteria

TA = TestAmerica, Seattle, WA

JS- = The result is estimated because the surrogate failed recovery criteria (low).

**Bold** = The sample result exceeded the waste criteria.

[ ] = Limit of detection (LOD)

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				CKAG01-F CK-AG-01-Fillet 18152-1 8/1/2012 TF STLP	CKAG01-W CK-AG-01-Whole 18152-2 8/1/2012 TF STLP	CKAG02-F CK-AG-02-Fillet 18152-3 8/1/2012 TF STLP	CKAG02-W CK-AG-02-Whole 18152-4 8/1/2012 TF STLP	CKAG03-F CK-AG-03-Fillet 18152-5 8/1/2012 TF STLP
Method	Analyte	Units	Project Action Limit					
BDTL	Total Lipids	Percent	-	0.53 [0.03]	2.1 [0.03]	0.29 [0.03]	1.1 [0.029]	0.63 [0.03]
SW7471B	Mercury	mg/kg	0.014	<b>0.082 [0.011]</b>	<b>0.043 [0.01]</b>	<b>0.064 [0.011]</b>	<b>0.041 [0.01]</b>	<b>0.061 [0.011]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.00029 [0.000054] J	0.00055 [0.000055]	0.00083 [0.000054] J	0.00091 [0.000055] J	0.00095 [0.000055] J
SW8081B	4,4'-DDE	mg/kg	0.0093	0.00015 [0.000063] J	0.00065 [0.000063]	ND [0.000063]	0.00014 [0.000063] J	ND [0.000063]
SW8081B	4,4'-DDT	mg/kg	0.0093	ND [0.000062]	0.00012 [0.000063] J	ND [0.000062]	ND [0.000063]	ND [0.000063]
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000074]	ND [0.000075]	ND [0.000074]	ND [0.000075]	ND [0.000075]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000082]	0.00084 [0.000083] J	ND [0.000082]	ND [0.000083]	ND [0.000083]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]
SW8081B	Dieldrin	mg/kg	0.0002	ND [0.000069]	0.00015 [0.00007] J	ND [0.000069]	ND [0.00007]	ND [0.00007]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000078]	ND [0.000079]	ND [0.000078]	ND [0.000079]	ND [0.000079]
SW8081B	Endosulfan II	mg/kg	0.811	0.0001 [0.000073] J	ND [0.000074]	ND [0.000073]	ND [0.000074]	ND [0.000074]
SW8081B	Endosulfan sulfate	mg/kg	0.811	ND [0.000043]	ND [0.000044]	ND [0.000043]	ND [0.000044]	ND [0.000044]
SW8081B	Endrin	mg/kg	0.041	ND [0.00008]	0.00013 [0.000081] J	ND [0.00008]	ND [0.000081]	ND [0.000081]
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.00008]	ND [0.000081]	ND [0.00008]	ND [0.000081]	ND [0.000081]
SW8081B	Endrin ketone	mg/kg	-	ND [0.000065]	0.00014 [0.000065] J	ND [0.000065]	ND [0.000065]	ND [0.000065]
SW8081B	gamma-BHC	mg/kg	0.0029	0.00024 [0.000073] J	0.00019 [0.000073] J	0.00019 [0.000073] J	0.00027 [0.000073] J	0.00023 [0.000073] J
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000081]	0.00013 [0.000082] J	ND [0.000081]	ND [0.000082]	ND [0.000082]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000092]	ND [0.000093]	ND [0.000092]	ND [0.000093]	ND [0.000093]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.00008]	ND [0.000081]	ND [0.00008]	ND [0.000081]	ND [0.000081]
SW8081B	Methoxychlor	mg/kg	0.676	0.00011 [0.000086] J, B	ND [0.000087]	ND [0.000086]	ND [0.000087]	ND [0.000087]
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.0008]	ND [0.00079]	ND [0.00079]	ND [0.0008]	ND [0.0008]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00068]	ND [0.00067]	ND [0.00067]	ND [0.00068]	ND [0.00068]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	ND [0.00059]	<b>0.023 [0.00059]</b>	<b>0.0079 [0.00059]</b>	ND [0.00059]	ND [0.00059]
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	0.0016 [0.00059] J	<b>0.015 [0.00059]</b>	<b>0.011 [0.00059]</b>	<b>0.0093 [0.00059]</b>	ND [0.00059]

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E (Italics)* = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				CKAG03-W CK-AG-03-Whole 18152-6 8/1/2012 TF STLP	CKAG04-F CK-AG-04-Fillet 18152-7 8/1/2012 TF STLP	CKAG04-W CK-AG-04-Whole 18152-8 8/1/2012 TF STLP	CKAG05-F CK-AG-05-Fillet 18152-9 8/1/2012 TF STLP	CKAG05-W CK-AG-05-Whole 18152-10 8/1/2012 TF STLP	CKAG06-F CK-AG-06-Fillet 18152-11 8/1/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	2.7 [0.029]	0.67 [0.029]	1.8 [0.03]	1.1 [0.03]	3.3 [0.03]	0.74 [0.03]
SW7471B	Mercury	mg/kg	0.014	<b>0.031 [0.011] J</b>	<b>0.22 [0.011]</b>	<b>0.14 [0.011]</b>	<b>0.2 [0.01]</b>	<b>0.084 [0.011]</b>	<b>0.049 [0.011]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.00016 [0.000054] J	0.00024 [0.000055] J	0.0011 [0.000054]	0.0002 [0.000054] J	0.00027 [0.000054] J	0.00026 [0.000055] J
SW8081B	4,4'-DDE	mg/kg	0.0093	0.00032 [0.000063] J	0.00069 [0.000063]	0.0034 [0.000063]	0.00034 [0.000063] J	0.0011 [0.000063]	0.00013 [0.000063] J
SW8081B	4,4'-DDT	mg/kg	0.0093	0.000071 [0.000062] J	0.000076 [0.000063] J	0.00039 [0.000062] J	ND [0.000062]	0.00015 [0.000062] J	ND [0.000063]
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000074]	ND [0.000075]	ND [0.000074]	ND [0.000074]	ND [0.000074]	ND [0.000075]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000082]	ND [0.000083]	0.00011 [0.000082] J	ND [0.000082]	0.000082 [0.000082] J	ND [0.000083]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]
SW8081B	Dieldrin	mg/kg	0.0002	ND [0.000069]	ND [0.00007]	ND [0.000069]	ND [0.000069]	0.00018 [0.000069] J	ND [0.00007]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000078]	ND [0.000079]	ND [0.000078]	ND [0.000078]	ND [0.000078]	ND [0.000079]
SW8081B	Endosulfan II	mg/kg	0.811	ND [0.000073]	ND [0.000074]	ND [0.000073]	ND [0.000073]	0.000091 [0.000073] J	ND [0.000074]
SW8081B	Endosulfan sulfate	mg/kg	0.811	0.0005 [0.000043]	ND [0.000044]	ND [0.000043]	ND [0.000043]	0.00022 [0.000043] J	ND [0.000044]
SW8081B	Endrin	mg/kg	0.041	ND [0.00008]	ND [0.000081]	0.00022 [0.00008] J	ND [0.00008]	0.000085 [0.00008] J	0.00016 [0.000081] J
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.00008]	ND [0.000081]	ND [0.00008]	ND [0.00008]	ND [0.00008]	ND [0.000081]
SW8081B	Endrin ketone	mg/kg	-	0.000095 [0.000065] J	ND [0.000065]	0.00028 [0.000065] J	ND [0.000065]	0.0011 [0.000065]	ND [0.000065]
SW8081B	gamma-BHC	mg/kg	0.0029	0.00018 [0.000073] J	0.00022 [0.000073] J	0.00021 [0.000073] J	0.00018 [0.000073] J	0.00019 [0.000073] J	0.000078 [0.000073] J
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000081]	ND [0.000082]	0.00035 [0.000081] J	ND [0.000081]	ND [0.000081]	ND [0.000082]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000092]	ND [0.000093]	ND [0.000092]	ND [0.000092]	ND [0.000092]	ND [0.000093]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.00008]	ND [0.000081]	0.00009 [0.00008] J	ND [0.00008]	ND [0.00008]	ND [0.000081]
SW8081B	Methoxychlor	mg/kg	0.676	ND [0.000086]	ND [0.000087]	ND [0.000086]	ND [0.000086]	0.00011 [0.000086] J, B	0.00026 [0.000087] J
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.00079]	ND [0.0008]	ND [0.00079]	ND [0.00079]	ND [0.0008]	ND [0.00079]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00067]	ND [0.00068]	ND [0.00067]	ND [0.00067]	ND [0.00068]	ND [0.00067]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	ND [0.00059]	<b>0.012 [0.00059]</b>	<b>0.0062 [0.00059]</b>	ND [0.00059]	ND [0.00059]	ND [0.00059]
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	0.0014 [0.00059] J	<b>0.02 [0.00059]</b>	<b>0.0088 [0.00059]</b>	0.0021 [0.00059] J	<b>0.015 [0.00059]</b>	<b>0.0028 [0.00059] J</b>

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E (Italics)* = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				CKAG06-W CK-AG-06-Whole 18152-12 8/1/2012 TF STLP	CKAG07-F CK-AG-07-Fillet 18152-13 8/1/2012 TF STLP	CKAG07-W CK-AG-07-Whole 18152-14 8/1/2012 TF STLP	CKAG08-F CK-AG-08-Fillet 18152-15 8/1/2012 TF STLP	CKAG08-W CK-AG-08-Whole 18152-16 8/1/2012 TF STLP
Method	Analyte	Units	Project Action Limit					
BDTL	Total Lipids	Percent	-	2.9 [0.03]	0.83 [0.03]	2.3 [0.03]	0.91 [0.03]	3.3 [0.03]
SW7471B	Mercury	mg/kg	0.014	<b>0.03 [0.01] J</b>	<b>0.11 [0.011]</b>	<b>0.068 [0.01]</b>	<b>0.046 [0.01]</b>	<b>0.022 [0.0098] J</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.00025 [0.000055] J	0.00022 [0.000055] J	0.00033 [0.000054] J	0.0002 [0.000055] J	0.00025 [0.000055] J
SW8081B	4,4'-DDE	mg/kg	0.0093	0.00041 [0.000063] J	ND [0.000063]	0.00031 [0.000063] J	0.00076 [0.000063] J	0.00036 [0.000063] J
SW8081B	4,4'-DDT	mg/kg	0.0093	0.00011 [0.000063] J	ND [0.000063]	0.0002 [0.000062] J	ND [0.000063]	0.00076 [0.000063] J
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000075]	ND [0.000075]	ND [0.000074]	ND [0.000075]	ND [0.000075]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000068]	ND [0.000068]	0.00018 [0.000068] J	ND [0.000068]	ND [0.000068]
SW8081B	alpha-Chlordane	mg/kg	0.009	0.00089 [0.000083] J	ND [0.000083]	ND [0.000082]	ND [0.000083]	ND [0.000083]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]
SW8081B	Dieldrin	mg/kg	0.0002	0.00018 [0.00007] J	ND [0.00007]	0.00073 [0.000069] J	ND [0.00007]	ND [0.00007]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000079]	ND [0.000079]	ND [0.000078]	ND [0.000079]	ND [0.000079]
SW8081B	Endosulfan II	mg/kg	0.811	0.00012 [0.000074] J	ND [0.000074]	0.00012 [0.000073] J	ND [0.000074]	ND [0.000074]
SW8081B	Endosulfan sulfate	mg/kg	0.811	ND [0.000044]	ND [0.000044]	0.00027 [0.000043] J	0.00015 [0.000044] J	0.00062 [0.000044]
SW8081B	Endrin	mg/kg	0.041	0.00011 [0.000081] J	0.00092 [0.000081] J	0.00011 [0.00008] J	0.00012 [0.000081] J	0.00015 [0.000081] J
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.000081]	ND [0.000081]	0.0001 [0.00008] J	ND [0.000081]	ND [0.000081]
SW8081B	Endrin ketone	mg/kg	-	ND [0.000065]	ND [0.000065]	0.0016 [0.000065]	ND [0.000065]	ND [0.000065]
SW8081B	gamma-BHC	mg/kg	0.0029	0.00025 [0.000073] J	0.00018 [0.000073] J	0.00013 [0.000073] J	ND [0.000073]	0.00019 [0.000073] J
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000082]	ND [0.000082]	ND [0.000081]	ND [0.000082]	ND [0.000082]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000093]	ND [0.000093]	ND [0.000092]	ND [0.000093]	ND [0.000093]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.000081]	ND [0.000081]	ND [0.00008]	ND [0.000081]	ND [0.000081]
SW8081B	Methoxychlor	mg/kg	0.676	0.00017 [0.000087] J, B	0.00014 [0.000087] J, B	0.00015 [0.000086] J, B	0.00014 [0.000087] J, B	0.00017 [0.000087] J, B
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.00079]	ND [0.0008]	ND [0.0008]	ND [0.0008]	ND [0.0008]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00067]	ND [0.00068]	ND [0.00068]	ND [0.00068]	ND [0.00068]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	ND [0.00059]	ND [0.00059]	ND [0.00059]	ND [0.00059]	ND [0.00059]
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.0035 [0.00059] J</b>	<b>0.0078 [0.00059]</b>	0.0016 [0.00059] J	ND [0.00059]	<b>0.0041 [0.00059] J</b>

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E (Italics)* = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				CKAG09-F CK-AG-09-Fillet 18152-17 8/1/2012 TF STLP	CKAG09-W CK-AG-09-Whole 18152-18 8/1/2012 TF STLP	CKAG10-F CK-AG-10-Fillet 18152-19 8/1/2012 TF STLP	CKAG10-W CK-AG-10-Whole 18152-20 8/1/2012 TF STLP	CKAG11-F CK-AG-11-Fillet 18152-21 8/1/2012 TF STLP
Method	Analyte	Units	Project Action Limit					
BDTL	Total Lipids	Percent	-	0.56 [0.03]	2.8 [0.03]	0.14 [0.029]	1 [0.029]	0.65 [0.03]
SW7471B	Mercury	mg/kg	0.014	<b>0.056 [0.011]</b>	<b>0.029 [0.01] J</b>	<b>0.051 [0.011]</b>	<b>0.03 [0.011] J</b>	<b>0.062 [0.01]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.00023 [0.000054] J	0.00034 [0.000054] J	0.00093 [0.000055] J	0.00066 [0.000055]	0.00036 [0.000054] J
SW8081B	4,4'-DDE	mg/kg	0.0093	0.00011 [0.000063] J	0.00081 [0.000063]	ND [0.000063]	0.00054 [0.000063]	0.00024 [0.000063] J
SW8081B	4,4'-DDT	mg/kg	0.0093	ND [0.000062]	0.00012 [0.000062] J	ND [0.000063]	0.00016 [0.000063] J	0.0011 [0.000062]
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000074]	ND [0.000074]	ND [0.000075]	ND [0.000075]	ND [0.000074]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000082]	ND [0.000082]	ND [0.000083]	ND [0.000083]	ND [0.000082]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]
SW8081B	Dieldrin	mg/kg	0.0002	ND [0.000069]	0.00013 [0.000069] J	ND [0.00007]	0.00012 [0.00007] J	<b>0.00058 [0.000069]</b>
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000078]	ND [0.000078]	ND [0.000079]	ND [0.000079]	0.00018 [0.000078] J
SW8081B	Endosulfan II	mg/kg	0.811	ND [0.000073]	ND [0.000073]	0.00013 [0.000074] J	ND [0.000074]	0.00012 [0.000073] J
SW8081B	Endosulfan sulfate	mg/kg	0.811	ND [0.000043]	ND [0.000043]	ND [0.000044]	0.000054 [0.000044] J	0.00037 [0.000043] J
SW8081B	Endrin	mg/kg	0.041	0.00011 [0.00008] J	0.00016 [0.00008] J	0.00012 [0.000081] J	0.00011 [0.000081] J	0.0038 [0.00008]
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.00008]	ND [0.00008]	ND [0.000081]	0.00011 [0.000081] J	ND [0.00008]
SW8081B	Endrin ketone	mg/kg	-	ND [0.000065]	0.00043 [0.000065]	ND [0.000065]	0.00025 [0.000065] J	0.0016 [0.000065]
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.000073]	0.00019 [0.000073] J	0.00018 [0.000073] J	0.00019 [0.000073] J	0.00021 [0.000073] J
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000081]	ND [0.000081]	ND [0.000082]	ND [0.000082]	0.00088 [0.000081]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000092]	ND [0.000092]	ND [0.000093]	ND [0.000093]	ND [0.000092]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.00008]	ND [0.00008]	ND [0.000081]	0.000084 [0.000081] J	0.00019 [0.00008] J
SW8081B	Methoxychlor	mg/kg	0.676	0.00017 [0.000086] J, B	0.00013 [0.000086] J, B	ND [0.000087]	ND [0.000087]	0.0025 [0.000086]
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.0008]	ND [0.00079]	ND [0.0008]	ND [0.00079]	ND [0.0008]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00068]	ND [0.00067]	ND [0.00068]	ND [0.00067]	ND [0.00068]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	<b>0.089 [0.00059]</b>	ND [0.00059]	ND [0.00059]	ND [0.00059]	ND [0.00059]
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	ND [0.00059]	<b>0.0032 [0.00059] J</b>	<b>0.003 [0.00059] J</b>	<b>0.0074 [0.00059]</b>	<b>0.0034 [0.00059] J</b>

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E (Italics)* = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).



**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				CKAG11-W CK-AG-11-Whole 18152-22 8/1/2012 TF STLP	CKAG12-F CK-AG-12-Fillet 18152-23 8/1/2012 TF STLP	CKAG12-W CK-AG-12-Whole 18152-24 8/1/2012 TF STLP	CKAG13-F CK-AG-13-Fillet 18152-25 8/1/2012 TF STLP	CKAG13-W CK-AG-13-Whole 18152-26 8/1/2012 TF STLP	CKAG14-F CK-AG-14-Fillet 18152-27 8/1/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	2.6 [0.03]	0.5 [0.029]	2.5 [0.03]	0.82 [0.03]	3 [0.03]	0.17 [0.03]
SW7471B	Mercury	mg/kg	0.014	<b>0.026 [0.01] J</b>	<b>0.11 [0.011]</b>	<b>0.069 [0.011]</b>	<b>0.1 [0.01]</b>	<b>0.051 [0.011]</b>	<b>0.082 [0.011]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.00038 [0.000054] J	0.00011 [0.000055] J	0.00024 [0.000055] J	0.00013 [0.000055] J	0.00046 [0.000055] J	0.000078 [0.000055] J
SW8081B	4,4'-DDE	mg/kg	0.0093	0.00069 [0.000063] J	ND [0.000063]	0.00018 [0.000063] J	0.00015 [0.000063] J	0.00047 [0.000063] J	ND [0.000063]
SW8081B	4,4'-DDT	mg/kg	0.0093	0.00029 [0.000062] J	ND [0.000063]	ND [0.000063]	ND [0.000063]	0.0001 [0.000063] J	ND [0.000063]
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000074]	ND [0.000075]	ND [0.000075]	ND [0.000075]	ND [0.000075]	ND [0.000075]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]
SW8081B	alpha-Chlordane	mg/kg	0.009	0.000099 [0.000082] J	ND [0.000083]	ND [0.000083]	ND [0.000083]	ND [0.000083]	ND [0.000083]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]
SW8081B	Dieldrin	mg/kg	0.0002	0.00016 [0.000069] J	ND [0.00007]	ND [0.00007]	ND [0.00007]	0.000075 [0.00007] J	ND [0.00007]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000078]	ND [0.000079]	ND [0.000079]	ND [0.000079]	ND [0.000079]	ND [0.000079]
SW8081B	Endosulfan II	mg/kg	0.811	0.000081 [0.000073] J	0.00012 [0.000074] J	0.00015 [0.000074] J	ND [0.000074] J	0.0001 [0.000074] J	0.0001 [0.000074] J
SW8081B	Endosulfan sulfate	mg/kg	0.811	ND [0.000043]	ND [0.000044]	0.000047 [0.000044] J	ND [0.000044]	ND [0.000044]	ND [0.000044]
SW8081B	Endrin	mg/kg	0.041	0.00012 [0.00008] J	0.00017 [0.000081] J	0.0001 [0.000081] J	ND [0.000081]	0.00015 [0.000081] J	0.000093 [0.000081] J
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.00008]	ND [0.000081]	ND [0.000081]	ND [0.000081]	ND [0.000081]	ND [0.000081]
SW8081B	Endrin ketone	mg/kg	-	0.000078 [0.000065] J	0.00031 [0.000065] J	0.00044 [0.000065] J	ND [0.000065]	0.000073 [0.000065] J	ND [0.000065]
SW8081B	gamma-BHC	mg/kg	0.0029	0.00021 [0.000073] J	0.00021 [0.000073] J	0.0002 [0.000073] J	0.0002 [0.000073] J	0.00018 [0.000073] J	0.00018 [0.000073] J
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000081]	ND [0.000082]	ND [0.000082]	ND [0.000082]	ND [0.000082]	ND [0.000082]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000092]	ND [0.000093]	ND [0.000093]	ND [0.000093]	ND [0.000093]	ND [0.000093]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.00008]	ND [0.000081]	ND [0.000081]	ND [0.000081]	ND [0.000081]	ND [0.000081]
SW8081B	Methoxychlor	mg/kg	0.676	ND [0.000086]	0.00016 [0.000087] J	ND [0.000087]	0.00011 [0.000087] J	ND [0.000087]	ND [0.000087]
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.00079]	ND [0.0008]	ND [0.00079]	ND [0.0008]	ND [0.0008]	ND [0.0008]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00067]	ND [0.00068]	ND [0.00067]	ND [0.00068]	ND [0.00068]	ND [0.00068]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	ND [0.00059]	ND [0.00059]	<b>0.013 [0.00059]</b>	ND [0.00059]	ND [0.00059]	<b>0.0036 [0.00059] J</b>
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.0099 [0.00059]</b>	ND [0.00059]	<b>0.032 [0.00059]</b>	ND [0.00059]	ND [0.00059]	<b>0.011 [0.00059]</b>

**Notes:**

STLP = TestAmerica, Pittsburg, PA

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*E* (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				CKAG14-W CK-AG-14-Whole 18152-28 8/1/2012 TF STLP	CKAG15-F CK-AG-15-Fillet 18152-29 8/1/2012 TF STLP	CKAG15-W CK-AG-15-Whole 18152-30 8/1/2012 TF STLP	CKAG16-F CK-AG-16-Fillet 18152-31 8/5/2012 TF STLP	CKAG16-W CK-AG-16-Whole 18152-32 8/5/2012 TF STLP	CKAG17-F CK-AG-17-Fillet 18152-33 8/5/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	0.49 [0.029]	0.41 [0.029]	2.4 [0.029]	0.84 [0.03]	2 [0.029]	0.36 [0.03]
SW7471B	Mercury	mg/kg	0.014	<b>0.047 [0.01]</b>	<b>0.05 [0.011]</b>	<b>0.034 [0.011]</b>	<b>0.067 [0.011]</b>	<b>0.046 [0.011]</b>	<b>0.091 [0.01]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.00044 [0.000054]	0.000087 [0.000055] J	0.00046 [0.000055]	0.00011 [0.000055] J	0.00015 [0.000054] J	0.00011 [0.000054] J
SW8081B	4,4'-DDE	mg/kg	0.0093	0.00013 [0.000062] J	ND [0.000063]	0.00018 [0.000063] J	ND [0.000063]	0.00018 [0.000063] J	ND [0.000063]
SW8081B	4,4'-DDT	mg/kg	0.0093	ND [0.000062]	ND [0.000063]	0.000078 [0.000063] J	ND [0.000063]	ND [0.000062]	0.00031 [0.000062] J
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000074]	ND [0.000075]	ND [0.000075]	ND [0.000075]	ND [0.000074]	ND [0.000074]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000067]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000081]	ND [0.000083]	ND [0.000083]	ND [0.000083]	ND [0.000082]	ND [0.000082]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000063]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]
SW8081B	Dieldrin	mg/kg	0.0002	0.000091 [0.000069] J	ND [0.00007]	ND [0.00007]	ND [0.00007]	ND [0.000069]	ND [0.000069]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000077]	ND [0.000079]	ND [0.000079]	ND [0.000079]	ND [0.000078]	ND [0.000078]
SW8081B	Endosulfan II	mg/kg	0.811	ND [0.000073]	ND [0.000074]	ND [0.000074]	ND [0.000074]	0.00012 [0.000073] J	ND [0.000073]
SW8081B	Endosulfan sulfate	mg/kg	0.811	0.000055 [0.000043] J	ND [0.000044]	ND [0.000044]	ND [0.000044]	ND [0.000043]	ND [0.000043]
SW8081B	Endrin	mg/kg	0.041	0.00012 [0.00008] J	ND [0.000081]	0.00013 [0.000081] J	0.00011 [0.000081] J	0.000081 [0.00008] J	0.00011 [0.00008] J
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.00008]	ND [0.000081]	0.000083 [0.000081] J	ND [0.000081]	ND [0.00008]	ND [0.00008]
SW8081B	Endrin ketone	mg/kg	-	ND [0.000064]	ND [0.000065]	0.00013 [0.000065] J	ND [0.000065]	0.00011 [0.000065] J	ND [0.000065]
SW8081B	gamma-BHC	mg/kg	0.0029	0.00018 [0.000072] J	0.0002 [0.000073] J	0.0002 [0.000073] J	0.0002 [0.000073] J	0.00019 [0.000073] J	0.00018 [0.000073] J
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000081]	ND [0.000082]	ND [0.000082]	ND [0.000082]	ND [0.000081]	ND [0.000081]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000091]	ND [0.000093]	ND [0.000093]	ND [0.000093]	ND [0.000092]	ND [0.000092]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	0.00013 [0.00008] J	ND [0.000081]	0.00018 [0.000081] J	ND [0.000081]	ND [0.00008]	ND [0.00008]
SW8081B	Methoxychlor	mg/kg	0.676	ND [0.000086]	ND [0.000087]	ND [0.000087]	ND [0.000087]	ND [0.000086]	ND [0.000086]
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0027]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.0008]	ND [0.0008]	ND [0.00079]	ND [0.0008]	ND [0.00079]	ND [0.0008]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00068]	ND [0.00068]	ND [0.00067]	ND [0.00068]	ND [0.00067]	ND [0.00068]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	ND [0.00059]	ND [0.00059]	ND [0.00059]	ND [0.00059]	ND [0.00059]	<b>0.02 [0.00059]</b>
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	ND [0.00059]	ND [0.00059]	ND [0.00059]	ND [0.00059]	ND [0.00059]	<b>0.018 [0.00059]</b>

**Notes:**

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*E* (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				CKAG17-W CK-AG-17-Whole 18152-34 8/5/2012 TF STLP	CKAG18-F CK-AG-18-Fillet 18152-35 8/5/2012 TF STLP	CKAG18-W CK-AG-18-Whole 18152-36 8/5/2012 TF STLP	CKAG19-F CK-AG-19-Fillet 18152-37 8/5/2012 TF STLP	CKAG19-W CK-AG-19-Whole 18152-38 8/5/2012 TF STLP	CKAG20-F CK-AG-20-Fillet 18152-39 8/5/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	1.6 [0.03]	0.4 [0.03]	1.1 [0.03]	0.49 [0.03]	2.6 [0.029]	0.39 [0.03]
SW7471B	Mercury	mg/kg	0.014	<b>0.063 [0.0098]</b>	<b>0.095 [0.011]</b>	<b>0.058 [0.011]</b>	<b>0.073 [0.01]</b>	<b>0.05 [0.011]</b>	<b>0.13 [0.011]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.00031 [0.000055] J	0.00012 [0.000054] J	0.0002 [0.000054] J	0.00088 [0.000055] J	0.0006 [0.000055]	0.00012 [0.000055] J
SW8081B	4,4'-DDE	mg/kg	0.0093	0.00025 [0.000063] J	0.00017 [0.000063] J	0.00039 [0.000063] J	ND [0.000063]	0.00036 [0.000063] J	0.00097 [0.000063] J
SW8081B	4,4'-DDT	mg/kg	0.0093	0.00094 [0.000063] J	ND [0.000062]	0.00066 [0.000062] J	ND [0.000063]	ND [0.000063]	ND [0.000063]
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000075]	ND [0.000074]	ND [0.000074]	ND [0.000075]	ND [0.000075]	ND [0.000075]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000083]	ND [0.000082]	ND [0.000082]	ND [0.000083]	ND [0.000083]	ND [0.000083]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]
SW8081B	Dieldrin	mg/kg	0.0002	ND [0.00007]	ND [0.000069]	ND [0.000069]	ND [0.00007]	0.00012 [0.00007] J	ND [0.00007]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000079]	ND [0.000078]	ND [0.000078]	ND [0.000079]	ND [0.000079]	ND [0.000079]
SW8081B	Endosulfan II	mg/kg	0.811	ND [0.000074]	ND [0.000073]	ND [0.000073]	ND [0.000074]	ND [0.000074]	0.0001 [0.000074] J
SW8081B	Endosulfan sulfate	mg/kg	0.811	ND [0.000044]	ND [0.000043]	ND [0.000043]	ND [0.000044]	ND [0.000044]	ND [0.000044]
SW8081B	Endrin	mg/kg	0.041	0.00011 [0.000081] J	ND [0.00008]	0.00013 [0.00008] J	0.0001 [0.000081] J	ND [0.000081]	ND [0.000081]
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.000081]	ND [0.00008]	ND [0.00008]	ND [0.000081]	ND [0.000081]	ND [0.000081]
SW8081B	Endrin ketone	mg/kg	-	0.00021 [0.000065] J	0.000097 [0.000065] J	0.000075 [0.000065] J	ND [0.000065]	ND [0.000065]	ND [0.000065]
SW8081B	gamma-BHC	mg/kg	0.0029	0.00018 [0.000073] J	0.00019 [0.000073] J	0.00018 [0.000073] J	0.0002 [0.000073] J	0.00019 [0.000073] J	0.0002 [0.000073] J
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000082]	ND [0.000081]	0.00011 [0.000081] J	ND [0.000082]	ND [0.000082]	ND [0.000082]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000093]	ND [0.000092]	ND [0.000092]	ND [0.000093]	ND [0.000093]	ND [0.000093]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.000081]	ND [0.00008]	ND [0.00008]	ND [0.000081]	ND [0.000081]	ND [0.000081]
SW8081B	Methoxychlor	mg/kg	0.676	ND [0.000087]	ND [0.000086]	ND [0.000086]	ND [0.000087]	ND [0.000087]	ND [0.000087]
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.00079]	ND [0.00079]	ND [0.00079]	ND [0.00079]	ND [0.0008]	ND [0.00079]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00067]	ND [0.00067]	ND [0.00067]	ND [0.00067]	ND [0.00068]	ND [0.00067]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	ND [0.00059]	ND [0.00059]	ND [0.00059]	ND [0.00059]	ND [0.00059]	ND [0.00059]
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	ND [0.00059]	ND [0.00059]	ND [0.00059]	ND [0.00059]	<b>0.012 [0.00059]</b>	<b>0.019 [0.00059]</b>

