



Prepared for the Alaska Department of Environmental Conservation

Evaluate Fecal Coliform Bacteria for Two Anchorage Watersheds

Alaska Clean Water Actions Grant # 18-07– Task 8

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February 28, 2019

This project has been funded wholly or in part by the United States EPA under assistance agreement number 00J84603 to the Department of Environmental Conservation (DEC) through the Alaska Clean Water Actions (ACWA) program. The contents of this document do not necessarily reflect the views and policies of the EPA or DEC, nor does the EPA or DEC endorse trade names or recommend the use of commercial product mentioned in this document.

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Introduction

This report summarizes the findings from ACWA grant 18-07 for task 8 on Chester Creek and Little Campbell Creek within the Municipality of Anchorage (MOA). Anchorage Waterways Council's (AWC) water quality monitoring program is described as well as how the fecal coliform bacteria (FC) monitoring data between 2010 and 2017 was vetted through Quality Assurance and Quality Control procedures (QA/QC). A discussion follows regarding the use of the FC data for screening or analytical purposes. Included in the report are four GIS maps depicting monitoring sites and potential sources of fecal coliform from different factors. The conclusion covers areas where additional monitoring would be useful.

Anchorage Waterways Council's Monitoring Program

AWC began its volunteer water quality monitoring program in 1998 when Robert Shipley, a board member and biologist, took the first samples. Known as CEMP, citizens' environmental monitoring program, to date it has trained over 300 volunteer monitors to collect baseline water quality data which is used to identify trends and detect pollution. About half of Anchorage's 26 watersheds have had some monitoring, and several creeks have had multiple sites monitored. Baseline goals for individual sites are to achieve 5 years of monitoring with a total of 20 monthly site visits during winter months (monitoring cannot be achieved when the site is frozen, but we still require visits) and a total of 40 monthly site visits during summer over the 5 year period.

AWC's monitoring protocols have changed over the years as has the equipment. Initially, Hanna meters were used which necessitated 3-day trainings and frequent recertifications. In 2016, the lack of adequate funding for the program, the expense of replacing Hanna meters every 18 months, and the cost for shipping hazardous chemicals by 2nd day air made the existing program nonviable. In order to keep our monitoring program going, some changes had to be made. As an example, pH testing had been done by a colorimetric system as well as by a Hanna meter which needed 3 chemicals just for calibrating. pH testing is now done solely by using test strips, and they seem to be fairly comparable to colorimetric tests.

Dissolved oxygen (DO), previously done through titration (which uses sulfuric acid), was changed to a comparator system. Ampoules that have been vacuum-sealed with a chemical reagent are inserted into a vial containing the sample. The tip of the ampoule is broken off and within seconds the ampoule sucks in the water and changes to a shade of blue which is then matched to a color/reading in the comparator. It's usually not crucial to worry about the difference between a 7 or 8. What is important is to make sure the DO is not 6 or less. And that is easily discernable.

There have been no changes in the fecal coliform (FC) testing which uses Micrology's Coliscan® or measuring turbidity with LaMotte's Standard Turbidity Reagent. These remain according to the original Quality Assurance Project Plan (QAPP) approved by the Alaska Department of Environmental Conservation (DEC) in 2008. It must be remembered that these tests are used as water quality screening and a means of alerting AWC to potential problems, and, if necessary, the need for more extensive testing. It would require a much greater cost per sampling to have laboratory analytical results all the time.

In reviewing the differences between AWC's previous testing and that which began in 2016— we have reduced the need for chemical reagents from ten to one (turbidity) which has given our program the ability to continue. And we are satisfied with the quality of the results¹ for screening purposes.

¹ Calibrating Hanna meters and performing DO titration require careful attention and skill by the monitor. This probably was the case with about half the monitors, the other half had difficulty performing the calibrations and test, which resulted