**Notes:**

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*E (Italics)* = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				CKAG20-W CK-AG-20-Whole 18152-40 8/5/2012 TF STLP	CKAG21-F CK-AG-21-Fillet 18152-41 8/1/2012 TF STLP	Crag01-F CR-AG-01-Fillet 18147-1 8/3/2012 TF STLP	Crag01-W CR-AG-01-Whole 18147-2 8/3/2012 TF STLP	Crag02-F CR-AG-02-Fillet 18147-3 8/3/2012 TF STLP	Crag02-W CR-AG-02-Whole 18147-4 8/3/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	1.7 [0.029]	0.7 [0.03]	0.29 [0.029]	1.1 [0.03]	0.088 [0.03] J	0.97 [0.03]
SW7471B	Mercury	mg/kg	0.014	<b>0.091 [0.01]</b>	<b>0.18 [0.011]</b>	<b>0.33 [0.0099]</b>	<b>0.22 [0.011]</b>	<b>0.22 [0.01]</b>	<b>0.11 [0.011]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.00039 [0.000054] J	0.00022 [0.000055] J	0.00018 [0.000054] J	0.0006 [0.000055]	0.00012 [0.000055] J	0.00032 [0.000055] J
SW8081B	4,4'-DDE	mg/kg	0.0093	0.00027 [0.000062] J	0.00022 [0.000063] J	0.00028 [0.000063] J	0.0013 [0.000063]	0.00014 [0.000063] J	0.00075 [0.000063]
SW8081B	4,4'-DDT	mg/kg	0.0093	0.0001 [0.000062] J	ND [0.000063]	ND [0.000062]	0.000079 [0.000063] J	ND [0.000063]	ND [0.000063]
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000074]	ND [0.000075]	ND [0.000074]	ND [0.000075]	ND [0.000075]	ND [0.000075]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000067]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000081]	ND [0.000083]	ND [0.000082]	ND [0.000083]	ND [0.000083]	ND [0.000083]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000063]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]
SW8081B	Dieldrin	mg/kg	0.0002	0.00014 [0.000069] J	ND [0.00007]	ND [0.000069]	ND [0.00007]	ND [0.00007]	ND [0.00007]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000077]	ND [0.000079]	ND [0.000078]	ND [0.000079]	ND [0.000079]	ND [0.000079]
SW8081B	Endosulfan II	mg/kg	0.811	ND [0.000073]	ND [0.000074]	0.00014 [0.000073] J	0.000077 [0.000074] J	0.0001 [0.000074] J	ND [0.000074]
SW8081B	Endosulfan sulfate	mg/kg	0.811	ND [0.000043]	ND [0.000044]	ND [0.000043]	ND [0.000044]	ND [0.000044]	ND [0.000044]
SW8081B	Endrin	mg/kg	0.041	ND [0.00008]	ND [0.000081]	ND [0.00008]	0.00011 [0.000081] J	ND [0.000081]	ND [0.000081]
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.00008]	ND [0.000081]	ND [0.00008]	ND [0.000081]	ND [0.000081]	ND [0.000081]
SW8081B	Endrin ketone	mg/kg	-	0.0001 [0.000064] J	ND [0.000065]	ND [0.000065]	0.00021 [0.000065] J	ND [0.000065]	ND [0.000065]
SW8081B	gamma-BHC	mg/kg	0.0029	0.00022 [0.000072] J	0.00018 [0.000073] J	0.00011 [0.000073] J	0.00011 [0.000073] J	ND [0.000073]	ND [0.000073]
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000081]	ND [0.000082]	ND [0.000081]	ND [0.000082]	ND [0.000082]	ND [0.000082]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000091]	ND [0.000093]	ND [0.000092]	ND [0.000093]	ND [0.000093]	ND [0.000093]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.00008]	ND [0.000081]	ND [0.00008]	ND [0.000081]	ND [0.000081]	ND [0.000081]
SW8081B	Methoxychlor	mg/kg	0.676	ND [0.000086]	ND [0.000087]	ND [0.000086]	ND [0.000087]	ND [0.000087]	ND [0.000087]
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0027]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.0008]	ND [0.0008]	ND [0.00079]	ND [0.00079]	ND [0.0008]	ND [0.00079]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00068]	ND [0.00068]	ND [0.00067]	ND [0.00067]	ND [0.00068]	ND [0.00067]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	<b>0.0054 [0.00059]</b>	ND [0.00059]	ND [0.00059]	ND [0.00059]	ND [0.00059]	ND [0.00059]
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.051 [0.00059]</b>	<b>0.014 [0.00059]</b>	0.0025 [0.00059] J	<b>0.0055 [0.00059]</b>	0.0016 [0.00059] J	0.0021 [0.00059] J

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E* (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				CRAG03-F CR-AG-03-Fillet 18147-5 8/3/2012 TF STLP	CRAG03-W CR-AG-03-Whole 18147-6 8/3/2012 TF STLP	CRAG04-F CR-AG-04-Fillet 18147-7 8/3/2012 TF STLP	CRAG04-W CR-AG-04-Whole 18147-8 8/3/2012 TF STLP	CRAG05-F CR-AG-05-Fillet 18147-9 8/3/2012 TF STLP	CRAG05-W CR-AG-05-Whole 18147-10 8/3/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	0.49 [0.03]	1.9 [0.03]	0.43 [0.03]	1.1 [0.029]	0.9 [0.029]	3.8 [0.029]
SW7471B	Mercury	mg/kg	0.014	<b>0.06 [0.011]</b>	<b>0.04 [0.011]</b>	<b>0.23 [0.011]</b>	<b>0.15 [0.01]</b>	<b>0.057 [0.01]</b>	<b>0.038 [0.0098]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.00022 [0.000055] J	0.00096 [0.000055]	0.00017 [0.000055] J	0.00064 [0.000055]	0.00033 [0.000055] J	0.0011 [0.000054]
SW8081B	4,4'-DDE	mg/kg	0.0093	0.00047 [0.000063]	0.0026 [0.000063]	0.0005 [0.000063]	0.0012 [0.000063]	0.00042 [0.000063]	0.0021 [0.000063]
SW8081B	4,4'-DDT	mg/kg	0.0093	0.000073 [0.000063] J	0.00035 [0.000063] J	ND [0.000063]	0.00015 [0.000063] J	0.0001 [0.000063] J	0.00014 [0.000062] J
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000075]	ND [0.000075]	ND [0.000075]	ND [0.000075]	ND [0.000075]	ND [0.000074]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000083]	ND [0.000083]	ND [0.000083]	ND [0.000083]	ND [0.000083]	ND [0.000082]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]
SW8081B	Dieldrin	mg/kg	0.0002	ND [0.00007]	0.00012 [0.00007] J	ND [0.00007]	0.0001 [0.00007] J	ND [0.00007]	0.000078 [0.000069] J
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000079]	ND [0.000079]	ND [0.000079]	ND [0.000079]	ND [0.000079]	ND [0.000078]
SW8081B	Endosulfan II	mg/kg	0.811	0.00014 [0.000074] J	ND [0.000074]	0.00012 [0.000074] J	0.000076 [0.000074] J	0.00011 [0.000074] J	ND [0.000073]
SW8081B	Endosulfan sulfate	mg/kg	0.811	ND [0.000044]	0.000078 [0.000044] J	ND [0.000044]	ND [0.000044]	ND [0.000044]	0.000057 [0.000043] J
SW8081B	Endrin	mg/kg	0.041	0.00012 [0.000081] J	0.00013 [0.000081] J	ND [0.000081]	0.000095 [0.000081] J	ND [0.000081]	ND [0.00008]
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.000081]	ND [0.000081]	ND [0.000081]	ND [0.000081]	ND [0.000081]	ND [0.00008]
SW8081B	Endrin ketone	mg/kg	-	ND [0.000065]	ND [0.000065]	ND [0.000065]	ND [0.000065]	ND [0.000065]	0.00021 [0.000065] J
SW8081B	gamma-BHC	mg/kg	0.0029	0.0002 [0.000073] J	0.0002 [0.000073] J	0.00013 [0.000073] J	0.00019 [0.000073] J	0.0002 [0.000073] J	0.00021 [0.000073] J
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000082]	0.00011 [0.000082] J	ND [0.000082]	ND [0.000082]	ND [0.000082]	ND [0.000081]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000093]	ND [0.000093]	ND [0.000093]	ND [0.000093]	ND [0.000093]	ND [0.000092]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.000081]	ND [0.000081]	ND [0.000081]	ND [0.000081]	ND [0.000081]	ND [0.00008]
SW8081B	Methoxychlor	mg/kg	0.676	ND [0.000087]	0.00023 [0.000087] J	ND [0.000087]	0.000099 [0.000087] J	ND [0.000087]	0.00018 [0.000086] J
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.0008]	ND [0.0008]	ND [0.0008]	ND [0.0008]	ND [0.0008]	ND [0.0008]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00068]	ND [0.00068]	ND [0.00068]	ND [0.00068]	ND [0.00068]	ND [0.00068]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	ND [0.00059]	<b>0.0066 [0.00059]</b>	ND [0.00059]	ND [0.00059]	ND [0.00059]	<b>0.0083 [0.00059]</b>
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.0058 [0.00059]</b>	<b>0.008 [0.00059]</b>	0.0011 [0.00059] J	<b>0.005 [0.00059]</b>	0.0024 [0.00059] J	<b>0.019 [0.00059]</b>

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E* (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				CRAG06-F CR-AG-06-Fillet 18147-11 8/3/2012 TF STLP	CRAG06-W CR-AG-06-Whole 18147-12 8/3/2012 TF STLP	CRAG07-F CR-AG-07-Fillet 18147-13 8/3/2012 TF STLP	CRAG07-W CR-AG-07-Whole 18147-14 8/3/2012 TF STLP	CRAG08-F CR-AG-08-Fillet 18147-15 8/3/2012 TF STLP	CRAG08-W CR-AG-08-Whole 18147-16 8/3/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	0.61 [0.029]	2.1 [0.03]	1.6 [0.029]	6.6 [0.03]	0.39 [0.03]	0.95 [0.029]
SW7471B	Mercury	mg/kg	0.014	<b>0.15 [0.011]</b>	<b>0.099 [0.01]</b>	ND [0.011]	ND [0.011]	<b>0.13 [0.011]</b>	<b>0.088 [0.011]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.00023 [0.000054] J	0.00036 [0.000053] J	0.00032 [0.000055] J	0.00088 [0.000055]	0.00014 [0.000054] J	0.0004 [0.000055] J
SW8081B	4,4'-DDE	mg/kg	0.0093	0.00035 [0.000062] J	0.0013 [0.000062]	0.00047 [0.000063]	0.002 [0.000063]	0.00032 [0.000063] J	0.0015 [0.000063]
SW8081B	4,4'-DDT	mg/kg	0.0093	0.00052 [0.000062]	0.00018 [0.000061] J	ND [0.000063]	0.00017 [0.000063] J	ND [0.000062]	0.00027 [0.000063] J
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000074]	ND [0.000073]	ND [0.000075]	ND [0.000075]	ND [0.000074]	ND [0.000075]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000067]	ND [0.000067]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000081]	ND [0.000081]	ND [0.000083]	ND [0.000083]	ND [0.000082]	ND [0.000083]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000063]	ND [0.000063]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]
SW8081B	Dieldrin	mg/kg	0.0002	0.00016 [0.000069] J	ND [0.000068]	ND [0.00007]	0.000082 [0.00007] J	ND [0.000069]	ND [0.00007]
SW8081B	Endosulfan I	mg/kg	0.811	0.00012 [0.000077] J	ND [0.000077]	ND [0.000079]	ND [0.000079]	ND [0.000078]	ND [0.000079]
SW8081B	Endosulfan II	mg/kg	0.811	0.00013 [0.000073] J	ND [0.000072]	ND [0.000074]	ND [0.000074]	0.00011 [0.000073] J	0.00011 [0.000074] J
SW8081B	Endosulfan sulfate	mg/kg	0.811	0.00016 [0.000043] J	ND [0.000043]	ND [0.000044]	ND [0.000044]	ND [0.000043]	ND [0.000044]
SW8081B	Endrin	mg/kg	0.041	0.0016 [0.00008]	ND [0.000079]	ND [0.000081]	0.00022 [0.000081] J	ND [0.00008]	0.0001 [0.000081] J
SW8081B	Endrin aldehyde	mg/kg	0.041	0.00022 [0.00008] J	ND [0.000079]	ND [0.000081]	ND [0.000081]	ND [0.00008]	ND [0.000081]
SW8081B	Endrin ketone	mg/kg	-	0.00073 [0.000064]	0.000095 [0.000064] J	ND [0.000065]	0.000072 [0.000065] J	ND [0.000065]	0.00021 [0.000065] J
SW8081B	gamma-BHC	mg/kg	0.0029	0.00021 [0.000072] J	0.00026 [0.000072] J	0.0002 [0.000073] J	0.00024 [0.000073] J	0.0002 [0.000073] J	0.00014 [0.000073] J
SW8081B	gamma-Chlordane	mg/kg	0.009	0.00035 [0.000081] J	ND [0.00008]	ND [0.000082]	ND [0.000082]	ND [0.000081]	ND [0.000082]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000091]	ND [0.000091]	ND [0.000093]	ND [0.000093]	ND [0.000092]	ND [0.000093]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.00008]	ND [0.000079]	ND [0.000081]	ND [0.000081]	ND [0.00008]	ND [0.000081]
SW8081B	Methoxychlor	mg/kg	0.676	0.0011 [0.000086]	ND [0.000085]	ND [0.000087]	0.0001 [0.000087] J	ND [0.000086]	0.00011 [0.000087] J
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0027]	ND [0.0027]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00061]	ND [0.00061]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.0008]	ND [0.0008]	ND [0.00079]	ND [0.0008]	ND [0.00078]	ND [0.00078]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.0007]	ND [0.0007]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00068]	ND [0.00068]	ND [0.00067]	ND [0.00068]	ND [0.00067]	ND [0.00067]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	ND [0.00059]	<b>0.0091 [0.00059]</b>	<b>0.019 [0.00059]</b>	ND [0.00059]	<b>0.0092 [0.00058]</b>	<b>0.019 [0.00058]</b>
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.014 [0.00059]</b>	<b>0.015 [0.00059]</b>	<b>0.029 [0.00059]</b>	<b>0.017 [0.00059]</b>	<b>0.0089 [0.00058]</b>	<b>0.021 [0.00058]</b>

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E* (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				CRA09-F CR-AG-09-Fillet 18147-17 8/3/2012 TF STLP	CRA09-W CR-AG-09-Whole 18147-18 8/3/2012 TF STLP	CRA10-F CR-AG-10-Fillet 18147-19 8/3/2012 TF STLP	CRA10-W CR-AG-10-Whole 18147-20 8/3/2012 TF STLP	CRA11-F CR-AG-11-Fillet 18147-21 8/3/2012 TF STLP	CRA11-W CR-AG-11-Whole 18147-22 8/3/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	0.99 [0.03]	2.7 [0.03]	0.81 [0.03]	2.8 [0.029]	0.28 [0.029]	0.74 [0.03]
SW7471B	Mercury	mg/kg	0.014	<b>0.13 [0.01]</b>	<b>0.12 [0.011]</b>	<b>0.24 [0.011]</b>	<b>0.091 [0.011]</b>	<b>0.16 [0.01]</b>	<b>0.094 [0.01]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.00033 [0.000055] J	0.00045 [0.000054]	0.0002 [0.000055] J	0.00056 [0.000055]	0.00036 [0.000055] J	0.00047 [0.000054]
SW8081B	4,4'-DDE	mg/kg	0.0093	0.0012 [0.000063]	0.0008 [0.000063]	0.00016 [0.000063] J	0.0018 [0.000063]	0.00068 [0.000063]	0.0015 [0.000063]
SW8081B	4,4'-DDT	mg/kg	0.0093	0.00032 [0.000063] J	0.00012 [0.000062] J	ND [0.000063]	0.000069 [0.000063] J	0.00012 [0.000063] J	0.000076 [0.000062] J
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000075]	ND [0.000074]	ND [0.000075]	ND [0.000075]	ND [0.000075]	ND [0.000074]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000083]	ND [0.000082]	ND [0.000083]	ND [0.000083]	ND [0.000083]	ND [0.000082]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]
SW8081B	Dieldrin	mg/kg	0.0002	ND [0.00007]	ND [0.000069]	ND [0.00007]	ND [0.00007]	ND [0.00007]	ND [0.000069]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000079]	ND [0.000078]	ND [0.000079]	ND [0.000079]	ND [0.000079]	ND [0.000078]
SW8081B	Endosulfan II	mg/kg	0.811	0.000095 [0.000074] J	0.000075 [0.000073] J	ND [0.000074]	ND [0.000074]	ND [0.000074]	ND [0.000073]
SW8081B	Endosulfan sulfate	mg/kg	0.811	ND [0.000044]	ND [0.000043]	ND [0.000044]	0.000061 [0.000044] J	ND [0.000044]	ND [0.000043]
SW8081B	Endrin	mg/kg	0.041	0.00011 [0.000081] J	0.000085 [0.00008] J	ND [0.000081]	0.000083 [0.000081] J	0.000093 [0.000081] J	ND [0.00008]
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.000081]	ND [0.00008]	ND [0.000081]	ND [0.000081]	ND [0.000081]	ND [0.00008]
SW8081B	Endrin ketone	mg/kg	-	ND [0.000065]	0.000071 [0.000065] J	ND [0.000065]	ND [0.000065]	0.00019 [0.000065] J	0.00044 [0.000065]
SW8081B	gamma-BHC	mg/kg	0.0029	0.00024 [0.000073] J	ND [0.000073]	ND [0.000073]	ND [0.000073]	ND [0.000073]	ND [0.000073]
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000082]	ND [0.000081]	ND [0.000082]	ND [0.000082]	ND [0.000082]	ND [0.000081]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000093]	ND [0.000092]	ND [0.000093]	ND [0.000093]	ND [0.000093]	ND [0.000092]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.000081]	ND [0.00008]	ND [0.000081]	ND [0.000081]	ND [0.000081]	ND [0.00008]
SW8081B	Methoxychlor	mg/kg	0.676	0.000096 [0.000087] J	0.00011 [0.000086] J	ND [0.000087]	0.0002 [0.000087] J	0.00022 [0.000087] J	ND [0.000086]
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00061]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.00078]	ND [0.0008]	ND [0.00079]	ND [0.0008]	ND [0.0008]	ND [0.0008]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.0007]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00067]	ND [0.00068]	ND [0.00067]	ND [0.00068]	ND [0.00068]	ND [0.00068]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	ND [0.00058]	ND [0.00059]	ND [0.00059]	ND [0.00059]	ND [0.00059]	ND [0.00059]
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.013 [0.00058]</b>	0.0023 [0.00059] J	0.0011 [0.00059] J	<b>0.0028 [0.00059] J</b>	0.0023 [0.00059] J	<b>0.0042 [0.00059]</b>

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E* (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				CRAG12-F CR-AG-12-Fillet 18147-23 8/3/2012 TF STLP	CRAG12-W CR-AG-12-Whole 18147-24 8/3/2012 TF STLP	CRAG13-F CR-AG-13-Fillet 18147-25 8/6/2012 TF STLP	CRAG13-W CR-AG-13-Whole 18147-26 8/6/2012 TF STLP	CRAG14-F CR-AG-14-Fillet 18147-27 8/6/2012 TF STLP	CRAG14-W CR-AG-14-Whole 18147-28 8/6/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	0.18 [0.029]	0.63 [0.03]	0.59 [0.03]	1.4 [0.029]	0.6 [0.03]	1.6 [0.029]
SW7471B	Mercury	mg/kg	0.014	<b>0.22 [0.0098]</b>	<b>0.15 [0.0099]</b>	<b>0.17 [0.011]</b>	<b>0.12 [0.011]</b>	<b>0.2 [0.01]</b>	<b>0.13 [0.01]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.00023 [0.000055] J	0.00031 [0.000055] J	0.00028 [0.000054] J	0.00061 [0.000055]	0.00025 [0.000054] J	0.00026 [0.000055] J
SW8081B	4,4'-DDE	mg/kg	0.0093	0.00028 [0.000063] J	0.0015 [0.000063]	0.00055 [0.000063]	0.0018 [0.000063]	0.00053 [0.000063]	0.001 [0.000063]
SW8081B	4,4'-DDT	mg/kg	0.0093	0.00012 [0.000063] J	0.00011 [0.000063] J	0.00021 [0.000062] J	0.0047 [0.000063]	0.00011 [0.000062] J	0.00022 [0.000063] J
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000075]	ND [0.000075]	ND [0.000074]	ND [0.000075]	ND [0.000074]	ND [0.000075]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000083]	ND [0.000083]	ND [0.000082]	ND [0.000083]	ND [0.000082]	ND [0.000083]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]
SW8081B	Dieldrin	mg/kg	0.0002	ND [0.00007]	ND [0.00007]	ND [0.000069]	<b>0.0003 [0.00007] J</b>	ND [0.000069]	ND [0.00007]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000079]	ND [0.000079]	ND [0.000078]	0.00012 [0.000079] J	ND [0.000078]	ND [0.000079]
SW8081B	Endosulfan II	mg/kg	0.811	0.000089 [0.000074] J	ND [0.000074]	ND [0.000073]	0.000098 [0.000074] J	ND [0.000073]	ND [0.000074]
SW8081B	Endosulfan sulfate	mg/kg	0.811	ND [0.000044]	ND [0.000044]	ND [0.000043]	0.00033 [0.000044] J	ND [0.000043]	ND [0.000044]
SW8081B	Endrin	mg/kg	0.041	0.000091 [0.000081] J	0.00009 [0.000081] J	ND [0.00008]	0.0018 [0.000081]	ND [0.00008]	ND [0.000081]
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.000081]	ND [0.000081]	ND [0.00008]	0.000093 [0.000081] J	ND [0.00008]	ND [0.000081]
SW8081B	Endrin ketone	mg/kg	-	ND [0.000065]	0.00027 [0.000065] J	ND [0.000065]	0.0018 [0.000065]	0.000097 [0.000065] J	0.00014 [0.000065] J
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.000073]	ND [0.000073]	ND [0.000073]	ND [0.000073]	ND [0.000073]	ND [0.000073]
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000082]	ND [0.000082]	ND [0.000081]	0.00077 [0.000082]	ND [0.000081]	ND [0.000082]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000093]	ND [0.000093]	ND [0.000092]	ND [0.000093]	ND [0.000092]	ND [0.000093]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.000081]	ND [0.000081]	ND [0.00008]	0.00013 [0.000081] J	ND [0.00008]	ND [0.000081]
SW8081B	Methoxychlor	mg/kg	0.676	0.00011 [0.000087] J	0.00022 [0.000087] J	0.00018 [0.000086] J	0.002 [0.000087]	ND [0.000086]	ND [0.000087]
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.00079]	ND [0.0008]	ND [0.00079]	ND [0.0008]	ND [0.0008]	ND [0.00079]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00067]	ND [0.00068]	ND [0.00067]	ND [0.00068]	ND [0.00068]	ND [0.00067]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	ND [0.00059]	ND [0.00059]	<b>0.0019 [0.00059] J</b>	ND [0.00059]	<b>0.0026 [0.00059] J</b>	<b>0.0019 [0.00059] J</b>
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	0.0018 [0.00059] J	<b>0.0051 [0.00059]</b>	<b>0.0042 [0.00059]</b>	<b>0.0071 [0.00059]</b>	<b>0.0042 [0.00059]</b>	<b>0.0027 [0.00059] J</b>

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E* (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).



**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				CRAG15-F CR-AG-15-Fillet 18147-29 8/6/2012 TF STLP	CRAG15-W CR-AG-15-Whole 18147-30 8/6/2012 TF STLP	CRAG16-F CR-AG-16-Fillet 18147-31 8/6/2012 TF STLP	CRAG16-W CR-AG-16-Whole 18147-32 8/6/2012 TF STLP	CRAG17-F CR-AG-17-Fillet 18147-33 8/6/2012 TF STLP	CRAG17-W CR-AG-17-Whole 18147-34 8/6/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	0.36 [0.029]	1.3 [0.03]	0.87 [0.03]	2.3 [0.029]	0.16 [0.029]	0.46 [0.03]
SW7471B	Mercury	mg/kg	0.014	<b>0.48 [0.01]</b>	<b>0.23 [0.01]</b>	<b>0.067 [0.011]</b>	<b>0.032 [0.011] J</b>	<b>0.33 [0.011]</b>	<b>0.21 [0.011]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.00011 [0.000055] J	0.00029 [0.000055] J	0.00022 [0.000054] J	0.00038 [0.000055] J	0.00016 [0.000054] J	0.00031 [0.000055] J
SW8081B	4,4'-DDE	mg/kg	0.0093	0.00015 [0.000063] J	0.00055 [0.000063]	0.00027 [0.000063] J	0.00086 [0.000063]	0.00038 [0.000063] J	0.00046 [0.000063]
SW8081B	4,4'-DDT	mg/kg	0.0093	ND [0.000063]	ND [0.000063]	ND [0.000062]	0.00021 [0.000063] J	0.000096 [0.000062] J	0.00014 [0.000063] J
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000075]	ND [0.000075]	ND [0.000074]	ND [0.000075]	ND [0.000074]	ND [0.000075]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000083]	ND [0.000083]	ND [0.000082]	ND [0.000083]	ND [0.000082]	ND [0.000083]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]
SW8081B	Dieldrin	mg/kg	0.0002	ND [0.00007]	ND [0.00007]	ND [0.000069]	ND [0.00007]	ND [0.000069]	ND [0.00007]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000079]	ND [0.000079]	ND [0.000078]	ND [0.000079]	ND [0.000078]	ND [0.000079]
SW8081B	Endosulfan II	mg/kg	0.811	0.000089 [0.000074] J	0.000074 [0.000074] J	ND [0.000073]	0.00011 [0.000074] J	ND [0.000073]	ND [0.000074]
SW8081B	Endosulfan sulfate	mg/kg	0.811	ND [0.000044]	ND [0.000044]	ND [0.000043]	ND [0.000044]	ND [0.000043]	ND [0.000044]
SW8081B	Endrin	mg/kg	0.041	0.000087 [0.000081] J	0.00015 [0.000081] J	ND [0.00008]	ND [0.000081]	ND [0.00008]	0.00013 [0.000081] J
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.000081]	ND [0.000081]	ND [0.00008]	ND [0.000081]	ND [0.00008]	ND [0.000081]
SW8081B	Endrin ketone	mg/kg	-	ND [0.000065]	ND [0.000065]	ND [0.000065]	0.000083 [0.000065] J	ND [0.000065]	0.00024 [0.000065] J
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.000073]	ND [0.000073]	ND [0.000073]	ND [0.000073]	ND [0.000073]	ND [0.000073]
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000082]	ND [0.000082]	ND [0.000081]	ND [0.000082]	ND [0.000081]	ND [0.000082]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000093]	ND [0.000093]	ND [0.000092]	ND [0.000093]	ND [0.000092]	ND [0.000093]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.000081]	ND [0.000081]	ND [0.00008]	ND [0.000081]	ND [0.00008]	ND [0.000081]
SW8081B	Methoxychlor	mg/kg	0.676	ND [0.000087]	ND [0.000087]	ND [0.000086]	0.0002 [0.000087] J	ND [0.000086]	ND [0.000087]
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.00079]	ND [0.00079]	ND [0.0008]	ND [0.00079]	ND [0.0008]	ND [0.0008]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00067]	ND [0.00067]	ND [0.00068]	ND [0.00067]	ND [0.00068]	ND [0.00068]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	<b>0.002 [0.00059] J</b>	<b>0.021 [0.00059]</b>	<b>0.016 [0.00059]</b>	<b>0.0086 [0.00059]</b>	ND [0.00059]	ND [0.00059]
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	0.0024 [0.00059] J	<b>0.016 [0.00059]</b>	<b>0.018 [0.00059]</b>	<b>0.008 [0.00059]</b>	0.00082 [0.00059] J	ND [0.00059]

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E* (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				CRAG18-F CR-AG-18-Fillet 18147-35 8/6/2012 TF STLP	CRAG18-W CR-AG-18-Whole 18147-36 8/6/2012 TF STLP	CRAG19-F CR-AG-19-Fillet 18147-37 8/6/2012 TF STLP	CRAG19-W CR-AG-19-Whole 18147-38 8/6/2012 TF STLP	CRAG20-F CR-AG-20-Fillet 18147-39 8/6/2012 TF STLP	CRAG20-W CR-AG-20-Whole 18147-40 8/6/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	0.6 [0.03]	1.2 [0.03]	0.23 [0.029]	1.2 [0.029]	0.48 [0.029]	1.5 [0.03]
SW7471B	Mercury	mg/kg	0.014	<b>0.39 [0.011]</b>	<b>0.25 [0.011]</b>	<b>0.15 [0.011]</b>	<b>0.099 [0.01]</b>	<b>0.048 [0.011]</b>	<b>0.035 [0.011]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.00027 [0.000055] J	0.00036 [0.000055] J	0.00018 [0.000054] J	0.00022 [0.000055] J	0.00037 [0.000054] J	0.00098 [0.000053]
SW8081B	4,4'-DDE	mg/kg	0.0093	0.00051 [0.000063]	0.00092 [0.000063]	0.00012 [0.000062] J	0.00043 [0.000063]	0.0011 [0.000062]	0.0029 [0.000062]
SW8081B	4,4'-DDT	mg/kg	0.0093	0.00014 [0.000063] J	0.000089 [0.000063] J	ND [0.000062]	0.000071 [0.000063] J	0.00024 [0.000062] J	0.0003 [0.000061] J
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000075]	ND [0.000075]	ND [0.000074]	ND [0.000075]	ND [0.000074]	ND [0.000073]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000068]	ND [0.000068]	ND [0.000067]	ND [0.000068]	ND [0.000067]	ND [0.000067]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000083]	ND [0.000083]	ND [0.000081]	ND [0.000083]	ND [0.000081]	ND [0.000081]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000064]	ND [0.000064]	ND [0.000063]	ND [0.000064]	ND [0.000063]	ND [0.000063]
SW8081B	Dieldrin	mg/kg	0.0002	ND [0.00007]	ND [0.00007]	ND [0.000069]	ND [0.00007]	ND [0.000069]	ND [0.000068]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000079]	ND [0.000079]	ND [0.000077]	ND [0.000079]	ND [0.000077]	ND [0.000077]
SW8081B	Endosulfan II	mg/kg	0.811	ND [0.000074]	ND [0.000074]	ND [0.000073]	ND [0.000074]	ND [0.000073]	ND [0.000072]
SW8081B	Endosulfan sulfate	mg/kg	0.811	ND [0.000044]	ND [0.000044]	ND [0.000043]	0.000048 [0.000044] J	ND [0.000043]	0.00013 [0.000043] J
SW8081B	Endrin	mg/kg	0.041	0.00011 [0.000081] J	0.00011 [0.000081] J	ND [0.00008]	ND [0.000081]	ND [0.00008]	ND [0.000079]
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.000081]	ND [0.000081]	ND [0.00008]	ND [0.000081]	ND [0.00008]	ND [0.000079]
SW8081B	Endrin ketone	mg/kg	-	ND [0.000065]	ND [0.000065]	ND [0.000064]	ND [0.000065]	0.00027 [0.000064] J	0.0004 [0.000064] J
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.000073]	ND [0.000073]	ND [0.000072]	ND [0.000073]	ND [0.000072]	ND [0.000072]
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000082]	ND [0.000082]	ND [0.000081]	ND [0.000082]	ND [0.000081]	ND [0.00008]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000093]	ND [0.000093]	ND [0.000091]	ND [0.000093]	ND [0.000091]	ND [0.000091]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.000081]	ND [0.000081]	ND [0.00008]	ND [0.000081]	ND [0.00008]	ND [0.000079]
SW8081B	Methoxychlor	mg/kg	0.676	ND [0.000087]	ND [0.000087]	ND [0.000086]	ND [0.000087]	0.00015 [0.000086] J, B	0.0003 [0.000085] J
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0028]	ND [0.0028]	ND [0.0027]	ND [0.0028]	ND [0.0027]	ND [0.0027]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.0008]	ND [0.0008]	ND [0.0008]	ND [0.00079]	ND [0.00079]	ND [0.0008]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00068]	ND [0.00068]	ND [0.00068]	ND [0.00067]	ND [0.00067]	ND [0.00068]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	<b>0.0032 [0.00059] J</b>	ND [0.00059]	<b>0.039 [0.00059]</b>	ND [0.00059]	ND [0.00059]	ND [0.00059]
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.006 [0.00059]</b>	<b>0.0027 [0.00059] J</b>	ND [0.00059]	0.0019 [0.00059] J	<b>0.0034 [0.00059] J</b>	<b>0.0088 [0.00059]</b>

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E* (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				CRAG21-F CR-AG-21-Fillet 18147-41 8/3/2012 TF STLP	GSAG01-F GS-AG-01-FILLET 13438-1 7/31/2012 TF STLP	GSAG01-W GS-AG-01-WHOLE 13438-2 7/31/2012 TF STLP	GSAG02-G GS-AG-02-FILLET 13438-3 7/31/2012 TF STLP	GSAG02-W GS-AG-02-WHOLE 13438-4 7/31/2012 TF STLP	GSAG03-F GS-AG-03-FILLET 13438-5 7/31/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	0.51 [0.029]	0.68 [0.03]	2.9 [0.029]	1 [0.03]	4.3 [0.03]	1.2 [0.029]
SW7471B	Mercury	mg/kg	0.014	<b>0.22 [0.01]</b>	<b>0.03 [0.011] J</b>	<b>0.024 [0.01] J</b>	<b>0.039 [0.011]</b>	<b>0.022 [0.01] J</b>	<b>0.038 [0.01]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.000099 [0.000055] J	<b>0.056 [0.00053]</b>	<b>0.27 [0.00053]</b>	<b>0.066 [0.00053]</b>	<b>0.3 [0.00054]</b>	<b>0.21 [0.00055]</b>
SW8081B	4,4'-DDE	mg/kg	0.0093	0.00021 [0.000063] J	<b>0.021 [0.00062]</b>	<b>0.1 [0.00061]</b>	<b>0.013 [0.00061]</b>	<b>0.08 [0.00062]</b>	<b>0.078 [0.00063]</b>
SW8081B	4,4'-DDT	mg/kg	0.0093	ND [0.000063]	0.0016 [0.00061] J	0.0048 [0.00061]	0.0015 [0.00061] J	0.0047 [0.00062]	0.0069 [0.00063]
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000075]	ND [0.00073]	ND [0.00073]	ND [0.00073]	ND [0.00074]	ND [0.00075]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000068]	ND [0.00067]	ND [0.00066]	ND [0.00066]	ND [0.00067]	ND [0.00068]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000083]	ND [0.00081]	ND [0.0008]	ND [0.0008]	0.00095 [0.00081] J	ND [0.00083]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.0011]	ND [0.0011]	ND [0.0011]	ND [0.0011]	ND [0.0011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000064]	ND [0.00063]	ND [0.00062]	ND [0.00062]	ND [0.00063]	ND [0.00064]
SW8081B	Dieldrin	mg/kg	0.0002	ND [0.00007]	ND [0.00068]	<b>0.0017 [0.00068] J</b>	ND [0.00068]	ND [0.00069]	ND [0.0007]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000079]	ND [0.00077]	ND [0.00076]	ND [0.00076]	ND [0.00077]	ND [0.00079]
SW8081B	Endosulfan II	mg/kg	0.811	ND [0.000074]	ND [0.00072]	0.0015 [0.00072] J	ND [0.00072]	0.0021 [0.00073] J	0.0012 [0.00074] J
SW8081B	Endosulfan sulfate	mg/kg	0.811	ND [0.000044]	0.00063 [0.00043] J	0.0022 [0.00042] J	0.00044 [0.00042] J	0.0018 [0.00043] J	0.0015 [0.00044] J
SW8081B	Endrin	mg/kg	0.041	ND [0.000081]	0.0018 [0.00079] J	0.0062 [0.00079]	0.0012 [0.00079] J	0.0032 [0.0008] J	0.0037 [0.00081] J
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.000081]	ND [0.00079]	ND [0.00079]	ND [0.00079]	ND [0.0008]	ND [0.00081]
SW8081B	Endrin ketone	mg/kg	-	ND [0.000065]	0.0044 [0.00064]	0.013 [0.00063]	0.0057 [0.00063]	0.053 [0.00064]	0.0096 [0.00065]
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.000073]	ND [0.00072]	ND [0.00071]	ND [0.00071]	ND [0.00072]	ND [0.00073]
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000082]	ND [0.0008]	ND [0.0008]	ND [0.0008]	ND [0.00081]	ND [0.00082]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000093]	ND [0.00091]	ND [0.0009]	ND [0.0009]	ND [0.00091]	ND [0.00093]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.000081]	ND [0.00079]	ND [0.00079]	<b>0.0011 [0.00079] J</b>	<b>0.0051 [0.0008]</b>	<b>0.0012 [0.00081] J</b>
SW8081B	Methoxychlor	mg/kg	0.676	ND [0.000087]	0.0016 [0.00085] J	0.0069 [0.00085] J	0.0014 [0.00085] J	0.0059 [0.00086] J	0.0055 [0.00087] J
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0028]	ND [0.027]	ND [0.027]	ND [0.027]	ND [0.027]	ND [0.028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00061]	ND [0.00062]	ND [0.00061]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.00078]	ND [0.00079]	ND [0.00078]	ND [0.00079]	ND [0.0008]	ND [0.0008]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.0007]	ND [0.00071]	ND [0.0007]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00067]	ND [0.00067]	ND [0.00067]	ND [0.00067]	ND [0.00068]	ND [0.00068]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	ND [0.00058]	<b>0.026 [0.00059]</b>	<b>0.1 [0.00058]</b>	<b>0.03 [0.00059]</b>	<b>0.12 [0.00059]</b>	<b>0.089 [0.00059]</b>
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.0033 [0.00058] J</b>	<b>0.086 [0.00059]</b>	<b>0.27 [0.00058]</b>	<b>0.07 [0.00059]</b>	<b>0.21 [0.00059]</b>	<b>0.18 [0.00059]</b>

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

E (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				GSAG03-W GS-AG-03-WHOLE 13438-6 7/31/2012 TF STLP	GSAG04-F GS-AG-04-FILLET 13438-7 7/31/2012 TF STLP	GSAG04-W GS-AG-04-WHOLE 13438-8 7/31/2012 TF STLP	GSAG05-F GS-AG-05-FILLET 13438-9 7/31/2012 TF STLP	GSAG05-W GS-AG-05-WHOLE 13438-10 7/31/2012 TF STLP	GSAG06-F GS-AG-06-FILLET 13438-11 7/31/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	3.1 [0.029]	0.94 [0.03]	3 [0.03]	0.49 [0.03]	3.1 [0.029]	1.2 [0.029]
SW7471B	Mercury	mg/kg	0.014	<b>0.027 [0.011] J</b>	<b>0.025 [0.01] J</b>	<b>0.02 [0.011] J</b>	<b>0.032 [0.011]</b>	0.011 [0.011] J	<b>0.018 [0.0098] J</b>
SW8081B	4,4'-DDD	mg/kg	0.013	<b>0.43 [0.00053]</b>	<b>0.18 [0.00053]</b>	<b>0.3 [0.00054]</b>	<b>0.031 [0.00053]</b>	<b>0.22 [0.00053]</b>	<b>0.092 [0.00054]</b>
SW8081B	4,4'-DDE	mg/kg	0.0093	<b>0.15 [0.00062]</b>	<b>0.069 [0.00061]</b>	<b>0.11 [0.00062]</b>	0.0047 [0.00061]	<b>0.052 [0.00061]</b>	<b>0.039 [0.00063]</b>
SW8081B	4,4'-DDT	mg/kg	0.0093	<b>0.013 [0.00061]</b>	0.0057 [0.00061]	0.0073 [0.00062]	0.0007 [0.00061] J	0.0038 [0.00061] J	0.0031 [0.00062] J
SW8081B	Aldrin	mg/kg	0.00019	ND [0.00073]	ND [0.00073]	ND [0.00074]	ND [0.00073]	ND [0.00073]	ND [0.00074]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.00067]	ND [0.00066]	ND [0.00067]	ND [0.00066]	ND [0.00066]	ND [0.00068]
SW8081B	alpha-Chlordane	mg/kg	0.009	0.00098 [0.00081] J	ND [0.0008]	ND [0.00081]	ND [0.0008]	ND [0.0008]	ND [0.00082]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.0011]	ND [0.0011]	ND [0.0011]	ND [0.0011]	ND [0.0011]	ND [0.0011]
SW8081B	delta-BHC	mg/kg	-	ND [0.00063]	ND [0.00062]	ND [0.00063]	ND [0.00062]	ND [0.00062]	ND [0.00064]
SW8081B	Dieldrin	mg/kg	0.0002	<b>0.0019 [0.00068] J</b>	ND [0.00068]	<b>0.0016 [0.00069] J</b>	ND [0.00068]	ND [0.00068]	ND [0.00069]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.00077]	ND [0.00076]	ND [0.00077]	ND [0.00076]	ND [0.00076]	ND [0.00078]
SW8081B	Endosulfan II	mg/kg	0.811	0.0024 [0.00072] J	0.0014 [0.00072] J	0.0016 [0.00073] J	ND [0.00072]	0.00097 [0.00072] J	0.0011 [0.00073] J
SW8081B	Endosulfan sulfate	mg/kg	0.811	0.0028 [0.00043] J	0.0015 [0.00042] J	0.003 [0.00043] J	ND [0.00042]	0.00098 [0.00042] J	0.00091 [0.00043] J
SW8081B	Endrin	mg/kg	0.041	0.0069 [0.00079]	0.0038 [0.00079] J	0.0057 [0.0008]	ND [0.00079]	0.0019 [0.00079] J	0.0023 [0.0008] J
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.00079]	ND [0.00079]	ND [0.0008]	ND [0.00079]	ND [0.00079]	ND [0.0008]
SW8081B	Endrin ketone	mg/kg	-	0.017 [0.00064]	0.017 [0.00063]	0.014 [0.00064]	0.0022 [0.00063] J	0.0059 [0.00063]	0.0068 [0.00065]
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.00072]	ND [0.00071]	ND [0.00072]	ND [0.00071]	ND [0.00071]	ND [0.00073]
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.0008]	ND [0.0008]	ND [0.00081]	ND [0.0008]	ND [0.0008]	ND [0.00081]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.00091]	ND [0.0009]	ND [0.00091]	ND [0.0009]	ND [0.0009]	ND [0.00092]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.00079]	ND [0.00079]	ND [0.0008]	ND [0.00079]	ND [0.00079]	ND [0.0008]
SW8081B	Methoxychlor	mg/kg	0.676	0.0088 [0.00085]	0.0053 [0.00085] J	0.0076 [0.00086] J	ND [0.00085]	ND [0.00085]	0.0035 [0.00086] J
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.027]	ND [0.027]	ND [0.027]	ND [0.027]	ND [0.027]	ND [0.028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.0008]	ND [0.00079]	ND [0.0008]	ND [0.0008]	ND [0.0008]	ND [0.00079]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00068]	ND [0.00067]	ND [0.00068]	ND [0.00068]	ND [0.00068]	ND [0.00067]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	<b>0.13 [0.00059]</b>	<b>0.054 [0.00059]</b>	<b>0.13 [0.00059]</b>	<b>0.15 [0.00059]</b>	<b>0.11 [0.00059]</b>	<b>0.095 [0.00059]</b>
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.45 [0.00059]</b>	<b>0.13 [0.00059]</b>	<b>0.26 [0.00059]</b>	<b>0.13 [0.00059]</b>	<b>0.2 [0.00059]</b>	<b>0.21 [0.00059]</b>

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E* (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

				Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory	GSAG06-W GS-AG-06-WHOLE 13438-12 7/31/2012 TF STLP	GSAG07-F GS-AG-07-FILLET 13438-13 8/2/2012 TF STLP	GSAG07-W GS-AG-07-WHOLE 13438-14 8/2/2012 TF STLP	GSAG08-F GS-AG-08-FILLET 13438-15 8/2/2012 TF STLP	GSAG08-W GS-AG-08-WHOLE 13438-16 8/2/2012 TF STLP	GSAG09-F GS-AG-09-FILLET 13438-17 8/2/2012 TF STLP
Method	Analyte	Units	Project Action Limit							
BDTL	Total Lipids	Percent	-	2 [0.029]	0.7 [0.03]	2.7 [0.03]	0.74 [0.03]	3.2 [0.03]	0.76 [0.03]	
SW7471B	Mercury	mg/kg	0.014	<b>0.023 [0.011] J</b>	<b>0.033 [0.011]</b>	0.013 [0.011] J	<b>0.059 [0.01]</b>	<b>0.037 [0.011]</b>	<b>0.026 [0.011] J</b>	
SW8081B	4,4'-DDD	mg/kg	0.013	<b>0.25 [0.00053]</b>	<b>0.053 [0.00054]</b>	<b>0.24 [0.00054]</b>	<b>0.091 [0.00053]</b>	<b>0.37 [0.00053]</b>	<b>0.056 [0.00055]</b>	
SW8081B	4,4'-DDE	mg/kg	0.0093	<b>0.11 [0.00062]</b>	<b>0.01 [0.00062]</b>	<b>0.073 [0.00062]</b>	<b>0.036 [0.00062]</b>	<b>0.14 [0.00062]</b>	<b>0.02 [0.00063]</b>	
SW8081B	4,4'-DDT	mg/kg	0.0093	<b>0.031 [0.00061]</b>	0.0011 [0.00062] J	0.003 [0.00062] J	0.0025 [0.00061] J	0.0086 [0.00061]	0.0017 [0.00063] J	
SW8081B	Aldrin	mg/kg	0.00019	ND [0.00073]	ND [0.00074]	ND [0.00074]	ND [0.00073]	ND [0.00073]	ND [0.00075]	
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.00067]	ND [0.00067]	ND [0.00067]	ND [0.00067]	ND [0.00067]	ND [0.00068]	
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.00081]	ND [0.00081]	0.0015 [0.00081] J	ND [0.00081]	0.00086 [0.00081] J	ND [0.00083]	
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.0011]	ND [0.0011]	ND [0.0011]	ND [0.0011]	ND [0.0011]	ND [0.0011]	
SW8081B	delta-BHC	mg/kg	-	ND [0.00063]	ND [0.00063]	ND [0.00063]	ND [0.00063]	ND [0.00063]	ND [0.00064]	
SW8081B	Dieldrin	mg/kg	0.0002	<b>0.0016 [0.00068] J</b>	ND [0.00069]	ND [0.00069]	ND [0.00068]	<b>0.0017 [0.00068] J</b>	ND [0.0007]	
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.00077]	ND [0.00077]	ND [0.00077]	ND [0.00077]	ND [0.00077]	ND [0.00079]	
SW8081B	Endosulfan II	mg/kg	0.811	0.0015 [0.00072] J	ND [0.00073]	0.00097 [0.00073] J	ND [0.00072]	0.0017 [0.00072] J	ND [0.00074]	
SW8081B	Endosulfan sulfate	mg/kg	0.811	0.0027 [0.00043] J	0.00046 [0.00043] J	0.0013 [0.00043] J	0.0012 [0.00043] J	0.0028 [0.00043] J	0.00054 [0.00044] J	
SW8081B	Endrin	mg/kg	0.041	0.0044 [0.00079]	0.0011 [0.0008] J	0.003 [0.0008] J	0.0021 [0.00079] J	0.0066 [0.00079]	0.0012 [0.00081] J	
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.00079]	ND [0.0008]	ND [0.0008]	ND [0.00079]	ND [0.00079]	ND [0.00081]	
SW8081B	Endrin ketone	mg/kg	-	0.018 [0.00064]	0.0027 [0.00064] J	0.0048 [0.00064]	0.0041 [0.00064]	0.014 [0.00064]	0.0029 [0.00065] J	
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.00072]	ND [0.00072]	ND [0.00072]	ND [0.00072]	ND [0.00072]	ND [0.00073]	
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.0008]	ND [0.00081]	ND [0.00081]	ND [0.0008]	ND [0.0008]	ND [0.00082]	
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.00091]	ND [0.00091]	ND [0.00091]	ND [0.00091]	ND [0.00091]	ND [0.00093]	
SW8081B	Heptachlor epoxide	mg/kg	0.00035	<b>0.0024 [0.00079] J</b>	ND [0.0008]	<b>0.0034 [0.0008] J</b>	<b>0.00094 [0.00079] J</b>	<b>0.0045 [0.00079]</b>	ND [0.00081]	
SW8081B	Methoxychlor	mg/kg	0.676	ND [0.00085]	ND [0.00086]	0.0027 [0.00086] J	0.0025 [0.00085] J	0.0083 [0.00085]	0.0015 [0.00087] J	
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.027]	ND [0.027]	ND [0.027]	ND [0.027]	ND [0.027]	ND [0.028]	
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.0008]	ND [0.0008]	ND [0.0008]	ND [0.00079]	ND [0.0008]	ND [0.00079]	
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00068]	ND [0.00068]	ND [0.00068]	ND [0.00067]	ND [0.00068]	ND [0.00067]	
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	<b>0.2 [0.00059]</b>	<b>0.061 [0.00059]</b>	<b>0.1 [0.00059]</b>	<b>0.045 [0.00059]</b>	<b>0.15 [0.00059]</b>	<b>0.046 [0.00059]</b>	
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.36 [0.00059]</b>	<b>0.067 [0.00059]</b>	<b>0.15 [0.00059]</b>	<b>0.091 [0.00059]</b>	<b>0.43 [0.00059]</b>	<b>0.16 [0.00059]</b>	

**Notes:**

STLP = TestAmerica, Pittsburg, PA

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*E (Italics)* = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				GSAG09-W GS-AG-09-WHOLE 13438-18 8/2/2012 TF STLP	GSAG10-F GS-AG-10-FILLET 13438-19 8/2/2012 TF STLP	GSAG10-W GS-AG-10-WHOLE 13438-20 8/2/2012 TF STLP	GSAG11-F GS-AG-11-FILLET 13438-21 8/2/2012 TF STLP	GSAG11-W GS-AG-11-WHOLE 13438-22 8/2/2012 TF STLP	GSAG12-F GS-AG-12-FILLET 13438-23 8/2/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	4.2 [0.03]	0.72 [0.029]	3.2 [0.03]	2 [0.03]	6 [0.029]	1.2 [0.03]
SW7471B	Mercury	mg/kg	0.014	<b>0.022 [0.0099] J</b>	<b>0.031 [0.011] J</b>	0.013 [0.011] J	<b>0.034 [0.011]</b>	<b>0.014 [0.01] J</b>	<b>0.047 [0.011]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	<b>0.29 [0.0054]</b>	<b>0.12 [0.0054]</b>	<b>0.52 [0.0054]</b>	<b>0.16 [0.0055]</b>	<b>1.5 [0.0027]</b>	<b>0.13 [0.0054]</b>
SW8081B	4,4'-DDE	mg/kg	0.0093	<b>0.1 [0.00063]</b>	<b>0.041 [0.00063]</b>	<b>0.16 [0.00062]</b>	<b>0.055 [0.00063]</b>	<b>0.51 [0.0031]</b>	<b>0.049 [0.00063]</b>
SW8081B	4,4'-DDT	mg/kg	0.0093	0.0053 [0.00062]	0.0038 [0.00062] J	<b>0.012 [0.00062]</b>	0.0046 [0.00063]	<b>0.044 [0.0031]</b>	0.0035 [0.00062] J
SW8081B	Aldrin	mg/kg	0.00019	ND [0.00074]	ND [0.00074]	ND [0.00074]	ND [0.00075]	ND [0.0037]	ND [0.00074]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.00068]	ND [0.00068]	ND [0.00067]	ND [0.00068]	ND [0.0034]	ND [0.00068]
SW8081B	alpha-Chlordane	mg/kg	0.009	0.00084 [0.00082] J	ND [0.00082]	0.00082 [0.00081] J	ND [0.00083]	0.0087 [0.0041] J	ND [0.00082]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.0011]	ND [0.0011]	ND [0.0011]	ND [0.0011]	ND [0.0054]	ND [0.0011]
SW8081B	delta-BHC	mg/kg	-	ND [0.00064]	ND [0.00064]	ND [0.00063]	ND [0.00064]	ND [0.0032]	ND [0.00064]
SW8081B	Dieldrin	mg/kg	0.0002	<b>0.0018 [0.00069] J</b>	ND [0.00069]	<b>0.0022 [0.00069] J</b>	<b>0.003 [0.0007] J</b>	<b>0.011 [0.0035] J</b>	<b>0.0025 [0.00069] J</b>
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.00078]	ND [0.00078]	ND [0.00077]	ND [0.00079]	ND [0.0039]	ND [0.00078]
SW8081B	Endosulfan II	mg/kg	0.811	0.0018 [0.00073] J	0.0008 [0.00073] J	0.0023 [0.00073] J	0.0012 [0.00074] J	0.0069 [0.0037] J	ND [0.00073]
SW8081B	Endosulfan sulfate	mg/kg	0.811	0.0023 [0.00043] J	0.00081 [0.00043] J	0.0029 [0.00043] J	0.0011 [0.00044] J	0.011 [0.0022] J	0.00092 [0.00043] J
SW8081B	Endrin	mg/kg	0.041	0.0037 [0.0008] J	0.0017 [0.0008] J	0.0061 [0.0008]	0.0028 [0.00081] J	0.024 [0.004]	0.0026 [0.0008] J
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.0008]	ND [0.0008]	ND [0.0008]	0.0011 [0.00081] J	ND [0.004]	0.0009 [0.0008] J
SW8081B	Endrin ketone	mg/kg	-	0.012 [0.00065]	0.0059 [0.00065]	0.022 [0.00064]	ND [0.00065]	ND [0.0032]	ND [0.00065]
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.00073]	ND [0.00073]	ND [0.00072]	ND [0.00073]	ND [0.0036]	ND [0.00073]
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.00081]	ND [0.00081]	ND [0.00081]	<b>0.025 [0.00082]</b>	<b>0.2 [0.0041]</b>	<b>0.021 [0.00081]</b>
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.00092]	ND [0.00092]	ND [0.00091]	ND [0.00093]	ND [0.0046]	ND [0.00092]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	<b>0.0038 [0.0008] J</b>	<b>0.0011 [0.0008] J</b>	<b>0.0041 [0.0008]</b>	<b>0.00089 [0.00081] J</b>	<b>0.0047 [0.004] J</b>	ND [0.0008]
SW8081B	Methoxychlor	mg/kg	0.676	0.0067 [0.00086] J	0.0032 [0.00086] J	0.012 [0.00086]	0.005 [0.00087] J	0.043 [0.0043]	0.0039 [0.00086] J
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.028]	ND [0.028]	ND [0.027]	ND [0.028]	ND [0.14]	ND [0.028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00061]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.0008]	ND [0.0008]	ND [0.0008]	ND [0.00078]	ND [0.00079]	ND [0.00079]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.0007]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00068]	ND [0.00068]	ND [0.00068]	ND [0.00067]	ND [0.00067]	ND [0.00067]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	<b>0.23 [0.00059]</b>	<b>0.039 [0.00059]</b>	<b>0.34 [0.00059]</b>	<b>0.085 [0.00058]</b>	<b>0.4 [0.00059]</b>	<b>0.1 [0.00059]</b>
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.5 [0.00059]</b>	<b>0.083 [0.00059]</b>	<b>0.81 [0.00059]</b>	<b>0.22 [0.00058]</b>	<b>0.98 [0.00059]</b>	<b>0.19 [0.00059]</b>

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*E* (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				GSAG12-W GS-AG-12-WHOLE 13438-24 8/2/2012 TF STLP	GSAG13-F GS-AG-13-FILLET 13438-25 8/2/2012 TF STLP	GSAG13-W GS-AG-13-WHOLE 13438-26 8/2/2012 TF STLP	GSAG14-F GS-AG-14-FILLET 13438-27 8/2/2012 TF STLP	GSAG14-W GS-AG-14-WHOLE 13438-28 8/2/2012 TF STLP	GSAG15-F GS-AG-15-FILLET 13438-29 8/2/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	3.1 [0.03]	1.1 [0.03]	3.6 [0.029]	1.9 [0.03]	2.9 [0.029]	0.68 [0.029]
SW7471B	Mercury	mg/kg	0.014	<b>0.023 [0.0099] J</b>	<b>0.046 [0.011]</b>	<b>0.024 [0.011] J</b>	<b>0.033 [0.011]</b>	<b>0.02 [0.01] J</b>	<b>0.056 [0.01]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	<b>0.28 [0.00055]</b>	<b>0.086 [0.00027]</b>	<b>0.27 [0.00054]</b>	<b>0.34 [0.00054]</b>	<b>1.2 [0.0027]</b>	<b>0.12 [0.00055]</b>
SW8081B	4,4'-DDE	mg/kg	0.0093	<b>0.1 [0.00063]</b>	<b>0.028 [0.00032]</b>	<b>0.084 [0.00063]</b>	<b>0.13 [0.00063]</b>	<b>0.43 [0.0032]</b>	<b>0.046 [0.00063]</b>
SW8081B	4,4'-DDT	mg/kg	0.0093	0.0078 [0.00063]	0.0016 [0.00031] J	0.0046 [0.00062]	<b>0.013 [0.00062]</b>	<b>0.044 [0.0031]</b>	0.0041 [0.00063] J
SW8081B	Aldrin	mg/kg	0.00019	ND [0.00075]	ND [0.00037]	ND [0.00074]	ND [0.00074]	ND [0.0037]	ND [0.00075]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.00068]	ND [0.00034]	ND [0.00068]	ND [0.00068]	ND [0.0034]	ND [0.00068]
SW8081B	alpha-Chlordane	mg/kg	0.009	0.002 [0.00083] J	0.0005 [0.00041] J	0.0013 [0.00082] J	0.0016 [0.00082] J	0.0058 [0.0041] J	ND [0.00083]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.0011]	ND [0.00054]	ND [0.0011]	ND [0.0011]	ND [0.0054]	ND [0.0011]
SW8081B	delta-BHC	mg/kg	-	ND [0.00064]	ND [0.00032]	ND [0.00064]	ND [0.00064]	ND [0.0032]	ND [0.00064]
SW8081B	Dieldrin	mg/kg	0.0002	<b>0.0021 [0.0007] J</b>	<b>0.0013 [0.00035] J</b>	<b>0.0035 [0.00069] J</b>	<b>0.0061 [0.00069]</b>	<b>0.021 [0.0035]</b>	<b>0.002 [0.0007] J</b>
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.00079]	ND [0.00039]	ND [0.00078]	ND [0.00078]	ND [0.0039]	ND [0.00079]
SW8081B	Endosulfan II	mg/kg	0.811	0.0014 [0.00074] J	0.00041 [0.00037] J	0.00097 [0.00073] J	0.0017 [0.00073] J	0.006 [0.0037] J	ND [0.00074]
SW8081B	Endosulfan sulfate	mg/kg	0.811	0.0019 [0.00044] J	0.00058 [0.00022] J	0.0014 [0.00043] J	0.0027 [0.00043] J	0.0096 [0.0022] J	0.001 [0.00044] J
SW8081B	Endrin	mg/kg	0.041	0.005 [0.00081]	0.0016 [0.00041] J	0.0041 [0.00081]	0.0077 [0.0008]	0.025 [0.0041]	0.0026 [0.00081] J
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.00081]	0.00047 [0.00041] J	ND [0.0008]	0.00081 [0.0008] J	ND [0.0041]	ND [0.00081]
SW8081B	Endrin ketone	mg/kg	-	ND [0.00065]	ND [0.00033]	ND [0.00065]	ND [0.00065]	ND [0.0033]	ND [0.00065]
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.00073]	0.00037 [0.00037] J	ND [0.00073]	ND [0.00073]	ND [0.0037]	ND [0.00073]
SW8081B	gamma-Chlordane	mg/kg	0.009	<b>0.047 [0.00082]</b>	<b>0.013 [0.00041]</b>	<b>0.036 [0.00081]</b>	<b>0.051 [0.00081]</b>	<b>0.18 [0.0041]</b>	<b>0.018 [0.00082]</b>
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.00093]	ND [0.00046]	ND [0.00092]	ND [0.00092]	ND [0.0046]	ND [0.00093]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.00081]	ND [0.00041]	ND [0.0008]	<b>0.00083 [0.0008] J</b>	ND [0.0041]	ND [0.00081]
SW8081B	Methoxychlor	mg/kg	0.676	0.0073 [0.00087] J	0.002 [0.00044] J	0.005 [0.00086] J	0.0096 [0.00086]	0.033 [0.0044] J	0.0039 [0.00087] J
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.028]	ND [0.014]	ND [0.028]	ND [0.028]	ND [0.14]	ND [0.028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.0008]	ND [0.0008]	ND [0.0008]	ND [0.00079]	ND [0.0008]	ND [0.0008]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00068]	ND [0.00068]	ND [0.00068]	ND [0.00067]	ND [0.00068]	ND [0.00068]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	<b>0.19 [0.00059]</b>	<b>0.057 [0.00059]</b>	<b>0.12 [0.00059]</b>	<b>0.19 [0.00059]</b>	<b>0.29 [0.00059]</b>	<b>0.064 [0.00059]</b>
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.35 [0.00059]</b>	<b>0.083 [0.00059]</b>	<b>0.19 [0.00059]</b>	<b>0.36 [0.00059]</b>	<b>0.56 [0.00059]</b>	<b>0.19 [0.00059]</b>

**Notes:**

STLP = TestAmerica, Pittsburg, PA

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*E* (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				GSAG15-W GS-AG-15-WHOLE 13438-30 8/2/2012 TF STLP	GSAG16-F GS-AG-16-FILLET 13438-31 8/5/2012 TF STLP	GSAG16-W GS-AG-16-WHOLE 13438-32 8/5/2012 TF STLP	GSAG17-F GS-AG-17-FILLET 13438-33 8/5/2012 TF STLP	GSAG17-W GS-AG-17-WHOLE 13438-34 8/5/2012 TF STLP	GSAG18-F GS-AG-18-FILLET 13438-35 8/5/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	2.6 [0.03]	1.3 [0.03]	2.7 [0.029]	1.4 [0.03]	4.4 [0.03]	0.93 [0.03]
SW7471B	Mercury	mg/kg	0.014	<b>0.037 [0.011]</b>	<b>0.039 [0.01]</b>	<b>0.018 [0.01] J</b>	<b>0.022 [0.011] J</b>	<b>0.014 [0.01] J</b>	<b>0.026 [0.011] J</b>
SW8081B	4,4'-DDD	mg/kg	0.013	<b>0.33 [0.00055]</b>	<b>0.16 [0.00055]</b>	<b>0.46 [0.00054]</b>	<b>0.19 [0.00054]</b>	<b>0.58 [0.0027]</b>	<b>0.14 [0.00054]</b>
SW8081B	4,4'-DDE	mg/kg	0.0093	<b>0.12 [0.00063]</b>	<b>0.052 [0.00063]</b>	<b>0.15 [0.00063]</b>	<b>0.056 [0.00063]</b>	<b>0.17 [0.0032]</b>	<b>0.058 [0.00063]</b>
SW8081B	4,4'-DDT	mg/kg	0.0093	<b>0.011 [0.00063]</b>	0.0048 [0.00063]	<b>0.019 [0.00062]</b>	0.0055 [0.00062]	<b>0.02 [0.0031] J</b>	0.0072 [0.00062]
SW8081B	Aldrin	mg/kg	0.00019	ND [0.00075]	ND [0.00075]	ND [0.00074]	ND [0.00074]	ND [0.0037]	ND [0.00074]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.00068]	ND [0.00068]	ND [0.00068]	ND [0.00068]	ND [0.0034]	ND [0.00068]
SW8081B	alpha-Chlordane	mg/kg	0.009	0.002 [0.00083] J	ND [0.00083]	0.0017 [0.00082] J	ND [0.00082]	ND [0.0041]	ND [0.00082]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.0011]	ND [0.0011]	ND [0.0011]	ND [0.0011]	ND [0.0054]	ND [0.0011]
SW8081B	delta-BHC	mg/kg	-	ND [0.00064]	ND [0.00064]	ND [0.00064]	ND [0.00064]	ND [0.0032]	ND [0.00064]
SW8081B	Dieldrin	mg/kg	0.0002	<b>0.0021 [0.0007] J</b>	<b>0.0029 [0.0007] J</b>	<b>0.0045 [0.00069]</b>	<b>0.0035 [0.00069] J</b>	<b>0.011 [0.0035] J</b>	<b>0.0028 [0.00069] J</b>
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.00079]	ND [0.00079]	0.0016 [0.00078] J	ND [0.00078]	ND [0.0039]	ND [0.00078]
SW8081B	Endosulfan II	mg/kg	0.811	0.0016 [0.00074] J	0.0008 [0.00074] J	0.0028 [0.00073] J	ND [0.00073]	ND [0.0037]	0.0011 [0.00073] J
SW8081B	Endosulfan sulfate	mg/kg	0.811	0.0027 [0.00044] J	0.00088 [0.00044] J	0.0094 [0.00043] J	0.002 [0.00043] J	0.0028 [0.0022] J	0.0012 [0.00043] J
SW8081B	Endrin	mg/kg	0.041	0.0072 [0.00081]	0.0023 [0.00081] J	0.016 [0.0008]	0.0025 [0.0008] J	0.0074 [0.0041] J	0.0039 [0.0008] J
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.00081]	ND [0.00081]	0.0022 [0.0008] J	0.0013 [0.0008] J	ND [0.0041]	ND [0.0008]
SW8081B	Endrin ketone	mg/kg	-	ND [0.00065]	ND [0.00065]	ND [0.00065]	ND [0.00065]	0.037 [0.0033]	ND [0.00065]
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.00073]	ND [0.00073]	ND [0.00073]	ND [0.00073]	ND [0.0037]	ND [0.00073]
SW8081B	gamma-Chlordane	mg/kg	0.009	<b>0.047 [0.00082]</b>	<b>0.027 [0.00082]</b>	<b>0.086 [0.00081]</b>	<b>0.027 [0.00081]</b>	ND [0.0041]	<b>0.023 [0.00081]</b>
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.00093]	ND [0.00093]	ND [0.00092]	ND [0.00092]	ND [0.0046]	ND [0.00092]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	<b>0.0013 [0.00081] J</b>	ND [0.00081]	<b>0.0013 [0.0008] J</b>	<b>0.002 [0.0008] J</b>	<b>0.0063 [0.0041] J</b>	ND [0.0008]
SW8081B	Methoxychlor	mg/kg	0.676	0.0095 [0.00087]	0.0045 [0.00087] J	0.019 [0.00086]	ND [0.00086]	0.014 [0.0044] J	0.0045 [0.00086] J
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.028]	ND [0.028]	ND [0.028]	ND [0.028]	ND [0.14]	ND [0.028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.00079]	ND [0.00079]	ND [0.0008]	ND [0.0008]	ND [0.0008]	ND [0.00079]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00067]	ND [0.00067]	ND [0.00068]	ND [0.00068]	ND [0.00068]	ND [0.00067]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	<b>0.42 [0.00059]</b>	<b>0.061 [0.00059]</b>	<b>0.26 [0.00059]</b>	<b>0.14 [0.00059]</b>	<b>0.34 [0.00059]</b>	<b>0.1 [0.00059]</b>
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.87 [0.00059]</b>	<b>0.23 [0.00059]</b>	<b>0.6 [0.00059]</b>	<b>0.29 [0.00059]</b>	<b>0.73 [0.00059]</b>	<b>0.26 [0.00059]</b>

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E* (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).



**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				GSAG18-W GS-AG-18-WHOLE 13438-36 8/5/2012 TF STLP	GSAG19-F GS-AG-19-FILLET 13438-37 8/5/2012 TF STLP	GSAG19-W GS-AG-19-WHOLE 13438-38 8/5/2012 TF STLP	GSAG20-F GS-AG-20-FILLET 13438-39 8/5/2012 TF STLP	GSAG20-W GS-AG-20-WHOLE 13438-40 8/5/2012 TF STLP	GSAG21-F GS-AG-21-FILLET 13438-41 8/2/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	2.9 [0.03]	1.4 [0.03]	2.8 [0.03]	0.7 [0.03]	4 [0.029]	1.2 [0.03]
SW7471B	Mercury	mg/kg	0.014	0.013 [0.011] J	<b>0.043 [0.01]</b>	<b>0.024 [0.011] J</b>	<b>0.038 [0.011]</b>	<b>0.024 [0.01] J</b>	<b>0.026 [0.01] J</b>
SW8081B	4,4'-DDD	mg/kg	0.013	<b>0.52 [0.00055]</b>	<b>0.17 [0.00055]</b>	<b>0.5 [0.00055]</b>	<b>0.03 [0.000055]</b>	<b>0.2 [0.00054]</b>	<b>0.058 [0.000055]</b>
SW8081B	4,4'-DDE	mg/kg	0.0093	<b>0.2 [0.00063]</b>	<b>0.055 [0.00063]</b>	<b>0.15 [0.00063]</b>	<b>0.01 [0.000063]</b>	<b>0.067 [0.00062]</b>	<b>0.019 [0.000063]</b>
SW8081B	4,4'-DDT	mg/kg	0.0093	<b>0.026 [0.00063]</b>	0.0045 [0.00063]	<b>0.013 [0.00063]</b>	0.00092 [0.000063]	0.0056 [0.00062]	0.0018 [0.000063]
SW8081B	Aldrin	mg/kg	0.00019	ND [0.00075]	ND [0.00075]	ND [0.00075]	ND [0.000075]	ND [0.00074]	ND [0.000075]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.00068]	ND [0.00068]	ND [0.00068]	ND [0.000068]	ND [0.00067]	ND [0.000068]
SW8081B	alpha-Chlordane	mg/kg	0.009	0.002 [0.00083] J	0.001 [0.00083] J	0.0019 [0.00083] J	0.00018 [0.000083] J	0.00082 [0.00081] J	0.00031 [0.000083] J
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.0011]	ND [0.0011]	ND [0.0011]	ND [0.00011]	ND [0.0011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.00064]	ND [0.00064]	ND [0.00064]	ND [0.000064]	ND [0.00063]	ND [0.000064]
SW8081B	Dieldrin	mg/kg	0.0002	<b>0.009 [0.0007]</b>	<b>0.0014 [0.0007] J</b>	<b>0.00094 [0.0007] J</b>	<b>0.00022 [0.00007] J</b>	<b>0.0013 [0.00069] J</b>	<b>0.00044 [0.00007] J</b>
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.00079]	ND [0.00079]	ND [0.00079]	ND [0.000079]	ND [0.00077]	ND [0.000079]
SW8081B	Endosulfan II	mg/kg	0.811	0.0027 [0.00074] J	0.00098 [0.00074] J	0.0026 [0.00074] J	0.00029 [0.000074] J	0.0015 [0.00073] J	0.00045 [0.000074] J
SW8081B	Endosulfan sulfate	mg/kg	0.811	0.0043 [0.00044] J	0.0012 [0.00044] J	0.0034 [0.00044] J	0.00025 [0.000044] J	0.0016 [0.00043] J	0.00046 [0.000044] J
SW8081B	Endrin	mg/kg	0.041	0.011 [0.00081]	0.0031 [0.00081] J	0.0072 [0.00081]	0.00058 [0.000081]	0.0035 [0.0008] J	0.00089 [0.000081]
SW8081B	Endrin aldehyde	mg/kg	0.041	0.0012 [0.00081] J	ND [0.00081]	0.00084 [0.00081] J	ND [0.000081]	ND [0.0008]	ND [0.000081]
SW8081B	Endrin ketone	mg/kg	-	ND [0.00065]	ND [0.00065]	ND [0.00065]	0.0033 [0.000065]	0.011 [0.00064]	0.0057 [0.000065]
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.00073]	ND [0.00073]	ND [0.00073]	0.000078 [0.000073] J	ND [0.00072]	0.0001 [0.000073] J
SW8081B	gamma-Chlordane	mg/kg	0.009	<b>0.075 [0.00082]</b>	ND [0.00082]	<b>0.063 [0.00082]</b>	ND [0.000082]	ND [0.00081]	ND [0.000082]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.00093]	ND [0.00093]	ND [0.00093]	ND [0.000093]	ND [0.00091]	ND [0.000093]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	<b>0.0014 [0.00081] J</b>	<b>0.0016 [0.00081] J</b>	ND [0.00081]	ND [0.000081]	ND [0.0008]	0.000083 [0.000081] J
SW8081B	Methoxychlor	mg/kg	0.676	0.014 [0.00087]	0.0054 [0.00087] J	0.013 [0.00087]	0.0011 [0.000087]	0.0055 [0.00086] J	0.0018 [0.000087]
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.028]	ND [0.028]	ND [0.028]	ND [0.0028]	ND [0.027]	ND [0.0028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.00079]	ND [0.0008]	ND [0.00079]	ND [0.00079]	ND [0.0008]	ND [0.0008]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00067]	ND [0.00068]	ND [0.00067]	ND [0.00067]	ND [0.00068]	ND [0.00068]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	<b>0.36 [0.00059]</b>	<b>0.1 [0.00059]</b>	<b>0.23 [0.00059]</b>	<b>0.013 [0.00059]</b>	<b>0.066 [0.00059]</b>	<b>0.022 [0.00059]</b>
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.78 [0.00059]</b>	<b>0.31 [0.00059]</b>	<b>0.54 [0.00059]</b>	<b>0.078 [0.00059]</b>	<b>0.13 [0.00059]</b>	<b>0.087 [0.00059]</b>

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[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				PSAG01-F PS-AG-01-Fillet 18159-1 8/2/2012 TF STLP	PSAG01-W PS-AG-01-Whole 18159-2 8/2/2012 TF STLP	PSAG02-F PS-AG-02-Fillet 18159-3 8/3/2012 TF STLP	PSAG02-W PS-AG-02-Whole 18159-4 8/3/2012 TF STLP	PSAG03-F PS-AG-03-Fillet 18159-5 8/3/2012 TF STLP	PSAG03-W PS-AG-03-Whole 18159-6 8/3/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	0.39 [0.029]	2.2 [0.03]	0.3 [0.029]	2.2 [0.03]	0.68 [0.029]	3 [0.029]
SW7471B	Mercury	mg/kg	0.014	<b>0.095 [0.011]</b>	<b>0.042 [0.011]</b>	<b>0.038 [0.011]</b>	<b>0.021 [0.011] J</b>	<b>0.042 [0.011]</b>	<b>0.03 [0.011] J</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.0012 [0.000054]	0.0039 [0.000054]	0.0033 [0.000054]	<b>0.024 [0.000055]</b>	<b>0.028 [0.000055]</b>	<b>0.14 [0.00053]</b>
SW8081B	4,4'-DDE	mg/kg	0.0093	0.0041 [0.000063]	<b>0.02 [0.000063]</b>	0.0014 [0.000063]	<b>0.01 [0.000063]</b>	<b>0.011 [0.000063]</b>	<b>0.056 [0.00062]</b>
SW8081B	4,4'-DDT	mg/kg	0.0093	0.00012 [0.000062] J	0.0019 [0.000062]	0.00014 [0.000062] J	0.00069 [0.000063]	0.00056 [0.000063]	0.0023 [0.00061] J
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000074]	ND [0.000074]	ND [0.000074]	ND [0.000075]	ND [0.000075]	ND [0.00073]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.00067]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000082]	ND [0.000082]	ND [0.000082]	ND [0.000083]	ND [0.000083]	ND [0.00081]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	0.00012 [0.00011] J	ND [0.00011]	ND [0.0011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.00063]
SW8081B	Dieldrin	mg/kg	0.0002	ND [0.000069]	ND [0.000069]	ND [0.000069]	<b>0.00034 [0.00007] J</b>	ND [0.00007]	ND [0.00068]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000078]	ND [0.000078]	ND [0.000078]	ND [0.000079]	ND [0.000079]	ND [0.00077]
SW8081B	Endosulfan II	mg/kg	0.811	ND [0.000073]	ND [0.000073]	ND [0.000073]	0.00024 [0.000074] J	0.00013 [0.000074] J	ND [0.00072]
SW8081B	Endosulfan sulfate	mg/kg	0.811	ND [0.000043]	0.00032 [0.000043] J	ND [0.000043]	0.00024 [0.000044] J	0.00018 [0.000044] J	0.00092 [0.00043] J
SW8081B	Endrin	mg/kg	0.041	0.000096 [0.00008] J	0.00041 [0.00008]	0.00013 [0.00008] J	0.0014 [0.000081]	0.00056 [0.000081]	0.0028 [0.00079] J
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.00008]	ND [0.00008]	ND [0.00008]	ND [0.000081]	ND [0.000081]	ND [0.00079]
SW8081B	Endrin ketone	mg/kg	-	0.0002 [0.000065] J	0.0012 [0.000065]	0.00021 [0.000065] J	0.0012 [0.000065]	0.00074 [0.000065]	0.0027 [0.00064] J
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.000073]	ND [0.000073]	ND [0.000073]	ND [0.000073]	ND [0.000073]	ND [0.00072]
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000081]	0.00012 [0.000081] J	ND [0.000081]	0.00032 [0.000082] J	ND [0.000082]	ND [0.0008]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000092]	ND [0.000092]	ND [0.000092]	ND [0.000093]	ND [0.000093]	ND [0.00091]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.00008]	ND [0.00008]	ND [0.00008]	<b>0.0011 [0.000081]</b>	0.00033 [0.000081] J	<b>0.0013 [0.00079] J</b>
SW8081B	Methoxychlor	mg/kg	0.676	0.00015 [0.000086] J, B	0.00075 [0.000086] J	ND [0.000086]	0.0012 [0.000087]	0.00091 [0.000087]	0.002 [0.00085] J
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.027]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00061]	ND [0.00061]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.00079]	ND [0.0008]	ND [0.0008]	ND [0.0008]	ND [0.00078]	ND [0.00078]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.0007]	ND [0.0007]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00067]	ND [0.00068]	ND [0.00068]	ND [0.00068]	ND [0.00067]	ND [0.00067]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	<b>0.0028 [0.00059] J</b>	<b>0.074 [0.00059]</b>	<b>0.005 [0.00059]</b>	<b>0.023 [0.00059]</b>	<b>0.015 [0.00058]</b>	<b>0.092 [0.00058]</b>
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.0067 [0.00059]</b>	<b>0.07 [0.00059]</b>	<b>0.004 [0.00059] J</b>	<b>0.029 [0.00059]</b>	<b>0.033 [0.00058]</b>	<b>0.17 [0.00058]</b>

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E (Italics)* = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				PSAG04-F PS-AG-04-Fillet 18159-7 8/3/2012 TF STLP	PSAG04-W PS-AG-04-Whole 18159-8 8/3/2012 TF STLP	PSAG05-F PS-AG-05-Fillet 18159-9 8/3/2012 TF STLP	PSAG05-W PS-AG-05-Whole 18159-10 8/3/2012 TF STLP	PSAG06-F PS-AG-06-Fillet 18159-11 8/3/2012 TF STLP
Method	Analyte	Units	Project Action Limit					
BDTL	Total Lipids	Percent	-	0.22 [0.029]	2 [0.03]	0.31 [0.03]	1.3 [0.03]	0.15 [0.029]
SW7471B	Mercury	mg/kg	0.014	<b>0.043 [0.011]</b>	<b>0.035 [0.01]</b>	<b>0.043 [0.011]</b>	<b>0.02 [0.01] J</b>	<b>0.057 [0.01]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.00045 [0.000055]	0.0021 [0.000054]	0.001 [0.000055]	0.0026 [0.000054]	0.00037 [0.000055] J
SW8081B	4,4'-DDE	mg/kg	0.0093	0.00082 [0.000063]	0.0055 [0.000063]	0.0015 [0.000063]	0.004 [0.000063]	0.0011 [0.000063]
SW8081B	4,4'-DDT	mg/kg	0.0093	ND [0.000063]	0.000087 [0.000062] J	0.00012 [0.000063] J	0.00023 [0.000062] J	0.00012 [0.000063] J
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000075]	ND [0.000074]	ND [0.000075]	ND [0.000074]	ND [0.000075]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000068]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000083]	ND [0.000082]	ND [0.000083]	ND [0.000082]	ND [0.000083]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000064]
SW8081B	Dieldrin	mg/kg	0.0002	ND [0.00007]	ND [0.000069]	ND [0.00007]	ND [0.000069]	ND [0.00007]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000079]	ND [0.000078]	ND [0.000079]	ND [0.000078]	ND [0.000079]
SW8081B	Endosulfan II	mg/kg	0.811	ND [0.000074]	ND [0.000073]	ND [0.000074]	ND [0.000073]	ND [0.000074]
SW8081B	Endosulfan sulfate	mg/kg	0.811	ND [0.000044]	0.00011 [0.000043] J	ND [0.000044]	ND [0.000043]	ND [0.000044]
SW8081B	Endrin	mg/kg	0.041	0.00012 [0.000081] J	ND [0.00008]	ND [0.000081]	ND [0.00008]	ND [0.000081]
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.000081]	ND [0.00008]	ND [0.000081]	ND [0.00008]	ND [0.000081]
SW8081B	Endrin ketone	mg/kg	-	ND [0.000065]	0.00028 [0.000065] J	ND [0.000065]	0.00023 [0.000065] J	0.000074 [0.000065] J
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.000073]	ND [0.000073]	ND [0.000073]	ND [0.000073]	ND [0.000073]
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000082]	ND [0.000081]	ND [0.000082]	ND [0.000081]	ND [0.000082]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000093]	ND [0.000092]	ND [0.000093]	ND [0.000092]	ND [0.000093]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.000081]	ND [0.00008]	ND [0.000081]	ND [0.00008]	ND [0.000081]
SW8081B	Methoxychlor	mg/kg	0.676	ND [0.000087]	0.00024 [0.000086] J, B	ND [0.000087]	0.00016 [0.000086] J, B	0.000087 [0.000087] J, B
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00061]	ND [0.00062]	ND [0.00062]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.00078]	ND [0.0008]	ND [0.0008]	ND [0.00079]	ND [0.00079]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.0007]	ND [0.00071]	ND [0.00071]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00067]	ND [0.00068]	ND [0.00068]	ND [0.00067]	ND [0.00067]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	<b>0.0034 [0.00058] J</b>	ND [0.00059]	ND [0.00059]	ND [0.00059]	ND [0.00059]
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.0066 [0.00058]</b>	<b>0.0041 [0.00059] J</b>	<b>0.0051 [0.00059]</b>	<b>0.006 [0.00059]</b>	0.0016 [0.00059] J

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E (Italics)* = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

				Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory	PSAG06-W PS-AG-06-Whole 18159-12 8/3/2012 TF STLP	PSAG07-F PS-AG-07-Fillet 18159-13 8/3/2012 TF STLP	PSAG07-W PS-AG-07-Whole 18159-14 8/3/2012 TF STLP	PSAG08-F PS-AG-08-Fillet 18159-15 8/4/2012 TF STLP	PSAG08-W PS-AG-08-Whole 18159-16 8/4/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	0.63 [0.029]	0.6 [0.029]	1.1 [0.029]	0.11 [0.029]	0.39 [0.03]	
SW7471B	Mercury	mg/kg	0.014	<b>0.045 [0.011]</b>	<b>0.037 [0.011]</b>	<b>0.025 [0.01] J</b>	<b>0.13 [0.0099]</b>	<b>0.063 [0.0099]</b>	
SW8081B	4,4'-DDD	mg/kg	0.013	0.0019 [0.000055]	0.0015 [0.000054]	0.0028 [0.000054]	0.00012 [0.000055] J	0.00022 [0.000054] J	
SW8081B	4,4'-DDE	mg/kg	0.0093	0.0055 [0.000063]	0.0038 [0.000063]	0.0065 [0.000062]	0.00021 [0.000063] J	0.00028 [0.000063] J	
SW8081B	4,4'-DDT	mg/kg	0.0093	0.00037 [0.000063] J	0.00032 [0.000062] J	0.0002 [0.000062] J	ND [0.000063]	0.000064 [0.000062] J	
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000075]	ND [0.000074]	ND [0.000074]	ND [0.000075]	ND [0.000074]	
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000068]	ND [0.000068]	ND [0.000067]	ND [0.000068]	ND [0.000068]	
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000083]	ND [0.000082]	ND [0.000081]	ND [0.000083]	ND [0.000082]	
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	
SW8081B	delta-BHC	mg/kg	-	ND [0.000064]	ND [0.000064]	ND [0.000063]	ND [0.000064]	ND [0.000064]	
SW8081B	Dieldrin	mg/kg	0.0002	ND [0.00007]	ND [0.000069]	ND [0.000069]	ND [0.00007]	ND [0.000069]	
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000079]	ND [0.000078]	ND [0.000077]	ND [0.000079]	ND [0.000078]	
SW8081B	Endosulfan II	mg/kg	0.811	ND [0.000074]	ND [0.000073]	ND [0.000073]	ND [0.000074]	ND [0.000073]	
SW8081B	Endosulfan sulfate	mg/kg	0.811	ND [0.000044]	ND [0.000043]	0.000097 [0.000043] J	ND [0.000044]	ND [0.000043]	
SW8081B	Endrin	mg/kg	0.041	ND [0.000081]	ND [0.00008]	0.00009 [0.00008] J	ND [0.000081]	ND [0.00008]	
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.000081]	ND [0.00008]	ND [0.00008]	ND [0.000081]	ND [0.00008]	
SW8081B	Endrin ketone	mg/kg	-	0.000072 [0.000065] J	ND [0.000065]	0.00012 [0.000064] J	ND [0.000065]	0.000098 [0.000065] J	
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.000073]	ND [0.000073]	ND [0.000072]	ND [0.000073]	ND [0.000073]	
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000082]	ND [0.000081]	ND [0.000081]	ND [0.000082]	ND [0.000081]	
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000093]	ND [0.000092]	ND [0.000091]	ND [0.000093]	ND [0.000092]	
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.000081]	ND [0.00008]	0.00011 [0.00008] J	ND [0.000081]	ND [0.00008]	
SW8081B	Methoxychlor	mg/kg	0.676	0.00017 [0.000087] J, B	0.00011 [0.000086] J, B	0.00014 [0.000086] J, B	ND [0.000087]	ND [0.000086]	
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0028]	ND [0.0028]	ND [0.0027]	ND [0.0028]	ND [0.0028]	
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00062]	ND [0.00061]	ND [0.00062]	ND [0.00061]	
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.00079]	ND [0.0008]	ND [0.00078]	ND [0.0008]	ND [0.00078]	
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.00071]	ND [0.0007]	ND [0.00071]	ND [0.0007]	
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00067]	ND [0.00068]	ND [0.00067]	ND [0.00068]	ND [0.00067]	
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	ND [0.00059]	ND [0.00059]	ND [0.00058]	ND [0.00059]	ND [0.00058]	
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.0057 [0.00059]</b>	0.0011 [0.00059] J	<b>0.004 [0.00058] J</b>	0.0016 [0.00059] J	0.0022 [0.00058] J	

**Notes:**

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**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E* (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				PSAG09-F PS-AG-09-Fillet 18159-17 8/4/2012 TF STLP	PSAG09-W PS-AG-09-Whole 18159-18 8/4/2012 TF STLP	PSAG10-F PS-AG-10-Fillet 18159-19 8/4/2012 TF STLP	PSAG10-W PS-AG-10-Whole 18159-20 8/4/2012 TF STLP	PSAG11-F PS-AG-11-Fillet 18159-21 8/4/2012 TF STLP	PSAG11-W PS-AG-11-Whole 18159-22 8/4/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	0.11 [0.03]	0.87 [0.03]	0.41 [0.029]	2.5 [0.03]	0.22 [0.03]	0.56 [0.029]
SW7471B	Mercury	mg/kg	0.014	<b>0.027 [0.011] J</b>	<b>0.039 [0.011]</b>	<b>0.059 [0.011]</b>	<b>0.052 [0.0099]</b>	<b>0.12 [0.01]</b>	<b>0.084 [0.01]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.0009 [0.000054]	0.0012 [0.000054]	0.00034 [0.000054] J	0.002 [0.000054]	0.00024 [0.000054] J	0.0003 [0.000055] J
SW8081B	4,4'-DDE	mg/kg	0.0093	0.0028 [0.000063]	0.007 [0.000063]	0.0007 [0.000062]	0.0059 [0.000063]	0.00035 [0.000062] J	0.0006 [0.000063]
SW8081B	4,4'-DDT	mg/kg	0.0093	0.00012 [0.000062] J	0.001 [0.000062]	ND [0.000062]	0.000086 [0.000062] J	ND [0.000062]	ND [0.000063]
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000074]	ND [0.000074]	ND [0.000074]	ND [0.000074]	ND [0.000074]	ND [0.000075]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000068]	0.000071 [0.000068] J	ND [0.000067]	ND [0.000068]	ND [0.000067]	ND [0.000068]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000082]	ND [0.000082]	ND [0.000081]	ND [0.000082]	ND [0.000081]	ND [0.000083]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000064]	ND [0.000064]	ND [0.000063]	ND [0.000064]	ND [0.000063]	ND [0.000064]
SW8081B	Dieldrin	mg/kg	0.0002	ND [0.000069]	ND [0.000069]	ND [0.000069]	ND [0.000069]	ND [0.000069]	ND [0.00007]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000078]	ND [0.000078]	ND [0.000077]	ND [0.000078]	ND [0.000077]	ND [0.000079]
SW8081B	Endosulfan II	mg/kg	0.811	ND [0.000073]	ND [0.000073]	ND [0.000073]	ND [0.000073]	ND [0.000073]	ND [0.000074]
SW8081B	Endosulfan sulfate	mg/kg	0.811	0.000098 [0.000043] J	0.00014 [0.000043] J	ND [0.000043]	ND [0.000043]	ND [0.000043]	ND [0.000044]
SW8081B	Endrin	mg/kg	0.041	0.000082 [0.00008] J	0.00039 [0.00008] J	ND [0.00008]	0.00013 [0.00008] J	ND [0.00008]	ND [0.000081]
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.00008]	ND [0.00008]	ND [0.00008]	ND [0.00008]	ND [0.00008]	ND [0.000081]
SW8081B	Endrin ketone	mg/kg	-	0.00024 [0.000065] J	0.00059 [0.000065]	ND [0.000064]	ND [0.000065]	ND [0.000064]	ND [0.000065]
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.000073]	ND [0.000073]	ND [0.000072]	ND [0.000073]	ND [0.000072]	ND [0.000073]
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000081]	0.000092 [0.000081] J	ND [0.000081]	ND [0.000081]	ND [0.000081]	ND [0.000082]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000092]	ND [0.000092]	ND [0.000091]	ND [0.000092]	ND [0.000091]	ND [0.000093]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.00008]	ND [0.00008]	ND [0.00008]	ND [0.00008]	ND [0.00008]	ND [0.000081]
SW8081B	Methoxychlor	mg/kg	0.676	0.00019 [0.000086] J, B	0.00041 [0.000086] J	ND [0.000086]	0.00014 [0.000086] J	ND [0.000086]	ND [0.000087]
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0028]	ND [0.0028]	ND [0.0027]	ND [0.0028]	ND [0.0027]	ND [0.0028]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00061]	ND [0.00062]	ND [0.00062]	ND [0.00061]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.0008]	ND [0.00078]	ND [0.0008]	ND [0.0008]	ND [0.00078]	ND [0.00079]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.0007]	ND [0.00071]	ND [0.00071]	ND [0.0007]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00068]	ND [0.00067]	ND [0.00068]	ND [0.00068]	ND [0.00067]	ND [0.00067]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	ND [0.00059]	ND [0.00058]	ND [0.00059]	ND [0.00059]	ND [0.00058]	ND [0.00059]
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.0054 [0.00059]</b>	<b>0.022 [0.00058]</b>	0.0025 [0.00059] J	<b>0.0061 [0.00059]</b>	<b>0.0028 [0.00058] J</b>	<b>0.0053 [0.00059]</b>

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E* (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				PSAG12-F PS-AG-12-Fillet 18159-23 8/4/2012 TF STLP	PSAG12-W PS-AG-12-Whole 18159-24 8/4/2012 TF STLP	PSAG13-F PS-AG-13-Fillet 18159-25 8/4/2012 TF STLP	PSAG13-W PS-AG-13-Whole 18159-26 8/4/2012 TF STLP	PSAG14-F PS-AG-14-Fillet 18159-27 8/7/2012 TF STLP	PSAG14-W PS-AG-14-Whole 18159-28 8/7/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	0.47 [0.029]	1.5 [0.029]	0.47 [0.03]	1.8 [0.029]	0.6 [0.029]	2.5 [0.03]
SW7471B	Mercury	mg/kg	0.014	<b>0.041 [0.01]</b>	<b>0.03 [0.01] J</b>	<b>0.038 [0.01]</b>	<b>0.027 [0.011] J</b>	<b>0.062 [0.011]</b>	<b>0.041 [0.011]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.00085 [0.000055]	0.0023 [0.00011]	0.0059 [0.000055]	<b>0.031 [0.000054]</b>	0.00047 [0.000055]	0.0031 [0.000053]
SW8081B	4,4'-DDE	mg/kg	0.0093	0.001 [0.000063]	0.0029 [0.00012]	0.0013 [0.000063]	<b>0.011 [0.000063]</b>	0.00051 [0.000063]	0.0042 [0.000062]
SW8081B	4,4'-DDT	mg/kg	0.0093	0.000066 [0.000063] J	0.00014 [0.00012] J	0.00025 [0.000063] J	0.0012 [0.000062]	ND [0.000063]	0.00013 [0.000061] J
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000075]	ND [0.00015]	ND [0.000075]	ND [0.000074]	ND [0.000075]	ND [0.000073]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000068]	ND [0.00013]	ND [0.000068]	ND [0.000068]	ND [0.000068]	ND [0.000067]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000083]	ND [0.00016]	ND [0.000083]	0.00028 [0.000082] J	ND [0.000083]	ND [0.000081]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00021]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000064]	ND [0.00013]	ND [0.000064]	ND [0.000064]	ND [0.000064]	ND [0.000063]
SW8081B	Dieldrin	mg/kg	0.0002	ND [0.00007]	ND [0.00014]	ND [0.00007]	ND [0.000069]	ND [0.00007]	ND [0.000068]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000079]	ND [0.00015]	ND [0.000079]	ND [0.000078]	ND [0.000079]	ND [0.000077]
SW8081B	Endosulfan II	mg/kg	0.811	ND [0.000074]	ND [0.00014]	ND [0.000074]	ND [0.000073]	ND [0.000074]	ND [0.000072]
SW8081B	Endosulfan sulfate	mg/kg	0.811	ND [0.000044]	ND [0.000085]	ND [0.000044]	0.00018 [0.000043] J	ND [0.000044]	ND [0.000043]
SW8081B	Endrin	mg/kg	0.041	0.000083 [0.000081] J	0.00029 [0.00016] J	ND [0.000081]	0.00041 [0.00008]	ND [0.000081]	0.00013 [0.000079] J
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.000081]	ND [0.00016]	ND [0.000081]	ND [0.00008]	ND [0.000081]	ND [0.000079]
SW8081B	Endrin ketone	mg/kg	-	ND [0.000065]	ND [0.00013]	ND [0.000065]	0.00037 [0.000065] J	ND [0.000065]	0.000097 [0.000064] J
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.000073]	ND [0.00014]	ND [0.000073]	ND [0.000073]	ND [0.000073]	ND [0.000072]
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000082]	ND [0.00016]	ND [0.000082]	0.00022 [0.000081] J	ND [0.000082]	ND [0.00008]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000093]	ND [0.00018]	ND [0.000093]	ND [0.000092]	ND [0.000093]	ND [0.000091]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.000081]	ND [0.00016]	ND [0.000081]	0.000084 [0.00008] J	ND [0.000081]	ND [0.000079]
SW8081B	Methoxychlor	mg/kg	0.676	ND [0.000087]	0.00023 [0.00017] J	ND [0.000087]	ND [0.000086]	ND [0.000087]	0.000095 [0.000085] J
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0028]	ND [0.0055]	ND [0.0028]	ND [0.0028]	ND [0.0028]	ND [0.0027]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	ND [0.00061]	ND [0.00062]	ND [0.00061]	ND [0.00062]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.0008]	ND [0.00078]	ND [0.00079]	ND [0.00078]	ND [0.00079]	ND [0.0008]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	ND [0.0007]	ND [0.00071]	ND [0.0007]	ND [0.00071]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00068]	ND [0.00067]	ND [0.00067]	ND [0.00067]	ND [0.00067]	ND [0.00068]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	ND [0.00059]	ND [0.00058]	<b>0.023 [0.00059]</b>	<b>0.036 [0.00058]</b>	ND [0.00059]	ND [0.00059]
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.0027 [0.00059] J</b>	<b>0.0081 [0.00058]</b>	<b>0.0071 [0.00059]</b>	<b>0.036 [0.00058]</b>	0.0013 [0.00059] J	ND [0.00059]

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E* (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				PSAG15-F PS-AG-15-Fillet 18159-29 8/7/2012 TF STLP	PSAG15-W PS-AG-15-Whole 18159-30 8/7/2012 TF STLP	PSAG16-F PS-AG-16-Fillet 18159-31 8/8/2012 TF STLP	PSAG16-W PS-AG-16-Whole 18159-32 8/8/2012 TF STLP	PSAG17-F PS-AG-17-Fillet 18159-33 8/8/2012 TF STLP	PSAG17-W PS-AG-17-Whole 18159-34 8/8/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	0.4 [0.03]	0.96 [0.029]	0.64 [0.029]	2 [0.03]	1.3 [0.03]	2 [0.03]
SW7471B	Mercury	mg/kg	0.014	<b>0.041 [0.0098]</b>	<b>0.038 [0.01]</b>	<b>0.052 [0.01]</b>	<b>0.033 [0.011]</b>	<b>0.04 [0.0099]</b>	<b>0.025 [0.011] J</b>
SW8081B	4,4'-DDD	mg/kg	0.013	0.00088 [0.000054]	0.0038 [0.000054]	0.00041 [0.000053]	0.0023 [0.000054]	<b>0.017 [0.000053]</b>	<b>0.038 [0.000054]</b>
SW8081B	4,4'-DDE	mg/kg	0.0093	0.0023 [0.000062]	<b>0.011 [0.000063]</b>	0.0007 [0.000062]	0.0045 [0.000062]	0.008 [0.000062]	<b>0.018 [0.000062]</b>
SW8081B	4,4'-DDT	mg/kg	0.0093	0.00051 [0.000062]	0.0021 [0.000062]	0.00012 [0.000061] J	0.00014 [0.000062] J	0.00054 [0.000061]	0.00053 [0.000062]
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000074]	ND [0.000074]	ND [0.000073]	ND [0.000074]	ND [0.000073]	ND [0.000074]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000067]	ND [0.000068]	ND [0.000067]	ND [0.000067]	ND [0.000067]	ND [0.000067]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000081]	ND [0.000082]	ND [0.000081]	ND [0.000081]	ND [0.000081]	0.000095 [0.000081] J
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]	ND [0.00011]
SW8081B	delta-BHC	mg/kg	-	ND [0.000063]	ND [0.000064]	ND [0.000063]	ND [0.000063]	ND [0.000063]	ND [0.000063]
SW8081B	Dieldrin	mg/kg	0.0002	ND [0.000069]	ND [0.000069]	ND [0.000068]	ND [0.000069]	ND [0.000068]	ND [0.000069]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000077]	ND [0.000078]	ND [0.000077]	ND [0.000077]	ND [0.000077]	ND [0.000077]
SW8081B	Endosulfan II	mg/kg	0.811	ND [0.000073]	ND [0.000073]	ND [0.000072]	ND [0.000073]	ND [0.000072]	ND [0.000073]
SW8081B	Endosulfan sulfate	mg/kg	0.811	0.000091 [0.000043] J	0.00023 [0.000043] J	ND [0.000043]	0.000071 [0.000043] J	0.00014 [0.000043] J	0.00031 [0.000043] J
SW8081B	Endrin	mg/kg	0.041	ND [0.00008]	0.00058 [0.00008]	ND [0.000079]	0.00011 [0.00008] J	0.00034 [0.000079] J	0.00083 [0.00008]
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.00008]	ND [0.00008]	ND [0.000079]	ND [0.00008]	ND [0.000079]	ND [0.00008]
SW8081B	Endrin ketone	mg/kg	-	0.00024 [0.000064] J	0.001 [0.000065]	ND [0.000064]	0.00017 [0.000064] J	0.00027 [0.000064] J	0.001 [0.000064]
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.000072]	ND [0.000073]	ND [0.000072]	ND [0.000072]	ND [0.000072]	ND [0.000072]
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000081]	0.00013 [0.000081] J	ND [0.00008]	ND [0.000081]	ND [0.00008]	ND [0.000081]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000091]	ND [0.000092]	ND [0.000091]	ND [0.000091]	ND [0.000091]	ND [0.000091]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.00008]	ND [0.00008]	ND [0.000079]	ND [0.00008]	ND [0.000079]	ND [0.00008]
SW8081B	Methoxychlor	mg/kg	0.676	0.00018 [0.000086] J	0.00081 [0.000086] J	ND [0.000085]	ND [0.000086]	0.00029 [0.000085] J	ND [0.000086]
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0027]	ND [0.0028]	ND [0.0027]	ND [0.0027]	ND [0.0027]	ND [0.0027]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00061]	ND [0.00061]	ND [0.00062]	ND [0.00062]	ND [0.00061]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.00078]	ND [0.00078]	ND [0.0008]	ND [0.0008]	ND [0.00078]	ND [0.0008]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.0007]	ND [0.0007]	ND [0.00071]	ND [0.00071]	ND [0.0007]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00067]	ND [0.00067]	ND [0.00068]	ND [0.00068]	ND [0.00067]	ND [0.00068]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	ND [0.00058]	<b>0.006 [0.00058]</b>	ND [0.00059]	<b>0.004 [0.00059] J</b>	<b>0.013 [0.00058]</b>	<b>0.032 [0.00059]</b>
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.0068 [0.00058]</b>	<b>0.034 [0.00058]</b>	ND [0.00059]	<b>0.006 [0.00059]</b>	<b>0.019 [0.00058]</b>	<b>0.067 [0.00059]</b>

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

E (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

Location ID Sample ID Lab Sample ID Date Collected Matrix Laboratory				PSAG18-F PS-AG-18-Fillet 18159-35 8/8/2012 TF STLP	PSAG18-W PS-AG-18-Whole 18159-36 8/8/2012 TF STLP	PSAG19-F PS-AG-19-Fillet 18159-37 8/8/2012 TF STLP	PSAG19-W PS-AG-19-Whole 18159-38 8/8/2012 TF STLP	PSAG20-F PS-AG-20-Fillet 18159-39 8/8/2012 TF STLP	PSAG20-W PS-AG-20-Whole 18159-40 8/8/2012 TF STLP
Method	Analyte	Units	Project Action Limit						
BDTL	Total Lipids	Percent	-	1.2 [0.029]	2.9 [0.03]	0.5 [0.029]	2.1 [0.03]	0.64 [0.029]	1.5 [0.03]
SW7471B	Mercury	mg/kg	0.014	<b>0.039 [0.01]</b>	<b>0.029 [0.011] J</b>	<b>0.046 [0.01]</b>	<b>0.024 [0.011] J</b>	<b>0.064 [0.011]</b>	<b>0.047 [0.01]</b>
SW8081B	4,4'-DDD	mg/kg	0.013	<b>0.12 [0.00055]</b>	<b>0.33 [0.00055]</b>	0.0006 [0.000055]	0.0018 [0.000054]	0.0025 [0.000055]	0.006 [0.000054]
SW8081B	4,4'-DDE	mg/kg	0.0093	<b>0.049 [0.00063]</b>	<b>0.14 [0.00063]</b>	0.00084 [0.000063]	0.0027 [0.000063]	0.0031 [0.000063]	0.0073 [0.000062]
SW8081B	4,4'-DDT	mg/kg	0.0093	<b>0.016 [0.00063]</b>	<b>0.041 [0.00063]</b>	0.00011 [0.000063] J	0.00013 [0.000062] J	0.00015 [0.000063] J	0.00036 [0.000062] J
SW8081B	Aldrin	mg/kg	0.00019	ND [0.00075]	ND [0.00075]	ND [0.00075]	ND [0.00074]	ND [0.00075]	ND [0.00074]
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.00068]	ND [0.00068]	ND [0.00068]	ND [0.00068]	ND [0.00068]	ND [0.00067]
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.00083]	0.0012 [0.00083] J	ND [0.00083]	ND [0.00082]	ND [0.00083]	ND [0.00081]
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.0011]	ND [0.0011]	ND [0.0011]	ND [0.0011]	ND [0.0011]	ND [0.0011]
SW8081B	delta-BHC	mg/kg	-	ND [0.00064]	ND [0.00064]	ND [0.00064]	ND [0.00064]	ND [0.00064]	ND [0.00063]
SW8081B	Dieldrin	mg/kg	0.0002	ND [0.0007]	ND [0.0007]	ND [0.0007]	ND [0.00069]	ND [0.0007]	ND [0.00069]
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.00079]	ND [0.00079]	ND [0.00079]	ND [0.00078]	ND [0.00079]	ND [0.00077]
SW8081B	Endosulfan II	mg/kg	0.811	ND [0.00074]	0.00089 [0.00074] J	ND [0.00074]	ND [0.00073]	ND [0.00074]	0.00027 [0.00073] J
SW8081B	Endosulfan sulfate	mg/kg	0.811	0.002 [0.00044] J	0.0041 [0.00044] J	ND [0.00044]	0.00071 [0.00043] J	0.00055 [0.00044] J	0.00028 [0.00043] J
SW8081B	Endrin	mg/kg	0.041	0.0041 [0.00081] J	0.011 [0.00081]	ND [0.00081]	0.00017 [0.00008] J	0.00013 [0.000081] J	0.00048 [0.00008]
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.00081]	ND [0.00081]	ND [0.00081]	ND [0.00008]	ND [0.00081]	ND [0.00008]
SW8081B	Endrin ketone	mg/kg	-	0.012 [0.00065]	0.022 [0.00065]	0.00012 [0.000065] J	0.00014 [0.000065] J	ND [0.00065]	0.0044 [0.000064]
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.00073]	ND [0.00073]	ND [0.00073]	ND [0.00073]	ND [0.00073]	ND [0.00072]
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.00082]	0.0028 [0.00082] J	ND [0.00082]	ND [0.00081]	ND [0.00082]	ND [0.00081]
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.00093]	ND [0.00093]	ND [0.00093]	ND [0.00092]	ND [0.00093]	ND [0.00091]
SW8081B	Heptachlor epoxide	mg/kg	0.00035	<b>0.0021 [0.00081] J</b>	<b>0.001 [0.00081] J</b>	ND [0.00081]	ND [0.00008]	ND [0.00081]	0.00013 [0.00008] J
SW8081B	Methoxychlor	mg/kg	0.676	ND [0.00087]	0.013 [0.00087]	ND [0.00087]	0.00012 [0.000086] J	0.00025 [0.000087] J	0.00084 [0.000086]
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.028]	ND [0.028]	ND [0.028]	ND [0.028]	ND [0.028]	ND [0.027]
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00061]	ND [0.00061]	ND [0.00062]	ND [0.00061]	ND [0.00061]	ND [0.00062]
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.00078]	ND [0.00078]	ND [0.00079]	ND [0.00078]	ND [0.00078]	ND [0.00079]
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.0007]	ND [0.0007]	ND [0.00071]	ND [0.0007]	ND [0.0007]	ND [0.00071]
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00067]	ND [0.00067]	ND [0.00067]	ND [0.00067]	ND [0.00067]	ND [0.00067]
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]	ND [0.00039]
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	<b>0.12 [0.00058]</b>	ND [0.00058]	<b>0.0028 [0.00059] J</b>	<b>0.0058 [0.00058]</b>	<b>0.01 [0.00058]</b>	ND [0.00059]
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.23 [0.00058]</b>	<b>0.84 [0.00058]</b>	0.0026 [0.00059] J	<b>0.01 [0.00058]</b>	<b>0.016 [0.00058]</b>	<b>0.022 [0.00059]</b>

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E (Italics)* = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).



**Fish Tissue Analytical Results  
Eielson AFB, Alaska**

				Location ID	PSAG21-F PS-AG-21-Fillet 18159-41 8/1/2012 TF STLP
				Sample ID	
				Lab Sample ID	
				Date Collected	
				Matrix Laboratory	
Method	Analyte	Units	Project Action Limit		
BDTL	Total Lipids	Percent	-	0.51 [0.03]	
SW7471B	Mercury	mg/kg	0.014	<b>0.095 [0.011]</b>	
SW8081B	4,4'-DDD	mg/kg	0.013	0.00079 [0.000054]	
SW8081B	4,4'-DDE	mg/kg	0.0093	0.0053 [0.000063]	
SW8081B	4,4'-DDT	mg/kg	0.0093	0.00018 [0.000062] J	
SW8081B	Aldrin	mg/kg	0.00019	ND [0.000074]	
SW8081B	alpha-BHC	mg/kg	0.0005	ND [0.000068]	
SW8081B	alpha-Chlordane	mg/kg	0.009	ND [0.000082]	
SW8081B	beta-BHC	mg/kg	0.0018	ND [0.00011]	
SW8081B	delta-BHC	mg/kg	-	ND [0.000064]	
SW8081B	Dieldrin	mg/kg	0.0002	0.000098 [0.000069] J	
SW8081B	Endosulfan I	mg/kg	0.811	ND [0.000078]	
SW8081B	Endosulfan II	mg/kg	0.811	ND [0.000073]	
SW8081B	Endosulfan sulfate	mg/kg	0.811	0.000073 [0.000043] J	
SW8081B	Endrin	mg/kg	0.041	0.00013 [0.00008] J	
SW8081B	Endrin aldehyde	mg/kg	0.041	ND [0.00008]	
SW8081B	Endrin ketone	mg/kg	-	ND [0.000065]	
SW8081B	gamma-BHC	mg/kg	0.0029	ND [0.000073]	
SW8081B	gamma-Chlordane	mg/kg	0.009	ND [0.000081]	
SW8081B	Heptachlor	mg/kg	0.0007	ND [0.000092]	
SW8081B	Heptachlor epoxide	mg/kg	0.00035	ND [0.00008]	
SW8081B	Methoxychlor	mg/kg	0.676	0.00029 [0.000086] J	
SW8081B	Toxaphene	mg/kg	0.0029	ND [0.0028]	
SW8082A	PCB-1016 (Aroclor 1016)	mg/kg	0.045	ND [0.00062]	
SW8082A	PCB-1221 (Aroclor 1221)	mg/kg	0.0016	ND [0.0008]	
SW8082A	PCB-1232 (Aroclor 1232)	mg/kg	0.0016	ND [0.00071]	
SW8082A	PCB-1242 (Aroclor 1242)	mg/kg	0.0016	ND [0.00068]	
SW8082A	PCB-1248 (Aroclor 1248)	mg/kg	0.0016	ND [0.00039]	
SW8082A	PCB-1254 (Aroclor 1254)	mg/kg	0.0016	<b>0.026 [0.00059]</b>	
SW8082A	PCB-1260 (Aroclor 1260)	mg/kg	0.00269	<b>0.027 [0.00059]</b>	

**Notes:**

STLP = TestAmerica, Pittsburg, PA

**Bold** = The sample result was greater than or equal to the Project Action Limit (PAL).

*E* (Italics) = The analyte was not detected in the sample and its LOD exceed the PAL.

[ ] = Limit of Detection (LOD)

Data qualifiers are defined in the Data Quality Assessment (DQA).

**EXHIBIT B**  
**Qualified Data Tables**

**Sample Results Qualified E due to Nondetect Results and the LOD Exceeding the Project Action Limit**

Sample Identification	Analyte	Result	LOD (mg/kg)	Project Action Limit (mg/kg)	Qualifier
GS-AG-01-FILLET	Aldrin	ND	0.00073	0.00019	E
GS-AG-01-FILLET	alpha-BHC	ND	0.00067	0.0005	E
GS-AG-01-FILLET	Dieldrin	ND	0.00068	0.0002	E
GS-AG-01-FILLET	Heptachlor epoxide	ND	0.00079	0.00035	E
GS-AG-01-FILLET	Heptachlor	ND	0.00091	0.0007	E
GS-AG-01-FILLET	Toxaphene	ND	0.027	0.0029	E
GS-AG-01-WHOLE	Aldrin	ND	0.00073	0.00019	E
GS-AG-01-WHOLE	alpha-BHC	ND	0.00066	0.0005	E
GS-AG-01-WHOLE	Heptachlor epoxide	ND	0.00079	0.00035	E
GS-AG-01-WHOLE	Heptachlor	ND	0.0009	0.0007	E
GS-AG-01-WHOLE	Toxaphene	ND	0.027	0.0029	E
GS-AG-02-FILLET	Aldrin	ND	0.00073	0.00019	E
GS-AG-02-FILLET	alpha-BHC	ND	0.00066	0.0005	E
GS-AG-02-FILLET	Dieldrin	ND	0.00068	0.0002	E
GS-AG-02-FILLET	Heptachlor	ND	0.0009	0.0007	E
GS-AG-02-FILLET	Toxaphene	ND	0.027	0.0029	E
GS-AG-02-WHOLE	Aldrin	ND	0.00074	0.00019	E
GS-AG-02-WHOLE	alpha-BHC	ND	0.00067	0.0005	E
GS-AG-02-WHOLE	Dieldrin	ND	0.00069	0.0002	E
GS-AG-02-WHOLE	Heptachlor	ND	0.00091	0.0007	E
GS-AG-02-WHOLE	Toxaphene	ND	0.027	0.0029	E
GS-AG-03-FILLET	Aldrin	ND	0.00075	0.00019	E
GS-AG-03-FILLET	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-03-FILLET	Dieldrin	ND	0.0007	0.0002	E
GS-AG-03-FILLET	Heptachlor	ND	0.00093	0.0007	E
GS-AG-03-FILLET	Toxaphene	ND	0.028	0.0029	E
GS-AG-03-WHOLE	Aldrin	ND	0.00073	0.00019	E
GS-AG-03-WHOLE	alpha-BHC	ND	0.00067	0.0005	E
GS-AG-03-WHOLE	Heptachlor epoxide	ND	0.00079	0.00035	E
GS-AG-03-WHOLE	Heptachlor	ND	0.00091	0.0007	E
GS-AG-03-WHOLE	Toxaphene	ND	0.027	0.0029	E
GS-AG-04-FILLET	Aldrin	ND	0.00073	0.00019	E
GS-AG-04-FILLET	alpha-BHC	ND	0.00066	0.0005	E
GS-AG-04-FILLET	Dieldrin	ND	0.00068	0.0002	E
GS-AG-04-FILLET	Heptachlor epoxide	ND	0.00079	0.00035	E
GS-AG-04-FILLET	Heptachlor	ND	0.0009	0.0007	E
GS-AG-04-FILLET	Toxaphene	ND	0.027	0.0029	E
GS-AG-04-WHOLE	Aldrin	ND	0.00074	0.00019	E
GS-AG-04-WHOLE	alpha-BHC	ND	0.00067	0.0005	E
GS-AG-04-WHOLE	Heptachlor epoxide	ND	0.0008	0.00035	E
GS-AG-04-WHOLE	Heptachlor	ND	0.00091	0.0007	E
GS-AG-04-WHOLE	Toxaphene	ND	0.027	0.0029	E
GS-AG-05-FILLET	Aldrin	ND	0.00073	0.00019	E
GS-AG-05-FILLET	alpha-BHC	ND	0.00066	0.0005	E
GS-AG-05-FILLET	Dieldrin	ND	0.00068	0.0002	E
GS-AG-05-FILLET	Heptachlor epoxide	ND	0.00079	0.00035	E
GS-AG-05-FILLET	Heptachlor	ND	0.0009	0.0007	E
GS-AG-05-FILLET	Toxaphene	ND	0.027	0.0029	E
GS-AG-05-WHOLE	Aldrin	ND	0.00073	0.00019	E
GS-AG-05-WHOLE	alpha-BHC	ND	0.00066	0.0005	E

**Sample Results Qualified E due to Nondetect Results and the LOD Exceeding the Project Action Limit**

Sample Identification	Analyte	Result	LOD (mg/kg)	Project Action Limit (mg/kg)	Qualifier
GS-AG-05-WHOLE	Dieldrin	ND	0.00068	0.0002	E
GS-AG-05-WHOLE	Heptachlor epoxide	ND	0.00079	0.00035	E
GS-AG-05-WHOLE	Heptachlor	ND	0.0009	0.0007	E
GS-AG-05-WHOLE	Toxaphene	ND	0.027	0.0029	E
GS-AG-06-FILLET	Aldrin	ND	0.00074	0.00019	E
GS-AG-06-FILLET	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-06-FILLET	Dieldrin	ND	0.00069	0.0002	E
GS-AG-06-FILLET	Heptachlor epoxide	ND	0.0008	0.00035	E
GS-AG-06-FILLET	Heptachlor	ND	0.00092	0.0007	E
GS-AG-06-FILLET	Toxaphene	ND	0.028	0.0029	E
GS-AG-06-WHOLE	Aldrin	ND	0.00073	0.00019	E
GS-AG-06-WHOLE	alpha-BHC	ND	0.00067	0.0005	E
GS-AG-06-WHOLE	Heptachlor	ND	0.00091	0.0007	E
GS-AG-06-WHOLE	Toxaphene	ND	0.027	0.0029	E
GS-AG-07-FILLET	Aldrin	ND	0.00074	0.00019	E
GS-AG-07-FILLET	alpha-BHC	ND	0.00067	0.0005	E
GS-AG-07-FILLET	Dieldrin	ND	0.00069	0.0002	E
GS-AG-07-FILLET	Heptachlor epoxide	ND	0.0008	0.00035	E
GS-AG-07-FILLET	Heptachlor	ND	0.00091	0.0007	E
GS-AG-07-FILLET	Toxaphene	ND	0.027	0.0029	E
GS-AG-07-WHOLE	Aldrin	ND	0.00074	0.00019	E
GS-AG-07-WHOLE	alpha-BHC	ND	0.00067	0.0005	E
GS-AG-07-WHOLE	Dieldrin	ND	0.00069	0.0002	E
GS-AG-07-WHOLE	Heptachlor	ND	0.00091	0.0007	E
GS-AG-07-WHOLE	Toxaphene	ND	0.027	0.0029	E
GS-AG-08-FILLET	Aldrin	ND	0.00073	0.00019	E
GS-AG-08-FILLET	alpha-BHC	ND	0.00067	0.0005	E
GS-AG-08-FILLET	Dieldrin	ND	0.00068	0.0002	E
GS-AG-08-FILLET	Heptachlor	ND	0.00091	0.0007	E
GS-AG-08-FILLET	Toxaphene	ND	0.027	0.0029	E
GS-AG-08-WHOLE	Aldrin	ND	0.00073	0.00019	E
GS-AG-08-WHOLE	alpha-BHC	ND	0.00067	0.0005	E
GS-AG-08-WHOLE	Heptachlor	ND	0.00091	0.0007	E
GS-AG-08-WHOLE	Toxaphene	ND	0.027	0.0029	E
GS-AG-09-FILLET	Aldrin	ND	0.00075	0.00019	E
GS-AG-09-FILLET	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-09-FILLET	Dieldrin	ND	0.0007	0.0002	E
GS-AG-09-FILLET	Heptachlor epoxide	ND	0.00081	0.00035	E
GS-AG-09-FILLET	Heptachlor	ND	0.00093	0.0007	E
GS-AG-09-FILLET	Toxaphene	ND	0.028	0.0029	E
GS-AG-09-WHOLE	Aldrin	ND	0.00074	0.00019	E
GS-AG-09-WHOLE	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-09-WHOLE	Heptachlor	ND	0.00092	0.0007	E
GS-AG-09-WHOLE	Toxaphene	ND	0.028	0.0029	E
GS-AG-10-FILLET	Aldrin	ND	0.00074	0.00019	E
GS-AG-10-FILLET	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-10-FILLET	Dieldrin	ND	0.00069	0.0002	E
GS-AG-10-FILLET	Heptachlor	ND	0.00092	0.0007	E
GS-AG-10-FILLET	Toxaphene	ND	0.028	0.0029	E
GS-AG-10-WHOLE	Aldrin	ND	0.00074	0.00019	E

**Sample Results Qualified E due to Nondetect Results and the LOD Exceeding the Project Action Limit**

Sample Identification	Analyte	Result	LOD (mg/kg)	Project Action Limit (mg/kg)	Qualifier
GS-AG-10-WHOLE	alpha-BHC	ND	0.00067	0.0005	E
GS-AG-10-WHOLE	Heptachlor	ND	0.00091	0.0007	E
GS-AG-10-WHOLE	Toxaphene	ND	0.027	0.0029	E
GS-AG-11-FILLET	Aldrin	ND	0.00075	0.00019	E
GS-AG-11-FILLET	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-11-FILLET	Heptachlor	ND	0.00093	0.0007	E
GS-AG-11-FILLET	Toxaphene	ND	0.028	0.0029	E
GS-AG-11-WHOLE	Aldrin	ND	0.0037	0.00019	E
GS-AG-11-WHOLE	alpha-BHC	ND	0.0034	0.0005	E
GS-AG-11-WHOLE	beta-BHC	ND	0.0054	0.0018	E
GS-AG-11-WHOLE	gamma-BHC	ND	0.0036	0.0029	E
GS-AG-11-WHOLE	Heptachlor	ND	0.0046	0.0007	E
GS-AG-11-WHOLE	Toxaphene	ND	0.14	0.0029	E
GS-AG-12-FILLET	Aldrin	ND	0.00074	0.00019	E
GS-AG-12-FILLET	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-12-FILLET	Heptachlor epoxide	ND	0.0008	0.00035	E
GS-AG-12-FILLET	Heptachlor	ND	0.00092	0.0007	E
GS-AG-12-FILLET	Toxaphene	ND	0.028	0.0029	E
GS-AG-12-WHOLE	Aldrin	ND	0.00075	0.00019	E
GS-AG-12-WHOLE	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-12-WHOLE	Heptachlor epoxide	ND	0.00081	0.00035	E
GS-AG-12-WHOLE	Heptachlor	ND	0.00093	0.0007	E
GS-AG-12-WHOLE	Toxaphene	ND	0.028	0.0029	E
GS-AG-13-FILLET	Aldrin	ND	0.00037	0.00019	E
GS-AG-13-FILLET	Heptachlor epoxide	ND	0.00041	0.00035	E
GS-AG-13-FILLET	Toxaphene	ND	0.014	0.0029	E
GS-AG-13-WHOLE	Aldrin	ND	0.00074	0.00019	E
GS-AG-13-WHOLE	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-13-WHOLE	Heptachlor epoxide	ND	0.0008	0.00035	E
GS-AG-13-WHOLE	Heptachlor	ND	0.00092	0.0007	E
GS-AG-13-WHOLE	Toxaphene	ND	0.028	0.0029	E
GS-AG-14-FILLET	Aldrin	ND	0.00074	0.00019	E
GS-AG-14-FILLET	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-14-FILLET	Heptachlor	ND	0.00092	0.0007	E
GS-AG-14-FILLET	Toxaphene	ND	0.028	0.0029	E
GS-AG-14-WHOLE	Aldrin	ND	0.0037	0.00019	E
GS-AG-14-WHOLE	alpha-BHC	ND	0.0034	0.0005	E
GS-AG-14-WHOLE	beta-BHC	ND	0.0054	0.0018	E
GS-AG-14-WHOLE	gamma-BHC	ND	0.0037	0.0029	E
GS-AG-14-WHOLE	Heptachlor epoxide	ND	0.0041	0.00035	E
GS-AG-14-WHOLE	Heptachlor	ND	0.0046	0.0007	E
GS-AG-14-WHOLE	Toxaphene	ND	0.14	0.0029	E
GS-AG-15-FILLET	Aldrin	ND	0.00075	0.00019	E
GS-AG-15-FILLET	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-15-FILLET	Heptachlor epoxide	ND	0.00081	0.00035	E
GS-AG-15-FILLET	Heptachlor	ND	0.00093	0.0007	E
GS-AG-15-FILLET	Toxaphene	ND	0.028	0.0029	E
GS-AG-15-WHOLE	Aldrin	ND	0.00075	0.00019	E
GS-AG-15-WHOLE	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-15-WHOLE	Heptachlor	ND	0.00093	0.0007	E

**Sample Results Qualified E due to Nondetect Results and the LOD Exceeding the Project Action Limit**

Sample Identification	Analyte	Result	LOD (mg/kg)	Project Action Limit (mg/kg)	Qualifier
GS-AG-15-WHOLE	Toxaphene	ND	0.028	0.0029	E
GS-AG-16-FILLET	Aldrin	ND	0.00075	0.00019	E
GS-AG-16-FILLET	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-16-FILLET	Heptachlor epoxide	ND	0.00081	0.00035	E
GS-AG-16-FILLET	Heptachlor	ND	0.00093	0.0007	E
GS-AG-16-FILLET	Toxaphene	ND	0.028	0.0029	E
GS-AG-16-WHOLE	Aldrin	ND	0.00074	0.00019	E
GS-AG-16-WHOLE	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-16-WHOLE	Heptachlor	ND	0.00092	0.0007	E
GS-AG-16-WHOLE	Toxaphene	ND	0.028	0.0029	E
GS-AG-17-FILLET	Aldrin	ND	0.00074	0.00019	E
GS-AG-17-FILLET	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-17-FILLET	Heptachlor	ND	0.00092	0.0007	E
GS-AG-17-FILLET	Toxaphene	ND	0.028	0.0029	E
GS-AG-17-WHOLE	Aldrin	ND	0.0037	0.00019	E
GS-AG-17-WHOLE	alpha-BHC	ND	0.0034	0.0005	E
GS-AG-17-WHOLE	beta-BHC	ND	0.0054	0.0018	E
GS-AG-17-WHOLE	gamma-BHC	ND	0.0037	0.0029	E
GS-AG-17-WHOLE	Heptachlor	ND	0.0046	0.0007	E
GS-AG-17-WHOLE	Toxaphene	ND	0.14	0.0029	E
GS-AG-18-FILLET	Aldrin	ND	0.00074	0.00019	E
GS-AG-18-FILLET	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-18-FILLET	Heptachlor epoxide	ND	0.0008	0.00035	E
GS-AG-18-FILLET	Heptachlor	ND	0.00092	0.0007	E
GS-AG-18-FILLET	Toxaphene	ND	0.028	0.0029	E
GS-AG-18-WHOLE	Aldrin	ND	0.00075	0.00019	E
GS-AG-18-WHOLE	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-18-WHOLE	Heptachlor	ND	0.00093	0.0007	E
GS-AG-18-WHOLE	Toxaphene	ND	0.028	0.0029	E
GS-AG-19-FILLET	Aldrin	ND	0.00075	0.00019	E
GS-AG-19-FILLET	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-19-FILLET	Heptachlor	ND	0.00093	0.0007	E
GS-AG-19-FILLET	Toxaphene	ND	0.028	0.0029	E
GS-AG-19-WHOLE	Aldrin	ND	0.00075	0.00019	E
GS-AG-19-WHOLE	alpha-BHC	ND	0.00068	0.0005	E
GS-AG-19-WHOLE	Heptachlor epoxide	ND	0.00081	0.00035	E
GS-AG-19-WHOLE	Heptachlor	ND	0.00093	0.0007	E
GS-AG-19-WHOLE	Toxaphene	ND	0.028	0.0029	E
GS-AG-20-WHOLE	Aldrin	ND	0.00074	0.00019	E
GS-AG-20-WHOLE	alpha-BHC	ND	0.00067	0.0005	E
GS-AG-20-WHOLE	Heptachlor epoxide	ND	0.0008	0.00035	E
GS-AG-20-WHOLE	Heptachlor	ND	0.00091	0.0007	E
GS-AG-20-WHOLE	Toxaphene	ND	0.027	0.0029	E
CR-AG-07-Fillet	Mercury	ND	0.011	0.014	E
CR-AG-07-Whole	Mercury	ND	0.011	0.014	E
PS-AG-03-Whole	Aldrin	ND	0.00073	0.00019	E
PS-AG-03-Whole	alpha-BHC	ND	0.00067	0.0005	E
PS-AG-03-Whole	Dieldrin	ND	0.00068	0.0002	E
PS-AG-03-Whole	Heptachlor	ND	0.00091	0.0007	E
PS-AG-03-Whole	Toxaphene	ND	0.027	0.0029	E

**Sample Results Qualified E due to Nondetect Results and the LOD Exceeding the Project Action Limit**

<b>Sample Identification</b>	<b>Analyte</b>	<b>Result</b>	<b>LOD (mg/kg)</b>	<b>Project Action Limit (mg/kg)</b>	<b>Qualifier</b>
PS-AG-04-Fillet	Aldrin	ND	0.000075	0.00019	E
PS-AG-12-Whole	Toxaphene	ND	0.0055	0.0029	E
PS-AG-18-Fillet	Aldrin	ND	0.00075	0.00019	E
PS-AG-18-Fillet	alpha-BHC	ND	0.00068	0.0005	E
PS-AG-18-Fillet	Dieldrin	ND	0.0007	0.0002	E
PS-AG-18-Fillet	Heptachlor	ND	0.00093	0.0007	E
PS-AG-18-Fillet	Toxaphene	ND	0.028	0.0029	E
PS-AG-18-Whole	Aldrin	ND	0.00075	0.00019	E
PS-AG-18-Whole	alpha-BHC	ND	0.00068	0.0005	E
PS-AG-18-Whole	Dieldrin	ND	0.0007	0.0002	E
PS-AG-18-Whole	Heptachlor	ND	0.00093	0.0007	E
PS-AG-18-Whole	Toxaphene	ND	0.028	0.0029	E

**Sample Results Qualified B due to Method Blank Contamination**

<b>Sample Identification</b>	<b>Analyte</b>	<b>Result (mg/kg)</b>	<b>Qualifiers</b>	<b>Analytical Batch</b>
Method Blank	Methoxychlor	0.0000475	-	61868
CK-AG-01-Fillet	Methoxychlor	0.00011	J, B	61868
CK-AG-05-Whole	Methoxychlor	0.00011	J, B	61868
CK-AG-06-Whole	Methoxychlor	0.00017	J, B	61868
CK-AG-07-Fillet	Methoxychlor	0.00014	J, B	61868
CK-AG-07-Whole	Methoxychlor	0.00015	J, B	61868
CK-AG-08-Fillet	Methoxychlor	0.00014	J, B	61868
CK-AG-08-Whole	Methoxychlor	0.00017	J, B	61868
CK-AG-09-Fillet	Methoxychlor	0.00017	J, B	61868
CK-AG-09-Whole	Methoxychlor	0.00013	J, B	61868
Method Blank	Methoxychlor	0.0000561	-	62279
CR-AG-20-Fillet	Methoxychlor	0.00015	J, B	62279
PS-AG-01-Fillet	Methoxychlor	0.00015	J, B	62279
PS-AG-04-Whole	Methoxychlor	0.00024	J, B	62279
PS-AG-05-Whole	Methoxychlor	0.00016	J, B	62279
PS-AG-06-Fillet	Methoxychlor	0.000087	J, B	62279
PS-AG-06-Whole	Methoxychlor	0.00017	J, B	62279
PS-AG-07-Fillet	Methoxychlor	0.00011	J, B	62279
PS-AG-07-Whole	Methoxychlor	0.00014	J, B	62279
PS-AG-09-Fillet	Methoxychlor	0.00019	J, B	62279



**EXHIBIT C**  
**Alaska Department of Environmental Conservation**  
**Laboratory Data Review Checklists**

**Laboratory Data Review Checklist**

**Completed by:**

**Title:**  **Date:**

**CS Report Name:**  **Report Date:**

**Consultant Firm:**

**Laboratory Name:**  **Laboratory Report Number:**

**ADEC File Number:**  **ADEC RecKey Number:**

**1. Laboratory**

a. Did an ADEC CS-approved laboratory receive and perform all of the submitted sample analyses?

Yes  No  NA (Please explain.) Comments:

Samples were sent to TestAmerica in Pittsburg, PA for the fish tissue analysis. The lab is DoD ELAP accredited, but does not hold ELAP certification for tissue analysis, nor do they hold certification for Alaska.

b. If the samples were transferred to another “network” laboratory or sub-contracted to an alternate laboratory, was the laboratory performing the analyses ADEC CS approved?

Yes  No  NA (Please explain.) Comments:

**2. Chain of Custody (CoC)**

a. CoC information completed, signed, and dated (including released/received by)?

Yes  No  NA (Please explain.) Comments:

b. Correct Analyses requested?

Yes  No  NA (Please explain.) Comments:

**3. Laboratory Sample Receipt Documentation**

a. Sample/cooler temperature documented and within range at receipt ( $4^{\circ} \pm 2^{\circ} \text{C}$ )?

Yes  No  NA (Please explain.) Comments:

Four coolers were received at temperatures of 2.8°C, 2.6°C, 2.6°C, and 1.9°C. The coolers contained all the fish tissue samples, including those in 3 other SDGs. Temperatures are indistinguishable between coolers and their associated samples, and therefore all four cooler temperatures have been listed.

b. Sample preservation acceptable – acidified waters, Methanol preserved VOC soil (GRO, BTEX, Volatile Chlorinated Solvents, etc.)?

Yes  No  NA (Please explain.) Comments:

c. Sample condition documented – broken, leaking (Methanol), zero headspace (VOC vials)?

Yes  No  NA (Please explain.) Comments:

d. If there were any discrepancies, were they documented? For example, incorrect sample containers/preservation, sample temperature outside of acceptable range, insufficient or missing samples, etc.?

Yes  No  NA (Please explain.) Comments:

The shipping temperature was noted. Samples were received in good condition.

ICVs 180-62115/27 and 180-62111/27 failed high for Aroclor 1232.

e. Data quality or usability affected? (Please explain.)

Comments:

The fish samples are unaffected by the shipping temperature. Upon arrival at the lab, the samples were stored in the freezer.

All associated sample results associated with the ICVs were nondetect and biased high; therefore, flags were not assigned and data quality and usability are minimally affected.

**4. Case Narrative**

a. Present and understandable?

Yes  No  NA (Please explain.) Comments:

b. Discrepancies, errors or QC failures identified by the lab?

Yes  No  NA (Please explain.) Comments:

c. Were all corrective actions documented?

Yes  No  NA (Please explain.) Comments:

d. What is the effect on data quality/usability according to the case narrative?

Comments:

See respective sections of this checklist for data quality and usability issues.

## 5. Samples Results

a. Correct analyses performed/reported as requested on COC?

Yes  No  NA (Please explain.)

Comments:

b. All applicable holding times met?

Yes  No  NA (Please explain.)

Comments:

Samples were analyzed outside of holding time for each analysis. Samples were shipped in August 2012 by EA Engineering and placed on hold at the time of arrival at the laboratory. Samples were released for analysis by Jacobs (after acquiring the contract) in January 2013, well over 100 days past holding time. Samples were placed in frozen storage between those months. The analyses are clearly out of hold, but sample data was not flagged because the holding time issue would apply to all sample results. It is uncertain whether being frozen would extend the applicable hold time. It is also unlikely these analytes (such as mercury and PCBs) would degrade or be affected leading to a potential low bias. The data was considered usable for the purposes of this project.

c. All soils reported on a dry weight basis?

Yes  No  NA (Please explain.)

Comments:

Only fish tissue was analyzed in this SDG.

d. Are the reported PQLs less than the Cleanup Level or the minimum required detection level for the project?

Yes  No  NA (Please explain.)

Comments:

Aldrin, alpha-BHC, beta-BHC, dieldrin, gamma-BHC, heptachlor, heptachlor epoxide, and toxaphene in one or more samples had nondetect results and LODs greater than the project action limit (PAL).

e. Data quality or usability affected?

Comments:

Data is considered usable for the purposes of this project.

## 6. QC Samples

a. Method Blank

i. One method blank reported per matrix, analysis and 20 samples?

Yes  No  NA (Please explain.)

Comments:

ii. All method blank results less than PQL?

Yes  No  NA (Please explain.)

Comments:

iii. If above PQL, what samples are affected?

Yes  No  NA (Please explain.)

Comments:

Method blank recoveries met criteria.

iv. Do the affected sample(s) have data flags and if so, are the data flags clearly defined?

Yes  No  NA (Please explain.)

Comments:

Method blank recoveries met criteria.

v. Data quality or usability affected? (please explain)

Comments:

Data quality and usability are not affected.

b. Laboratory Control Sample/Duplicate (LCS/LCSD)

i. Organics – One LCS/LCSD reported per matrix, analysis and 20 samples? (LCS/LCSD required per AK methods, LCS required per SW846)

Yes  No  NA (Please explain.)

Comments:

Not all laboratory batches had an associated LCSD. All batches had either an LCS/LCSD pair or an LCS and MS/MSD pair associated with it.

ii. Metals/Inorganics – one LCS and one sample duplicate reported per matrix, analysis and 20 samples?

Yes  No  NA (Please explain.)

Comments:

iii. Accuracy – All percent recoveries (%R) reported and within method or laboratory limits? And project specified DQOs, if applicable. (AK Petroleum methods: AK101 60%-120%, AK102 75%-125%, AK103 60%-120%; all other analyses see the laboratory QC pages)

Yes  No  NA (Please explain.)

Comments:

GS-AG-10-FILLET MS/MSD recoveries for 4,4'-DDD; 4,4'-DDE; dieldrin; endrin aldehyde; endrin ketone; heptachlor epoxide; and methoxychlor were outside of QC criteria.

GS-AG-10-WHOLE MS/MSD recoveries for Aroclor 1260 were outside of QC criteria.

iv. Precision – All relative percent differences (RPD) reported and less than method or laboratory limits? And project specified DQOs, if applicable. RPD reported from LCS/LCSD, MS/MSD, and or sample/sample duplicate. (AK Petroleum methods 20%; all other analyses see the laboratory QC pages)

Yes  No  NA (Please explain.)

Comments:

GS-AG-10-FILLET MS/MSD RPD for the following analytes were outside of QC criteria: 4,4'-DDD, 4,4'-DDE, dieldrin, endrin ketone, and heptachlor epoxide.

v. If %R or RPD is outside of acceptable limits, what samples are affected?

Comments:

GS-AG-10-FILLET, GS-AG-10-WHOLE

vi. Do the affected sample(s) have data flags? If so, are the data flags clearly defined?

Yes  No  NA (Please explain.) Comments:

All MS/MSD recoveries had dilution factors greater than or equal to 5.

vii. Data quality or usability affected? (Use comment box to explain.)

Comments:

Data quality and usability are minimally affected. The large dilution factors and RPDs outside of criteria suggest potential matrix interference.

c. Surrogates – Organics Only

i. Are surrogate recoveries reported for organic analyses – field, QC and laboratory samples?

Yes  No  NA (Please explain.) Comments:

ii. Accuracy – All percent recoveries (%R) reported and within method or laboratory limits? And project specified DQOs, if applicable. (AK Petroleum methods 50-150 %R; all other analyses see the laboratory report pages)

Yes  No  NA (Please explain.) Comments:

The surrogates decachlorobiphenyl and tetrachlorometaxylene for SW8081 had low recoveries for multiple samples and the surrogate decachlorobiphenyl for both SW8081 and SW8082 had high recoveries for multiple samples.

iii. Do the sample results with failed surrogate recoveries have data flags? If so, are the data flags clearly defined?

Yes  No  NA (Please explain.) Comments:

All affected samples had dilution factors greater than or equal to 5; therefore, flags were not assigned.

iv. Data quality or usability affected? (Use the comment box to explain.)

Comments:

Data quality and usability are not affected.

d. Trip blank – Volatile analyses only (GRO, BTEX, Volatile Chlorinated Solvents, etc.):  
Water and Soil

i. One trip blank reported per matrix, analysis and for each cooler containing volatile samples? (If not, enter explanation below.)

Yes  No  NA (Please explain.) Comments:

Trip blanks are not associated with SW7471, SW8081, and SW8082 analyses.

ii. Is the cooler used to transport the trip blank and VOA samples clearly indicated on the COC? (If not, a comment explaining why must be entered below)

Yes  No  NA (Please explain.) Comments:

N/A

iii. All results less than PQL?

Yes  No  NA (Please explain.)

Comments:

N/A

iv. If above PQL, what samples are affected?

Comments:

N/A

v. Data quality or usability affected? (Please explain.)

Comments:

N/A

e. Field Duplicate

i. One field duplicate submitted per matrix, analysis and 10 project samples?

Yes  No  NA (Please explain.)

Comments:

Field duplicates were not collected due to the nature of the samples (fish).

ii. Submitted blind to lab?

Yes  No  NA (Please explain.)

Comments:

Field duplicates were not collected.

iii. Precision – All relative percent differences (RPD) less than specified DQOs?  
(Recommended: 30% water, 50% soil)

$$\text{RPD (\%)} = \text{Absolute value of: } \frac{(R_1 - R_2)}{((R_1 + R_2)/2)} \times 100$$

Where  $R_1$  = Sample Concentration  
 $R_2$  = Field Duplicate Concentration

Yes  No  NA (Please explain.)

Comments:

Field duplicates were not collected

iv. Data quality or usability affected? (Use the comment box to explain why or why not.)

Comments:

Data quality and usability are not affected.

f. Decontamination or Equipment Blank (If not used explain why).

Yes  No  NA (Please explain.)

Comments:

The fish were sent to the lab whole. Due to this nature of the samples, decontamination/equipment blanks were not collected.

i. All results less than PQL?

Yes  No  NA (Please explain.)

Comments:

N/A

ii. If above PQL, what samples are affected?

Comments:

N/A

iii. Data quality or usability affected? (Please explain.)

Comments:

N/A

**7. Other Data Flags/Qualifiers (ACOE, AFCEE, Lab-Specific, etc.)**

a. Defined and appropriate?

Yes  No  NA (Please explain.)

Comments:



**Laboratory Data Review Checklist**

**Completed by:**

**Title:**  **Date:**

**CS Report Name:**  **Report Date:**

**Consultant Firm:**

**Laboratory Name:**  **Laboratory Report Number:**

**ADEC File Number:**  **ADEC RecKey Number:**

**1. Laboratory**

a. Did an ADEC CS-approved laboratory receive and perform all of the submitted sample analyses?

Yes  No  NA (Please explain.) Comments:

Samples were sent to TestAmerica in Pittsburg, PA for the fish tissue analysis. The lab is DoD ELAP accredited, but does not hold ELAP certification for tissue analysis, nor do they hold certification for Alaska.

b. If the samples were transferred to another “network” laboratory or sub-contracted to an alternate laboratory, was the laboratory performing the analyses ADEC CS approved?

Yes  No  NA (Please explain.) Comments:

Samples were not transferred.

**2. Chain of Custody (CoC)**

a. CoC information completed, signed, and dated (including released/received by)?

Yes  No  NA (Please explain.) Comments:

b. Correct Analyses requested?

Yes  No  NA (Please explain.) Comments:

**3. Laboratory Sample Receipt Documentation**

a. Sample/cooler temperature documented and within range at receipt ( $4^{\circ} \pm 2^{\circ} \text{C}$ )?

Yes  No  NA (Please explain.) Comments:

Four coolers were received at temperatures of 2.8°C, 2.6°C, 2.6°C, and 1.9°C. The coolers contained all the fish tissue samples, including those in 3 other SDGs. Temperatures are indistinguishable between coolers and their associated samples, and therefore all four cooler temperatures have been listed.

b. Sample preservation acceptable – acidified waters, Methanol preserved VOC soil (GRO, BTEX, Volatile Chlorinated Solvents, etc.)?

Yes  No  NA (Please explain.) Comments:

c. Sample condition documented – broken, leaking (Methanol), zero headspace (VOC vials)?

Yes  No  NA (Please explain.) Comments:

d. If there were any discrepancies, were they documented? For example, incorrect sample containers/preservation, sample temperature outside of acceptable range, insufficient or missing samples, etc.?

Yes  No  NA (Please explain.) Comments:

The shipping temperature was noted. Samples were received in good condition.

e. Data quality or usability affected? (Please explain.)

Comments:

The fish samples are unaffected by the shipping temperature. Upon arrival at the lab, the samples were stored in the freezer.

**4. Case Narrative**

a. Present and understandable?

Yes  No  NA (Please explain.) Comments:

b. Discrepancies, errors or QC failures identified by the lab?

Yes  No  NA (Please explain.) Comments:

c. Were all corrective actions documented?

Yes  No  NA (Please explain.) Comments:

d. What is the effect on data quality/usability according to the case narrative?

Comments:

See respective sections of this checklist for data quality and usability issues.

**5. Samples Results**

a. Correct analyses performed/reported as requested on COC?

Yes  No  NA (Please explain.)

Comments:

b. All applicable holding times met?

Yes  No  NA (Please explain.)

Comments:

Samples were analyzed outside of holding time for each analysis. Samples were shipped in August 2012 by EA Engineering placed on hold at the lab at that time. Samples were released for analysis by Jacobs (after acquiring the contract) in January 2013, well over 100 days past holding time. Samples were placed in frozen storage between those months. The analyses are clearly out of hold, but sample data was not flagged because the holding time issue would apply to all sample results. It is uncertain whether being frozen would extend the applicable hold time. It is also unlikely these analytes (such as mercury and PCBs) would degrade or be affected leading to a potential low bias. The data was considered usable for the purposes of this project.

c. All soils reported on a dry weight basis?

Yes  No  NA (Please explain.)

Comments:

Only fish tissue was analyzed in this SDG.

d. Are the reported PQLs less than the Cleanup Level or the minimum required detection level for the project?

Yes  No  NA (Please explain.)

Comments:

Mercury in samples CR-AG-07-Fillet and CR-AG-07-Whole had nondetect results and LODs higher than the project action limit (PAL).

e. Data quality or usability affected?

Comments:

Data is considered usable for the purposes of this project.

**6. QC Samples**

a. Method Blank

i. One method blank reported per matrix, analysis and 20 samples?

Yes  No  NA (Please explain.)

Comments:

ii. All method blank results less than PQL?

Yes  No  NA (Please explain.)

Comments:

Methoxychlor was detected in method blank 180-62279/1-B.

iii. If above PQL, what samples are affected?

Yes  No  NA (Please explain.)

Comments:

All samples in batch 180-62279 are affected.

iv. Do the affected sample(s) have data flags and if so, are the data flags clearly defined?

Yes  No  NA (Please explain.) Comments:

Only the sample result for methoxychlor in sample CR-AG-20-Fillet was flagged B due to having a detected result within 5 times the method blank concentration.

v. Data quality or usability affected? (please explain)

Comments:

Sample results flagged B are potentially biased high but all B qualified samples are less than the PAL. Data usability is not affected.

b. Laboratory Control Sample/Duplicate (LCS/LCSD)

i. Organics – One LCS/LCSD reported per matrix, analysis and 20 samples? (LCS/LCSD required per AK methods, LCS required per SW846)

Yes  No  NA (Please explain.) Comments:

All batches had either a LCS/LCSD pair or an LCS and MS/MSD pair.

ii. Metals/Inorganics – one LCS and one sample duplicate reported per matrix, analysis and 20 samples?

Yes  No  NA (Please explain.) Comments:

iii. Accuracy – All percent recoveries (%R) reported and within method or laboratory limits? And project specified DQOs, if applicable. (AK Petroleum methods: AK101 60%-120%, AK102 75%-125%, AK103 60%-120%; all other analyses see the laboratory QC pages)

Yes  No  NA (Please explain.) Comments:

CR-AG-02-Fillet MS/MSD for Aroclor 1260 had a high recovery.

CR-AG-02-Fillet MS/MSD for alpha-BHC and endrin aldehyde had low recoveries.

CR-AG-02-Whole MS/MSD for 4,4'-DDD, aldrin, alpha-BHC, beta-BHC, dieldrin, endosulfan I, endosulfan II, endosulfan sulfate, endrin aldehyde, endrin ketone, gamma-BHC, heptachlor, and heptachlor epoxide had low recoveries.

iv. Precision – All relative percent differences (RPD) reported and less than method or laboratory limits? And project specified DQOs, if applicable. RPD reported from LCS/LCSD, MS/MSD, and or sample/sample duplicate. (AK Petroleum methods 20%; all other analyses see the laboratory QC pages)

Yes  No  NA (Please explain.) Comments:

CR-AG-02-Fillet MS/MSD for alpha-BHC, endrin aldehyde, and Aroclor 1260 had RPDs outside of QC criteria.

v. If %R or RPD is outside of acceptable limits, what samples are affected?

Comments:

CR-AG-02-Fillet

vi. Do the affected sample(s) have data flags? If so, are the data flags clearly defined?

Yes  No  NA (Please explain.) Comments:

All MS/MSD recoveries had dilution factors greater than or equal to 5.

vii. Data quality or usability affected? (Use comment box to explain.)

Comments:

Data quality and usability are minimally affected. The large dilution factors and RPDs outside of criteria suggest potential matrix interference.

c. Surrogates – Organics Only

i. Are surrogate recoveries reported for organic analyses – field, QC and laboratory samples?

Yes  No  NA (Please explain.) Comments:

ii. Accuracy – All percent recoveries (%R) reported and within method or laboratory limits? And project specified DQOs, if applicable. (AK Petroleum methods 50-150 %R; all other analyses see the laboratory report pages)

Yes  No  NA (Please explain.) Comments:

Surrogates tetrachlorometaxylene for SW8081 and decachlorobiphenyl for SW8082 had low recoveries in one or more samples. The surrogate decachlorobiphenyl for SW8082 had high recoveries in one or more samples.

iii. Do the sample results with failed surrogate recoveries have data flags? If so, are the data flags clearly defined?

Yes  No  NA (Please explain.) Comments:

All affected samples had dilution factors greater than or equal to 5; therefore, flags were not assigned.

iv. Data quality or usability affected? (Use the comment box to explain.)

Comments:

Data quality and usability are not affected.

d. Trip blank – Volatile analyses only (GRO, BTEX, Volatile Chlorinated Solvents, etc.):  
Water and Soil

i. One trip blank reported per matrix, analysis and for each cooler containing volatile samples? (If not, enter explanation below.)

Yes  No  NA (Please explain.) Comments:

Trip blanks are not associated with SW7471, SW8081, and SW8082 analyses.

ii. Is the cooler used to transport the trip blank and VOA samples clearly indicated on the COC? (If not, a comment explaining why must be entered below)

Yes  No  NA (Please explain.) Comments:

N/A

iii. All results less than PQL?

Yes  No  NA (Please explain.)

Comments:

N/A

iv. If above PQL, what samples are affected?

Comments:

N/A

v. Data quality or usability affected? (Please explain.)

Comments:

N/A

e. Field Duplicate

i. One field duplicate submitted per matrix, analysis and 10 project samples?

Yes  No  NA (Please explain.)

Comments:

Field duplicates were not collected due to the nature of the samples (fish).

ii. Submitted blind to lab?

Yes  No  NA (Please explain.)

Comments:

Field duplicates were not collected.

iii. Precision – All relative percent differences (RPD) less than specified DQOs?  
(Recommended: 30% water, 50% soil)

$$\text{RPD (\%)} = \text{Absolute value of: } \frac{(R_1 - R_2)}{((R_1 + R_2) / 2)} \times 100$$

Where  $R_1$  = Sample Concentration  
 $R_2$  = Field Duplicate Concentration

Yes  No  NA (Please explain.)

Comments:

Field duplicates were not collected.

iv. Data quality or usability affected? (Use the comment box to explain why or why not.)

Comments:

Data quality and usability are not affected.

f. Decontamination or Equipment Blank (If not used explain why).

Yes  No  NA (Please explain.)

Comments:

The fish were sent to the lab whole. Due to this nature of the samples, decontamination/equipment blanks were not collected.

i. All results less than PQL?

Yes  No  NA (Please explain.)

Comments:

N/A

ii. If above PQL, what samples are affected?

Comments:

N/A

iii. Data quality or usability affected? (Please explain.)

Comments:

N/A

**7. Other Data Flags/Qualifiers (ACOE, AFCEE, Lab-Specific, etc.)**

a. Defined and appropriate?

Yes  No  NA (Please explain.)

Comments:

**Laboratory Data Review Checklist**

**Completed by:**

**Title:**  **Date:**

**CS Report Name:**  **Report Date:**

**Consultant Firm:**

**Laboratory Name:**  **Laboratory Report Number:**

**ADEC File Number:**  **ADEC RecKey Number:**

**1. Laboratory**

a. Did an ADEC CS-approved laboratory receive and perform all of the submitted sample analyses?

Yes  No  NA (Please explain.) Comments:

b. If the samples were transferred to another “network” laboratory or sub-contracted to an alternate laboratory, was the laboratory performing the analyses ADEC CS approved?

Yes  No  NA (Please explain.) Comments:

**2. Chain of Custody (CoC)**

a. CoC information completed, signed, and dated (including released/received by)?

Yes  No  NA (Please explain.) Comments:

b. Correct Analyses requested?

Yes  No  NA (Please explain.) Comments:



**3. Laboratory Sample Receipt Documentation**

a. Sample/cooler temperature documented and within range at receipt ( $4^{\circ} \pm 2^{\circ} \text{C}$ )?

Yes  No  NA (Please explain.)

Comments:

Four coolers were received at temperatures of 2.8°C, 2.6°C, 2.6°C, and 1.9°C. The coolers contained all the fish tissue samples, including those in 3 other SDGs. Temperatures are indistinguishable between coolers and their associated samples, and therefore all four cooler temperatures have been listed.

b. Sample preservation acceptable – acidified waters, Methanol preserved VOC soil (GRO, BTEX, Volatile Chlorinated Solvents, etc.)?

Yes  No  NA (Please explain.)

Comments:

c. Sample condition documented – broken, leaking (Methanol), zero headspace (VOC vials)?

Yes  No  NA (Please explain.)

Comments:

d. If there were any discrepancies, were they documented? For example, incorrect sample containers/preservation, sample temperature outside of acceptable range, insufficient or missing samples, etc.?

Yes  No  NA (Please explain.)

Comments:

The shipping temperature was noted. Samples were received in good condition.

e. Data quality or usability affected? (Please explain.)

Comments:

The fish samples are unaffected by the shipping temperature. Upon arrival at the lab, the samples were stored in the freezer.

**4. Case Narrative**

a. Present and understandable?

Yes  No  NA (Please explain.)

Comments:

b. Discrepancies, errors or QC failures identified by the lab?

Yes  No  NA (Please explain.)

Comments:

c. Were all corrective actions documented?

Yes  No  NA (Please explain.)

Comments:

d. What is the effect on data quality/usability according to the case narrative?

Comments:

See respective sections of this checklist for data quality and usability issues.

**5. Samples Results**

a. Correct analyses performed/reported as requested on COC?

Yes  No  NA (Please explain.)

Comments:

b. All applicable holding times met?

Yes  No  NA (Please explain.)

Comments:

Samples were analyzed outside of holding time for each analysis. Samples were shipped in August 2012 by EA Engineering placed on hold at the lab at that time. Samples were released for analysis by Jacobs (after acquiring the contract) in January 2013, well over 100 days past holding time. Samples were placed in frozen storage between those months. The analyses are clearly out of hold, but sample data was not flagged because the holding time issue would apply to all sample results. It is uncertain whether being frozen would extend the applicable hold time. It is also unlikely these analytes (such as mercury and PCBs) would degrade or be affected leading to a potential low bias. The data was considered usable for the purposes of this project.

c. All soils reported on a dry weight basis?

Yes  No  NA (Please explain.)

Comments:

Only fish tissue was analyzed in this SDG.

d. Are the reported PQLs less than the Cleanup Level or the minimum required detection level for the project?

Yes  No  NA (Please explain.)

Comments:

e. Data quality or usability affected?

Comments:

Data quality and usability are not affected.

**6. QC Samples**

a. Method Blank

i. One method blank reported per matrix, analysis and 20 samples?

Yes  No  NA (Please explain.)

Comments:

ii. All method blank results less than PQL?

Yes  No  NA (Please explain.)

Comments:

Methoxychlor was detected in method blank 180-61868/1-B.

iii. If above PQL, what samples are affected?

Yes  No  NA (Please explain.)

Comments:

All samples in batch 180-61868 are affected.

iv. Do the affected sample(s) have data flags and if so, are the data flags clearly defined?

Yes  No  NA (Please explain.)

Comments:

The sample results for methoxychlor in samples CK-AG-01-Fillet, CK-AG-05-Whole, CK-AG-06-Whole, CK-AG-07-Fillet, CK-AG-07-Whole, CK-AG-08-Fillet, CK-AG-08-Whole, CK-AG-09-Fillet, and CK-AG-09-Whole were flagged B due to having detected results less than 5 times the method blank concentration.

v. Data quality or usability affected? (please explain)

Comments:

Sample results flagged B are biased high but are less than PAL. Data usability is not affected.

b. Laboratory Control Sample/Duplicate (LCS/LCSD)

i. Organics – One LCS/LCSD reported per matrix, analysis and 20 samples? (LCS/LCSD required per AK methods, LCS required per SW846)

Yes  No  NA (Please explain.)

Comments:

All batches contain either an LCS/LCSD pair or an LCS and MS/MSD pair.

ii. Metals/Inorganics – one LCS and one sample duplicate reported per matrix, analysis and 20 samples?

Yes  No  NA (Please explain.)

Comments:

iii. Accuracy – All percent recoveries (%R) reported and within method or laboratory limits? And project specified DQOs, if applicable. (AK Petroleum methods: AK101 60%-120%, AK102 75%-125%, AK103 60%-120%; all other analyses see the laboratory QC pages)

Yes  No  NA (Please explain.)

Comments:

CK-AG-04-Fillet MS/MSD recoveries for alpha-BHC, delta-BHC, endosulfan sulfate, endrin aldehyde, and gamma-BHC were outside of QC criteria (biased low).

CK-AG-04-Whole MS/MSD recoveries for 4,4'-DDD, 4,4'-DDE, aldrin, alpha-BHC, beta-BHC, endrin aldehyde, gamma-BHC, heptachlor, and heptachlor epoxide were outside of QC criteria (biased low).

iv. Precision – All relative percent differences (RPD) reported and less than method or laboratory limits? And project specified DQOs, if applicable. RPD reported from LCS/LCSD, MS/MSD, and or sample/sample duplicate. (AK Petroleum methods 20%; all other analyses see the laboratory QC pages)

Yes  No  NA (Please explain.)

Comments:

CK-AG-04-Whole MS/MSD RPD for endrin aldehyde was outside of QC criteria.

CK-AG-04-Fillet MS/MSD RPD for Aroclor 1260 was outside of QC criteria.

v. If %R or RPD is outside of acceptable limits, what samples are affected?

Comments:

CK-AG-04-Fillet, CK-AG-04-Whole

vi. Do the affected sample(s) have data flags? If so, are the data flags clearly defined?

Yes  No  NA (Please explain.) Comments:

All MS/MSD recoveries had dilution factors greater than or equal to 5.

vii. Data quality or usability affected? (Use comment box to explain.)

Comments:

Data quality and usability are minimally affected. The large dilution factors and RPDs outside of criteria suggest potential matrix interference.

c. Surrogates – Organics Only

i. Are surrogate recoveries reported for organic analyses – field, QC and laboratory samples?

Yes  No  NA (Please explain.) Comments:

ii. Accuracy – All percent recoveries (%R) reported and within method or laboratory limits? And project specified DQOs, if applicable. (AK Petroleum methods 50-150 %R; all other analyses see the laboratory report pages)

Yes  No  NA (Please explain.) Comments:

Surrogate tetrachlorometaxylene (SW8081) had low recoveries in multiple samples. The surrogate decachlorobiphenyl (SW8082) had high recoveries in samples CK-AG-12-Whole and CK-AG-20-Whole.

iii. Do the sample results with failed surrogate recoveries have data flags? If so, are the data flags clearly defined?

Yes  No  NA (Please explain.) Comments:

All affected samples had dilution factors greater than or equal to 5; therefore, flags were not assigned.

iv. Data quality or usability affected? (Use the comment box to explain.)

Comments:

Data quality and usability are not affected.

d. Trip blank – Volatile analyses only (GRO, BTEX, Volatile Chlorinated Solvents, etc.):  
Water and Soil

i. One trip blank reported per matrix, analysis and for each cooler containing volatile samples? (If not, enter explanation below.)

Yes  No  NA (Please explain.) Comments:

Trip blanks are not associated with SW7471, SW8081, and SW8082 analyses.

ii. Is the cooler used to transport the trip blank and VOA samples clearly indicated on the COC? (If not, a comment explaining why must be entered below)

Yes  No  NA (Please explain.) Comments:

N/A

iii. All results less than PQL?

Yes  No  NA (Please explain.)

Comments:

N/A

iv. If above PQL, what samples are affected?

Comments:

N/A

v. Data quality or usability affected? (Please explain.)

Comments:

N/A

e. Field Duplicate

i. One field duplicate submitted per matrix, analysis and 10 project samples?

Yes  No  NA (Please explain.)

Comments:

Field duplicates were not collected due to the nature of the samples (fish).

ii. Submitted blind to lab?

Yes  No  NA (Please explain.)

Comments:

Field duplicates were not collected.

iii. Precision – All relative percent differences (RPD) less than specified DQOs?  
(Recommended: 30% water, 50% soil)

$$\text{RPD (\%)} = \text{Absolute value of: } \frac{(R_1 - R_2)}{((R_1 + R_2)/2)} \times 100$$

Where  $R_1$  = Sample Concentration  
 $R_2$  = Field Duplicate Concentration

Yes  No  NA (Please explain.)

Comments:

Field duplicates were not collected.

iv. Data quality or usability affected? (Use the comment box to explain why or why not.)

Comments:

Data quality and usability are not affected.

f. Decontamination or Equipment Blank (If not used explain why).

Yes  No  NA (Please explain.)

Comments:

The fish were sent to the lab whole. Due to the nature of sampling, decontamination/equipment blanks were not collected.

i. All results less than PQL?

Yes  No  NA (Please explain.)

Comments:

N/A

ii. If above PQL, what samples are affected?

Comments:

N/A

iii. Data quality or usability affected? (Please explain.)

Comments:

N/A

**7. Other Data Flags/Qualifiers (ACOE, AFCEE, Lab-Specific, etc.)**

a. Defined and appropriate?

Yes  No  NA (Please explain.)

Comments:

**Laboratory Data Review Checklist**

**Completed by:**

**Title:**  **Date:**

**CS Report Name:**  **Report Date:**

**Consultant Firm:**

**Laboratory Name:**  **Laboratory Report Number:**

**ADEC File Number:**  **ADEC RecKey Number:**

**1. Laboratory**

a. Did an ADEC CS-approved laboratory receive and perform all of the submitted sample analyses?

Yes  No  NA (Please explain.) Comments:

b. If the samples were transferred to another “network” laboratory or sub-contracted to an alternate laboratory, was the laboratory performing the analyses ADEC CS approved?

Yes  No  NA (Please explain.) Comments:

**2. Chain of Custody (CoC)**

a. CoC information completed, signed, and dated (including released/received by)?

Yes  No  NA (Please explain.) Comments:

b. Correct Analyses requested?

Yes  No  NA (Please explain.) Comments:

**3. Laboratory Sample Receipt Documentation**

a. Sample/cooler temperature documented and within range at receipt ( $4^{\circ} \pm 2^{\circ} \text{C}$ )?

Yes  No  NA (Please explain.)

Comments:

Four coolers were received at temperatures of 2.8°C, 2.6°C, 2.6°C, and 1.9°C. The coolers contained all the fish tissue samples, including those in 3 other SDGs. Temperatures are indistinguishable between coolers and their associated samples, and therefore all four cooler temperatures have been listed.

b. Sample preservation acceptable – acidified waters, Methanol preserved VOC soil (GRO, BTEX, Volatile Chlorinated Solvents, etc.)?

Yes  No  NA (Please explain.)

Comments:

c. Sample condition documented – broken, leaking (Methanol), zero headspace (VOC vials)?

Yes  No  NA (Please explain.)

Comments:

d. If there were any discrepancies, were they documented? For example, incorrect sample containers/preservation, sample temperature outside of acceptable range, insufficient or missing samples, etc.?

Yes  No  NA (Please explain.)

Comments:

The shipping temperature was noted. Samples were received in good condition.

e. Data quality or usability affected? (Please explain.)

Comments:

The fish samples are unaffected by the shipping temperature. Upon arrival at the lab, the samples were stored in the freezer.

**4. Case Narrative**

a. Present and understandable?

Yes  No  NA (Please explain.)

Comments:

b. Discrepancies, errors or QC failures identified by the lab?

Yes  No  NA (Please explain.)

Comments:

c. Were all corrective actions documented?

Yes  No  NA (Please explain.)

Comments:

d. What is the effect on data quality/usability according to the case narrative?

Comments:

See respective sections of this checklist for data quality and usability issues.



**5. Samples Results**

a. Correct analyses performed/reported as requested on COC?

Yes  No  NA (Please explain.)

Comments:

b. All applicable holding times met?

Yes  No  NA (Please explain.)

Comments:

Samples were analyzed outside of holding time for each analysis. Samples were shipped in August 2012 by EA Engineering placed on hold at the lab at that time. Samples were released for analysis by Jacobs (after acquiring the contract) in January 2013, well over 100 days past holding time. Samples were placed in frozen storage between those months. The analyses are clearly out of hold, but sample data was not flagged because the holding time issue would apply to all sample results. It is uncertain whether being frozen would extend the applicable hold time. It is also unlikely these analytes (such as mercury and PCBs) would degrade or be affected leading to a potential low bias. The data was considered usable for the purposes of this project.

c. All soils reported on a dry weight basis?

Yes  No  NA (Please explain.)

Comments:

Only fish tissue was analyzed in this SDG.

d. Are the reported PQLs less than the Cleanup Level or the minimum required detection level for the project?

Yes  No  NA (Please explain.)

Comments:

Aldrin, alpha-BHC, dieldrin, heptachlor, and toxaphene had nondetect results and LODs greater than the project action limit (PAL) for one or more samples. These sample results were flagged E.

e. Data quality or usability affected?

Comments:

Data is considered usable for the purposes of this project.

**6. QC Samples**

a. Method Blank

i. One method blank reported per matrix, analysis and 20 samples?

Yes  No  NA (Please explain.)

Comments:

ii. All method blank results less than PQL?

Yes  No  NA (Please explain.)

Comments:

Methoxychlor was detected in method blank 180-62279/1-B.

iii. If above PQL, what samples are affected?

Yes  No  NA (Please explain.)

Comments:

All samples in batch 180-62279 are affected.

iv. Do the affected sample(s) have data flags and if so, are the data flags clearly defined?

Yes  No  NA (Please explain.)

Comments:

Sample results for methoxychlor in samples PS-AG-01-Fillet, PS-AG-04-Whole, PS-AG-05-Whole, PS-AG-06-Fillet, PS-AG-06-Whole, PS-AG-07-Fillet, PS-AG-07-Whole, PS-AG-09-Fillet were flagged B due to detected results less than 5 times the method blank concentration.

v. Data quality or usability affected? (please explain)

Comments:

Sample results flagged B potentially are biased high but are less than PAL. Data usability is not affected.

b. Laboratory Control Sample/Duplicate (LCS/LCSD)

i. Organics – One LCS/LCSD reported per matrix, analysis and 20 samples? (LCS/LCSD required per AK methods, LCS required per SW846)

Yes  No  NA (Please explain.)

Comments:

All batches had either an LCS/LCSD pair or an LCS and MS/MSD pair.

ii. Metals/Inorganics – one LCS and one sample duplicate reported per matrix, analysis and 20 samples?

Yes  No  NA (Please explain.)

Comments:

iii. Accuracy – All percent recoveries (%R) reported and within method or laboratory limits? And project specified DQOs, if applicable. (AK Petroleum methods: AK101 60%-120%, AK102 75%-125%, AK103 60%-120%; all other analyses see the laboratory QC pages)

Yes  No  NA (Please explain.)

Comments:

PS-AG-09-Fillet MS/MSD recoveries for endosulfan sulfate and endrin aldehyde were outside of QC criteria (biased low).

PS-AG-09-Whole MS/MSD recoveries for 4,4'-DDD, 4,4'-DDE, 4,4'-DDT, aldrin, alpha-BHC, beta-BHC, dieldrin, endosulfan I, endosulfan II, endosulfan sulfate, endrin aldehyde, endrin ketone, gamma-BHC, heptachlor, heptachlor epoxide, and methoxychlor were outside of QC criteria (biased low).

iv. Precision – All relative percent differences (RPD) reported and less than method or laboratory limits? And project specified DQOs, if applicable. RPD reported from LCS/LCSD, MS/MSD, and or sample/sample duplicate. (AK Petroleum methods 20%; all other analyses see the laboratory QC pages)

Yes  No  NA (Please explain.)

Comments:

PS-AG-09-Fillet MS/MSD RPD for delta-BHC, endrin aldehyde, endosulfan sulfate, and gamma-BHC were outside of QC criteria.

PS-AG-09-Whole MS/MSD RPD for 4,4'-DDD, 4,4'-DDE, aldrin, alpha-BHC, alpha-Chlordane, beta-BHC, delta-BHC, dieldrin, endrin, endosulfan I, endosulfan II, gamma-BHC, gamma-Chlordane, heptachlor epoxide, heptachlor, and methoxychlor were outside of QC criteria.

v. If %R or RPD is outside of acceptable limits, what samples are affected?

Comments:

PS-AG-09-Fillet and PS-AG-09-Whole

vi. Do the affected sample(s) have data flags? If so, are the data flags clearly defined?

Yes  No  NA (Please explain.)

Comments:

All MS/MSD recoveries had dilution factors greater than or equal to 5.

vii. Data quality or usability affected? (Use comment box to explain.)

Comments:

Data quality and usability are minimally affected. The large dilution factors and RPDs outside of criteria suggest potential matrix interference.

c. Surrogates – Organics Only

i. Are surrogate recoveries reported for organic analyses – field, QC and laboratory samples?

Yes  No  NA (Please explain.)

Comments:

ii. Accuracy – All percent recoveries (%R) reported and within method or laboratory limits? And project specified DQOs, if applicable. (AK Petroleum methods 50-150 %R; all other analyses see the laboratory report pages)

Yes  No  NA (Please explain.)

Comments:

Surrogates decachlorobiphenyl and tetrachlorometaxylene (SW8081) had low recoveries in one or more samples. Surrogates decachlorobiphenyl and tetrachlorometaxylene (SW8082) had high recoveries in one or more samples.

iii. Do the sample results with failed surrogate recoveries have data flags? If so, are the data flags clearly defined?

Yes  No  NA (Please explain.)

Comments:

All affected samples had dilution factors greater than or equal to 5 therefore flags were not assigned.

iv. Data quality or usability affected? (Use the comment box to explain.)

Comments:

Data quality and usability are not affected.

d. Trip blank – Volatile analyses only (GRO, BTEX, Volatile Chlorinated Solvents, etc.):

Water and Soil

i. One trip blank reported per matrix, analysis and for each cooler containing volatile samples? (If not, enter explanation below.)

Yes  No  NA (Please explain.)

Comments:

Trip blanks are not associated with SW7471, SW8081, and SW8082 analyses.

ii. Is the cooler used to transport the trip blank and VOA samples clearly indicated on the COC?  
(If not, a comment explaining why must be entered below)

Yes  No  NA (Please explain.) Comments:

N/A

iii. All results less than PQL?

Yes  No  NA (Please explain.) Comments:

N/A

iv. If above PQL, what samples are affected?

Comments:

N/A

v. Data quality or usability affected? (Please explain.)

Comments:

N/A

e. Field Duplicate

i. One field duplicate submitted per matrix, analysis and 10 project samples?

Yes  No  NA (Please explain.) Comments:

Field duplicates were not collected due to the nature of the samples (fish).

ii. Submitted blind to lab?

Yes  No  NA (Please explain.) Comments:

Field duplicates were not collected.

iii. Precision – All relative percent differences (RPD) less than specified DQOs?  
(Recommended: 30% water, 50% soil)

$$\text{RPD (\%)} = \text{Absolute value of: } \frac{(R_1 - R_2)}{((R_1 + R_2)/2)} \times 100$$

Where  $R_1$  = Sample Concentration  
 $R_2$  = Field Duplicate Concentration

Yes  No  NA (Please explain.) Comments:

Field duplicates were not collected.

iv. Data quality or usability affected? (Use the comment box to explain why or why not.)

Comments:

Data quality and usability are not affected.

f. Decontamination or Equipment Blank (If not used explain why).

Yes  No  NA (Please explain.)

Comments:

The fish were sent to the lab whole. Due to this nature of the samples, decontamination/equipment blanks were not collected.

i. All results less than PQL?

Yes  No  NA (Please explain.)

Comments:

N/A

ii. If above PQL, what samples are affected?

Comments:

N/A

iii. Data quality or usability affected? (Please explain.)

Comments:

N/A

**7. Other Data Flags/Qualifiers (ACOE, AFCEE, Lab-Specific, etc.)**

a. Defined and appropriate?

Yes  No  NA (Please explain.)

Comments:

**Laboratory Data Review Checklist**

**Completed by:**

**Title:**  **Date:**

**CS Report Name:**  **Report Date:**

**Consultant Firm:**

**Laboratory Name:**  **Laboratory Report Number:**

**ADEC File Number:**  **ADEC RecKey Number:**

**1. Laboratory**

a. Did an ADEC CS-approved laboratory receive and perform all of the submitted sample analyses?

Yes    No    NA (Please explain.)                      Comments:

b. If the samples were transferred to another “network” laboratory or sub-contracted to an alternate laboratory, was the laboratory performing the analyses ADEC CS approved?

Yes    No    NA (Please explain.)                      Comments:

**2. Chain of Custody (CoC)**

a. CoC information completed, signed, and dated (including released/received by)?

Yes    No    NA (Please explain.)                      Comments:

b. Correct Analyses requested?

Yes    No    NA (Please explain.)                      Comments:

**3. Laboratory Sample Receipt Documentation**

a. Sample/cooler temperature documented and within range at receipt ( $4^{\circ} \pm 2^{\circ} \text{C}$ )?

Yes    No    NA (Please explain.)                      Comments:

b. Sample preservation acceptable – acidified waters, Methanol preserved VOC soil (GRO, BTEX, Volatile Chlorinated Solvents, etc.)?

Yes  No  NA (Please explain.)

Comments:

c. Sample condition documented – broken, leaking (Methanol), zero headspace (VOC vials)?

Yes  No  NA (Please explain.)

Comments:

d. If there were any discrepancies, were they documented? For example, incorrect sample containers/preservation, sample temperature outside of acceptable range, insufficient or missing samples, etc.?

Yes  No  NA (Please explain.)

Comments:

The low shipping temperature was noted.

Sample date and time matched on both the COC and the label, but as per client request, the date/time of the sample was changed.

e. Data quality or usability affected? (Please explain.)

Comments:

The samples is unaffected by the low shipping temperature. Samples were received in good condition and not frozen.

#### 4. Case Narrative

a. Present and understandable?

Yes  No  NA (Please explain.)

Comments:

b. Discrepancies, errors or QC failures identified by the lab?

Yes  No  NA (Please explain.)

Comments:

c. Were all corrective actions documented?

Yes  No  NA (Please explain.)

Comments:

d. What is the effect on data quality/usability according to the case narrative?

Comments:

See respective sections of this checklist for data quality and usability issues.

#### 5. Samples Results

a. Correct analyses performed/reported as requested on COC?

Yes  No  NA (Please explain.)

Comments:

b. All applicable holding times met?

Yes  No  NA (Please explain.)

Comments:

c. All soils reported on a dry weight basis?

Yes  No  NA (Please explain.)

Comments:

d. Are the reported PQLs less than the Cleanup Level or the minimum required detection level for the project?

Yes  No  NA (Please explain.)

Comments:

e. Data quality or usability affected?

Comments:

## 6. QC Samples

a. Method Blank

i. One method blank reported per matrix, analysis and 20 samples?

Yes  No  NA (Please explain.)

Comments:

ii. All method blank results less than PQL?

Yes  No  NA (Please explain.)

Comments:

iii. If above PQL, what samples are affected?

Yes  No  NA (Please explain.)

Comments:

iv. Do the affected sample(s) have data flags and if so, are the data flags clearly defined?

Yes  No  NA (Please explain.)

Comments:

v. Data quality or usability affected? (please explain)

Comments:

b. Laboratory Control Sample/Duplicate (LCS/LCSD)

i. Organics – One LCS/LCSD reported per matrix, analysis and 20 samples? (LCS/LCSD required per AK methods, LCS required per SW846)

Yes  No  NA (Please explain.)

Comments:



ii. Metals/Inorganics – one LCS and one sample duplicate reported per matrix, analysis and 20 samples?

Yes  No  NA (Please explain.) Comments:

An LCS/LCSD pair was analyzed for metals.

iii. Accuracy – All percent recoveries (%R) reported and within method or laboratory limits? And project specified DQOs, if applicable. (AK Petroleum methods: AK101 60%-120%, AK102 75%-125%, AK103 60%-120%; all other analyses see the laboratory QC pages)

Yes  No  NA (Please explain.) Comments:

iv. Precision – All relative percent differences (RPD) reported and less than method or laboratory limits? And project specified DQOs, if applicable. RPD reported from LCS/LCSD, MS/MSD, and or sample/sample duplicate. (AK Petroleum methods 20%; all other analyses see the laboratory QC pages)

Yes  No  NA (Please explain.) Comments:

v. If %R or RPD is outside of acceptable limits, what samples are affected?

Comments:

Accuracy and precision data was met.

vi. Do the affected sample(s) have data flags? If so, are the data flags clearly defined?

Yes  No  NA (Please explain.) Comments:

Accuracy and precision data was met.

vii. Data quality or usability affected? (Use comment box to explain.)

Comments:

Data quality and usability are not affected.

c. Surrogates – Organics Only

i. Are surrogate recoveries reported for organic analyses – field, QC and laboratory samples?

Yes  No  NA (Please explain.) Comments:

ii. Accuracy – All percent recoveries (%R) reported and within method or laboratory limits? And project specified DQOs, if applicable. (AK Petroleum methods 50-150 %R; all other analyses see the laboratory report pages)

Yes  No  NA (Please explain.) Comments:

The surrogate decachlorobiphenyl for SW8082 PCB analysis was outside of QC criteria (biased low).

iii. Do the sample results with failed surrogate recoveries have data flags? If so, are the data flags clearly defined?

Yes  No  NA (Please explain.) Comments:

All Aroclors in the sample 12EAFB-SS67-SO-W01 were flagged JS-.

iv. Data quality or usability affected? (Use the comment box to explain.)

Comments:

Sample results flagged JS- are considered estimated and biased low. Data quality is minimally affected because Aroclor 1260 had a result of 19 mg/kg, which is significantly greater than the ADEC cleanup criteria of 1 mg/kg and significantly less than the TSCA waste level of 50 mg/kg.

d. Trip blank – Volatile analyses only (GRO, BTEX, Volatile Chlorinated Solvents, etc.):

Water and Soil

i. One trip blank reported per matrix, analysis and for each cooler containing volatile samples? (If not, enter explanation below.)

Yes  No  NA (Please explain.) Comments:

Volatiles were not analyzed.

ii. Is the cooler used to transport the trip blank and VOA samples clearly indicated on the COC? (If not, a comment explaining why must be entered below)

Yes  No  NA (Please explain.) Comments:

N/A

iii. All results less than PQL?

Yes  No  NA (Please explain.) Comments:

N/A

iv. If above PQL, what samples are affected?

Comments:

N/A

v. Data quality or usability affected? (Please explain.)

Comments:

N/A

e. Field Duplicate

i. One field duplicate submitted per matrix, analysis and 10 project samples?

Yes  No  NA (Please explain.) Comments:

A field duplicate was not collected. The sample in this SDG was a waste characterization sample, which does not require a field duplicate.

ii. Submitted blind to lab?

Yes  No  NA (Please explain.)

Comments:

A field duplicate was not collected.

iii. Precision – All relative percent differences (RPD) less than specified DQOs?  
(Recommended: 30% water, 50% soil)

$$\text{RPD (\%)} = \text{Absolute value of: } \frac{(R_1 - R_2)}{((R_1 + R_2) / 2)} \times 100$$

Where  $R_1$  = Sample Concentration  
 $R_2$  = Field Duplicate Concentration

Yes  No  NA (Please explain.)

Comments:

A field duplicate was not collected.

iv. Data quality or usability affected? (Use the comment box to explain why or why not.)

Comments:

Data quality and usability are not affected.

f. Decontamination or Equipment Blank (If not used explain why).

Yes  No  NA (Please explain.)

Comments:

The waste characterization sample in this SDG was the only sample collected.

i. All results less than PQL?

Yes  No  NA (Please explain.)

Comments:

N/A

ii. If above PQL, what samples are affected?

Comments:

N/a

iii. Data quality or usability affected? (Please explain.)

Comments:

N/A

**7. Other Data Flags/Qualifiers (ACOE, AFCEE, Lab-Specific, etc.)**

a. Defined and appropriate?

Yes  No  NA (Please explain.)

Comments:

**EXHIBIT D**

**Laboratory Data Deliverables**

*(available separately on CD)*

**ATTACHMENT 6**  
**Waste Tracking**

**2013 Waste Tracking  
Eielson AFB, Alaska**

Site ID	Generation Date	Initials	Waste Container (ex. Supersack, Drum, etc)	Contents	Volume	Units	Estimated Weight (lbs)	Date Staged	Staging Area	Tracking #	Comments	Date Transferred	Transporter
SS67 Slough	11/12/2012	CJ	Super Sack®	IDW (PCB - nonTSCA)	0.5	CY	75 lbs	1/18/2013	Job Trailer	12EAFB-SS67-SS01	IDW from slough investigation Sample results show non-TSCA	2/19/2013	ELM

**NON-HAZARDOUS WASTE MANIFEST**

1. Generator ID Number: **AK1570028646** | 2. Page 1 of **1** | 3. Emergency Response Phone: **800-424-9300** | 4. Waste Tracking Number: **JEAFB0213-03**

5. Generator's Name and Mailing Address: **USAF EIELSON AFB 354 E CEOI 2910 CENTRAL AVENUE, SUITE 100 EIELSON AFB, AK 9. 02**  
 Generator's Site Address (if different than mailing address): **USAF EIELSON AFB - 354 CES/CEOIC Site. WP45/SS57 2681 to 2723 Flightline Blvd**  
 Generator's Phone: **907-372671**

6. Transporter 1 Company Name: **TOTEM TRAILER EXPRESS** / U.S. EPA ID Number: **WAD070397955**

7. Transporter 2 Company Name: **UNION PACIFIC RAILROAD** / U.S. EPA ID Number: **NED001792910**

8. Designated Facility Name and Site Address: **COLUMBIA RIDGE LANDFILL 18177 CEDAR SPRINGS LANE ARLINGTON, OR 97812** / U.S. EPA ID Number: **ORD987173457**  
 Facility's Phone: **541-454-2030**

9. Waste Shipping Name and Description	10. Containers		11. Total Quantity	12. Unit Wt./Vol.
	No.	Type		
1. <b>MATERIAL NOT REGULATED BY DOT (AMINATE OILS)</b>	<b>01</b>	<b>CM</b>		<b>T</b>
2.				
3.				
4.				

13. Special Handling Instructions and Additional Information  
**1) PROFILE # OR112211, POL / PCB CONTAMINATED SOILS AND DEBRIS**  
**CERTIFICATES OF DISPOSAL REQUIRED**

14. GENERATOR'S/OFFEROR'S CERTIFICATION: I hereby declare that the contents of this consignment are fully and accurately described above by the proper shipping name, and are classified, packaged, marked and labeled/placarded, and are in all respects in proper condition for transport according to applicable international and national governmental regulations.

Generator's/Offoror's Printed/Typed Name: **DAVID E. TIDWELL** / Signature: *[Signature]* / Month: **3** / Day: **14** / Year: **13**

15. International Shipments:  Import to U.S. /  Export from U.S. / Port of entry/exit: \_\_\_\_\_ / Date leaving U.S.: \_\_\_\_\_

16. Transporter Acknowledgment of Receipt of Materials  
 Transporter 1 Printed/Typed Name: **DAVID TIDWELL** / Signature: *[Signature]* / Month: **02** / Day: **20** / Year: **13**  
 Transporter 2 Printed/Typed Name: \_\_\_\_\_ / Signature: \_\_\_\_\_ / Month: \_\_\_\_\_ / Day: \_\_\_\_\_ / Year: \_\_\_\_\_

17. Discrepancy  
 17a. Discrepancy Indication Space:  Quantity /  Type /  Residue /  Partial Rejection /  Full Rejection  
 Manifest Reference Number: \_\_\_\_\_

17b. Alternate Facility (or Generator) / U.S. EPA ID Number: \_\_\_\_\_  
 Facility's Phone: \_\_\_\_\_  
 17c. Signature of Alternate Facility (or Generator) / Month: \_\_\_\_\_ / Day: \_\_\_\_\_ / Year: \_\_\_\_\_

18. Designated Facility Owner or Operator: Certification of receipt of materials covered by the manifest except as noted in Item 17a  
 Printed/Typed Name: \_\_\_\_\_ / Signature: \_\_\_\_\_ / Month: \_\_\_\_\_ / Day: \_\_\_\_\_ / Year: \_\_\_\_\_





DISPATCH 456 5031 RANDY  
TOTE ~ 452 1022

DRIVER # \_\_\_\_\_

# WEAVER BROTHERS, INC.

TRUCK # \_\_\_\_\_

278-4526 • 2230 Spar Ave. • Anchorage, Alaska 99501

## WORK ORDER

FROM FAB  
TO WBE  
BILLING/REFERENCE # \_\_\_\_\_  
TRAILER PREFIX WMLXU TRAILER NUMBER 6231  
DATE 2/19/13 CUSTOMER/NAME \_\_\_\_\_ PHONE # \_\_\_\_\_

BILL TO:  
HLA   
TOTE   
CUSTOMER   
CO-BIZ

ROUND TRIP TIME: START \_\_\_\_\_ AM-PM STOP \_\_\_\_\_ AM-PM \_\_\_\_\_ RESPOT  
LOAD TIME: START \_\_\_\_\_ AM-PM STOP \_\_\_\_\_ AM-PM \_\_\_\_\_ ON SITE RESPOT  
UNLOAD TIME: START \_\_\_\_\_ AM-PM STOP \_\_\_\_\_ AM-PM \_\_\_\_\_ HOUSEHOLD GOODS  
DELAY TIME: START 1425 AM-PM STOP 1530 AM-PM

WORK DESCRIPTION:  LOAD  EMPTY  EMPTY FOR SOUTH BOUND  LINEHAUL SUPPORT

STANDBY, while operator finished loading, VAD

Ida Petersen JACOBS August 2/19/13  
RECEIVED IN GOOD CONDITION EXCEPT AS NOTED COMPANY NAME SIGNATURE OF DRIVER



Requested Facility: Columbia Ridge Landfill
Check if there are multiple generator locations Attach locations
Renewal? Original Profile Number

A. GENERATOR INFORMATION (MATERIAL ORIGIN)
1 Generator Name USAF EIELSON AFB - 354 CES/CEOIC
2 Site Address 2310 CENTRAL AVENUE., SUITE 100
3 County
4 Contact Name Randy Smith
5 Email
6 Phone (907) 377-2574 7 Fax:
8 Generator EPA ID AK1570028646
9 State ID

B. BILLING INFORMATION
1 Billing Name: ELM Solutions Corp.
2 Billing Address: 17701 108th Ave. SE, Ste.427
3 Contact Name: Mike Gortner
4 Email: mikeg@elmsolutionscorp.com
5 Phone: (425) 591-2267 6 Fax: (888) 356-3299
7 WM Hauled?
8 PO Number:

C. MATERIAL INFORMATION
1 Common Name POL/PCB Contaminated Soils and Debris
Describe Process Generating Material. See Attached

Site Investigation

Table with 2 columns: Material Composition and Contaminants, and percentage values (50-100%, 0-20%, 0-20%, 0-5%, >=100%)

3. State Waste Codes:
4 Color Varies
5. Physical State at 70°F: Solid
6. Free Liquid Range Percentage: N/A (Solid)
7. pH: N/A (Solid)
8. Strong Odor: No
9. Flash Point: N/A (Solid)

D. REGULATORY INFORMATION
1. EPA Hazardous Waste?
2. State Hazardous Waste?
3. Excluded waste under 40 CFR 261.4 (a) or (b)?
4. Contains Underlying Hazardous Constituents?
5. Contains benzene and subject to Benzene NESHAP?
6. Facility remediation subject to 40 CFR 63 GGGGG?
7. CERCLA or State-mandated clean-up?
8. NRC or State-regulated radioactive or NORM waste?
9. Contains PCBs?
10. Regulated and/or Untreated Medical/Infectious Waste?
11. Contains Asbestos?

E. ANALYTICAL AND OTHER REPRESENTATIVE INFORMATION
1. Analytical attached
Please identify applicable samples and/or lab reports:

F. SHIPPING AND DOT INFORMATION
1. One-Time Event / Repeat Event/Ongoing Business
2. Estimated Quantity/Unit of Measure: 40
3. Container Type and Size: 20' Intermodal
4. USDOT Proper Shipping Name:

G. GENERATOR CERTIFICATION (PLEASE READ AND CERTIFY BY SIGNATURE)
By signing this EZ Profile™ form, I hereby certify that all information submitted in this and all attached documents contain true and accurate descriptions of this material, and that all relevant information necessary for proper material characterization and to identify known and suspected hazards has been provided.

If I am an agent signing on behalf of the Generator, I have confirmed with the Generator that information contained in this Profile is accurate and complete.
Name (Print) Date:
Title
Company

Certification Signature



# EZ Profile™ Addendum



Only complete this Addendum if prompted by responses on EZ Profile™ (page 1) or to provide additional information. Sections and question numbers correspond to EZ Profile™.

Profile Number 112211OR

### C. MATERIAL INFORMATION

Describe Process Generating Material (Continued from page 1).

If more space is needed, please attach additional pages

Material Composition and Contaminants (Continued from page 1)

If more space is needed, please attach additional pages.

5	PCB Contamination	0-49 ppm
6		
7		
8		
9		
10		
		≥100%

### D. REGULATORY INFORMATION

Only questions with a "Yes" response in Section D on the EZ Profile™ form (page 1) need to be answered here.

1 EPA Hazardous Waste

a Please list all USEPA listed and characteristic waste code numbers

- b Is the material subject to the Alternative Debris standards (40 CFR 268.45)?  Yes  No
- c Is the material subject to the Alternative Soil standards (40 CFR 268.49)? → If Yes, complete question 4.  Yes  No
- d Is the material exempt from Subpart CC Controls (40 CFR 264.1083 and 265.1084)?  Yes  No

→ If Yes, please select one of the following:

- Waste has been determined to be LDR exempt [265.1083(c)(4) and 265.1084(c)(4)] based on the fact that it meets all applicable organic treatment standards (including UHCs for D-coded characteristic wastes) or a Specified Technology has been utilized.
- Waste does not qualify for a LDR exemption, but the average VOC at the point of origination is <500 ppmw and this determination was based on analytical testing (upload copy of analysis) or generator knowledge.

2 State Hazardous Waste → Please list all state waste codes \_\_\_\_\_

3 Excluded Waste → Please select which of the following categories apply to your material.

- Delisted Hazardous Waste  Excluded Waste under 40 CFR 261.4 → Specify Exclusion: \_\_\_\_\_
- Treated Hazardous Waste Debris  Treated Characteristic Hazardous Waste → If checked, complete question 4.

4 Underlying Hazardous Constituents → Please list all Underlying Hazardous Constituents:

5. Benzene NESHAP → Please include benzene concentration and percent water/moisture in chemical composition.

a. Are you a TSDF? → If yes, please complete Benzene NESHAP questionnaire. If not, continue.

b. What is your facility's current total annual benzene quantity in Megagrams?

<1 Mg  1-9.99 Mg  ≥10 Mg

c. Is this waste soil from remediation at a closed facility?  Yes  No

d. Has material been treated to remove 99% of the benzene or to achieve <10 ppmw?  Yes  No

e. Is material exempt from controls in accordance with 40 CFR 61.342?  Yes  No

→ If yes, specify exemption: \_\_\_\_\_

f. Based on your knowledge of your waste and the BWON regulations, do you believe that this waste stream is subject to treatment and control requirements at an off-site TSDF?  Yes  No

6. 40 CFR 63 GGGGG → Does the material contain <500 ppw VOHAPs at the point of determination?  Yes  No

7. CERCLA or State-Mandated clean up → Please submit the Record of Decision or other documentation to assist others in the evaluation for proper disposal.

8. NRC or state regulated radioactive or NORM Waste → Please identify Isotopes and pCi/g \_\_\_\_\_

THINK GREEN.®

QUESTIONS? CALL 800 963 4776 FOR ASSISTANCE

Last Revised June 6, 2012  
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Thank you for choosing Carlile. For shipment information visit [www.carlile.biz](http://www.carlile.biz) or contact our Customer Service Department Toll Free (888) 399-3290 (907) 343-3290 Fax: (907) 278-0971 • Email: [customerservice@carlile.biz](mailto:customerservice@carlile.biz)

Subject to standard terms & conditions at [www.carlile.biz](http://www.carlile.biz)  
**Carlile 80692765** RE  
  
 Shippers # / P.O. #

**TRANSPORTATION SYSTEMS, INC.**

Corporate Office 1800 E. First Avenue • Anchorage, AK 99501

UNIFORM STRAIGHT BILL OF LADING ORIGINAL — NOT NEGOTIABLE

1 FROM	Company Name	Jacobs Engineering	TRAILER NUMBER	293357	B/L DATE	2/18/13
	Shippers Name		<b>HAZARDOUS MATERIALS EMERGENCY CONTACT NUMBER</b>			
	Phone Number	907 301 0342 (Toll) (907)				
	Address	2953 Flight Line Ave				
City	ELIOTON AFB	State	AK	99702	ERG#	
2 TO	Company Name	waste management	3	<input checked="" type="checkbox"/> COLLECT	<input type="checkbox"/> PREPAID	<input checked="" type="checkbox"/> THIRD PARTY
	Consignee Name		<input checked="" type="checkbox"/> EIM SOLUTIONS			
	Phone Number	(206) - 763 - 6641	Name	17701 108th Avenue		
	Address	700 ALASKA ST	Address	SEATTLE WA 98134		
City	SEATTLE	State	WA	98134		

Collect on Delivery \$ \_\_\_\_\_ Remit to: \_\_\_\_\_  
 Street \_\_\_\_\_ City \_\_\_\_\_ State \_\_\_\_\_ Signed \_\_\_\_\_  
 Carrier must collect cash or a certified check unless shipper signs here to accept company check

The property described below, in apparent good order, except as noted (contents and condition of contents of packages unknown), marked, consigned and destined as shown below, which said carrier agrees to carry to destination, if on its routes, otherwise to deliver to another carrier on the route to destination. Every service to be performed hereunder shall be subject to all the conditions not prohibited by law, whether printed or written, herein contained, including the conditions on back hereof, which are hereby agreed to by the shipper and accepted for himself and his assigns.

**DESCRIPTION OF ARTICLES, SPECIAL MARKS, AND EXCEPTIONS**

4	Number of Packages	Type of package (bx, cyl, crate, tube, drum, trailer)	HM X or RQ	Commodity Description Proper Shipping Name, Hazard Class, ID Number, Packing Group	Weight (lbs) (subject to correction)	Dimensions Length x Width x Height LWH	Freight Charges
	31	drums		waste water 10 Pallets	15,000		

**5 SPECIAL SERVICES (ADDITIONAL FEES MAY APPLY)**

<input type="checkbox"/> KFF	<input type="checkbox"/> Requested Delivery Date _____	<input type="checkbox"/> Delivered to Dock	<input type="checkbox"/> Special Labor Time
<input type="checkbox"/> Chill/Temperature	<input type="checkbox"/> Requested Delivery Time _____ (AM) (PM)	<input type="checkbox"/> Picked up at Dock	Start _____ End _____
<input type="checkbox"/> Freeze/Temperature	<input type="checkbox"/> Standby Start _____	<input type="checkbox"/> Saturday Pickup / Delivery	<input type="checkbox"/> Hazardous Waste
<input type="checkbox"/> Tarped	End _____	<input type="checkbox"/> After Hours Pickup / Delivery	<input type="checkbox"/> Non Regulated Waste
<input type="checkbox"/> Lift Gate	<input type="checkbox"/> Hold At Dock	<input type="checkbox"/> Pilot Car	<input type="checkbox"/> High HazMat class 1.1, 1.2, 1.3, 2.3, 4.3, 5.2, 6.1 PIH PGI, 7
<input type="checkbox"/> Oversize	<input type="checkbox"/> Storage/Dates _____ to _____	<input type="checkbox"/> Documentation Fee(s)	
<input type="checkbox"/> Residential Delivery		<input type="checkbox"/> Unstackable: Cubic Size _____	

**6 LIMITS OF LIABILITY FOR CARGO LOSS OR DAMAGE**

Where no value is declared below, the maximum liability on the shipment will be \$20.00 per pound up to a maximum of \$200,000 per shipment for new goods. Used goods will have a maximum liability of \$0.10 per pound up to a maximum of \$5,000 per shipment.  
 Declared Value of Shipment \$ \_\_\_\_\_ (Ref section 3 on reverse side)  
 NOTE: For information on Heavy Haul shipments, Declared Value shipments, limits of liability and claims see the back of this Bill of Lading.

**7 FOR FREIGHT COLLECT SHIPMENTS**

If this shipment is to be delivered to the consignee without recourse on the consignor, the consignor shall sign the following statement: The carrier shall not make delivery of this shipment without payment of freight and all other lawful charges.  
 X \_\_\_\_\_  
 (Signature of Consignor)

**8 SHIPPERS CERTIFICATION**

Shippers Signature \_\_\_\_\_ Total Pcs. Shipped \_\_\_\_\_ Date \_\_\_\_\_  
 This is to certify that the above-named materials are properly classified, described, packaged, marked and labeled and are in proper condition for transportation, according to the applicable regulations of the Department of Transportation.

**FOR CARLILE USE ONLY**

Carlile P/U By	Date	Time	AM	Total Pcs.	Carlile Del By:	Date	Time	AM	Total Pcs.
Pete 5155	2/18/13	3:20	(AM)	31				(PM)	

Consignee Signature \_\_\_\_\_ Received the above specified property in apparent good order  
 Total Pcs. Rec'd \_\_\_\_\_ Print Last Name \_\_\_\_\_ Date \_\_\_\_\_ Time \_\_\_\_\_  
**SHIPPER**

CCN# 670873

NON-HAZARDOUS WASTE MANIFEST

1. Generator ID Number

AK1570029646

2. Page 1 of 1

3. Emergency Response Phone

800-424-9300

4. Waste Tracking Number

LEAFB0213-02

5. Generator's Name and Mailing Address

USAF EIELSON AFB - 354 CES/CEIC  
2310 CENTRAL AVENUE, SUITE 100  
EIELSON AFB, AK 99702

Generator's Site Address (if different than mailing address)

USAF EIELSON AFB - 354 CES/CEIC  
Site: WP45/SS57  
2881 to 2723 Flightline Blvd.

Generator's Phone:

907-377-2574

6. Transporter 1 Company Name

TOTEM OCEAN TRAILER EXPRESS

U.S. EPA ID Number

WAC070307955

7. Transporter 2 Company Name

UNION PACIFIC RAILROAD

U.S. EPA ID Number

NEC001792910

8. Designated Facility Name and Site Address

COLUMBIA RIDGE LANDFILL  
18177 CEDAR SPRINGS LANE  
ARLINGTON, OR 97812

U.S. EPA ID Number

Facility's Phone:

541-454-2030

ORD987173467

9. Waste Shipping Name and Description

10. Containers

No. Type

11. Total Quantity

12. Unit Wt./Vol.

1. MATERIAL NOT REGULATED BY DOT  
(CONTAMINATED SOILS)

01

CM

T

2.

3.

4.

13. Special Handling Instructions and Additional Information

(1) PROFILE # ORI 12211, POL / PCB CONTAMINATED SOILS AND DEBRIS

CERTIFICATES OF DISPOSAL REQUIRED

14. GENERATOR'S/OFFEROR'S CERTIFICATION: I hereby declare that the contents of this consignment are fully and accurately described above by the proper shipping name, and are classified, packaged, marked and labeled/placarded, and are in all respects in proper condition for transport according to applicable international and national governmental regulations.

Generator's/Offeror's Printed/Typed Name

Signature

Month Day Year

15. International Shipments

Import to U.S.

Export from U.S.

Port of entry/exit:

Transporter Signature (for exports only):

Date leaving U.S.:

16. Transporter Acknowledgment of Receipt of Materials

Transporter 1 Printed/Typed Name

Signature

Month Day Year

Transporter 2 Printed/Typed Name

Signature

Month Day Year

17. Discrepancy

17a. Discrepancy Indication Space

Quantity

Type

Residue

Partial Rejection

Full Rejection

Manifest Reference Number:

17b. Alternate Facility (or Generator)

U.S. EPA ID Number

Facility's Phone:

17c. Signature of Alternate Facility (or Generator)

Month Day Year

18. Designated Facility Owner or Operator: Certification of receipt of materials covered by the manifest except as noted in Item 17a

Printed/Typed Name

Signature

Month Day Year

GENERATOR

INT'L

TRANSPORTER

DESIGNATED FACILITY

NON-HAZARDOUS WASTE MANIFEST

1. Generator ID Number

AK1570028646

2. Page 1 of

3

3. Emergency Response Phone

800-424-9200

4. Waste Tracking Number

JEAFB0213-01

5. Generator's Name and Mailing Address

USAF EIELSON AFB - 354 CES/CEOIC  
2310 CENTRAL AVENUE, SUITE 100  
EIELSON AFB, AK 99702

Generator's Site Address (if different than mailing address)

USAF EIELSON AFB - 354 CES/CEOIC  
Site: WP45/SS57  
2851 to 2723 Flightline Blvd

Generator's Phone:

907-377-2574

6. Transporter 1 Company Name

CARLILE TRANSPORTATION

U.S. EPA ID Number

WAH000018747

7. Transporter 2 Company Name

TOTEM OCEAN TRAILER EXPRESS

U.S. EPA ID Number

WAD070307855

8. Designated Facility Name and Site Address

CWMNW - ARLINGTON  
17829 CEDAR SPRINGS LANE  
ARLINGTON, OR 97812

U.S. EPA ID Number

Facility's Phone:

541-454-2643

ORD099452353

9. Waste Shipping Name and Description

1. MATERIAL NOT REGULATED BY DOT  
(POL CONTAMINATED WATER)

10. Containers

No. Type

11. Total Quantity

12. Unit Wt./Vol.

DM

1735

G

13. Special Handling Instructions and Additional Information

1) PROFILE # OR322492, POL WATER

CERTIFICATES OF DISPOSAL REQUIRED

14. GENERATOR'S/OFFEROR'S CERTIFICATION: I hereby declare that the contents of this consignment are fully and accurately described above by the proper shipping name, and are classified, packaged, marked and labeled/placarded, and are in all respects in proper condition for transport according to applicable international and national governmental regulations.

Generator's/Offeror's Printed/Typed Name

Signature

Month Day Year

22 JUNE 2013

[Signature]

6 14 13

15. International Shipments

Import to U.S.

Export from U.S.

Port of entry/exit:

Date leaving U.S.:

Transporter Signature (for exports only):

INT'L

16. Transporter Acknowledgment of Receipt of Materials

Transporter 1 Printed/Typed Name

Signature

Month Day Year

X D.F. NAPPAL RAY J

[Signature]

2 17 13

Transporter 2 Printed/Typed Name

Signature

Month Day Year

TRANSPORTER

17. Discrepancy

17a. Discrepancy Indication Space

Quantity

Type

Residue

Partial Rejection

Full Rejection

Manifest Reference Number:

17b. Alternate Facility (or Generator)

U.S. EPA ID Number

Facility's Phone:

17c. Signature of Alternate Facility (or Generator)

Month Day Year

DESIGNATED FACILITY

18. Designated Facility Owner or Operator: Certification of receipt of materials covered by the manifest except as noted in Item 17a

Printed/Typed Name

Signature

Month Day Year



**NON-HAZARDOUS WASTE MANIFEST  
(Continuation Sheet)**

19. Generator ID Number

AK1570028646

20. Page

3053

21. Waste Tracking Number

JEAFR0213-01

22. Generator's Name

USAF EIELSON AFB - 354 CESKCEIC  
2310 CENTRAL AVENUE, SUITE 100  
EIELSON AFB, AK 99782

23. Transporter \_\_\_\_\_ Company Name

UNION PACIFIC RAIL ROAD

U.S. EPA ID Number

NEF001702910

24. Transporter \_\_\_\_\_ Company Name

COLUMBIA RIDGE LANDFILL

U.S. EPA ID Number

ORD987173457

25. Waste Shipping Name and Description

MATERIAL NOT REGULATED BY DOT  
(POL-CONTAMINATED WATER)

26. Containers

No. Type

27. Total  
Quantity

28. Unit  
Wt./Vol.

GENERATOR

29. Special Handling Instructions and Additional Information

TRANSPORTER

30. Transporter \_\_\_\_\_ Acknowledgment of Receipt of Materials

Printed/Typed Name

Signature

Month Day Year

31. Transporter \_\_\_\_\_ Acknowledgment of Receipt of Materials

Printed/Typed Name

Signature

Month Day Year

DESIGNATED FACILITY

32. Discrepancy





DRIVER # 37

# WEAVER BROTHERS, INC.

278-4526 • 2230 Spar Ave. • Anchorage, Alaska 99501

TRUCK # 37

## WORK ORDER

FROM WBI

TO ELEISON AFB # HAZARDOUS WASTE FACILITY # 2963

BILLING/REFERENCE # \_\_\_\_\_

TRAILER PREFIX WMXU TRAILER NUMBER 6231

DATE 2/19/13 CUSTOMER/NAME \_\_\_\_\_ PHONE # \_\_\_\_\_

BILL TO:

HLA

TOTE

CUSTOMER

CO-BIZ

ROUND TRIP TIME: START \_\_\_\_\_ AM-PM STOP \_\_\_\_\_ AM-PM \_\_\_\_\_ RESPOT

LOAD TIME: START \_\_\_\_\_ AM-PM STOP \_\_\_\_\_ AM-PM \_\_\_\_\_ ON SITE RESPOT

UNLOAD TIME: START \_\_\_\_\_ AM-PM STOP \_\_\_\_\_ AM-PM \_\_\_\_\_ HOUSEHOLD GOODS

DELAY TIME: START \_\_\_\_\_ AM-PM STOP \_\_\_\_\_ AM-PM

WORK DESCRIPTION:  LOAD  EMPTY  EMPTY FOR SOUTH BOUND  LINEHAUL SUPPORT

SPOT MT FOR SB LOADING

[Signature] CIVIL/ELM GINA LEATON

RECEIVED IN GOOD CONDITION EXCEPT AS NOTED

COMPANY NAME

SIGNATURE OF DRIVER

KASTELER

AMERICA NORTH PRINTERS • 562-6416 • FAX 563-0033

ORIGINAL • OFFICE COPY

YELLOW • ACCOUNTING

PINK • CUSTOMER

DRIVER # \_\_\_\_\_

# WEAVER BROTHERS, INC.

TRUCK # \_\_\_\_\_

278-4526 • 2230 Spar Ave. • Anchorage, Alaska 99501

## WORK ORDER

FROM WBI \_\_\_\_\_

BILL TO:

TO EAFB \_\_\_\_\_

HLA

BILLING/REFERENCE # ~~05245901~~ \_\_\_\_\_

TOTE

TRAILER PREFIX Wwxu TRAILER NUMBER 6273 (CLASSIC FVX2733129) \_\_\_\_\_

CUSTOMER

DATE 2/19/13 CUSTOMER/NAME JACOBS/Ida Petersen PHONE # 907 451-0550 \_\_\_\_\_

CO-BIZ

ROUND TRIP TIME: START \_\_\_\_\_ AM-PM STOP \_\_\_\_\_ AM-PM \_\_\_\_\_ RESPOT

LOAD TIME: START \_\_\_\_\_ AM-PM STOP \_\_\_\_\_ AM-PM \_\_\_\_\_ ON SITE RESPOT

UNLOAD TIME: START \_\_\_\_\_ AM-PM STOP \_\_\_\_\_ AM-PM \_\_\_\_\_ HOUSEHOLD GOODS

DELAY TIME: START \_\_\_\_\_ AM-PM STOP \_\_\_\_\_ AM-PM

WORK DESCRIPTION:  LOAD  EMPTY  EMPTY FOR SOUTH BOUND  LINEHAUL SUPPORT

\_\_\_\_\_  
Empty to EAFB for loading

Ida Petersen (JACOBS) \_\_\_\_\_ Anchorage 2/19/13  
RECEIVED IN GOOD CONDITION EXCEPT AS NOTED COMPANY NAME SIGNATURE OF DRIVER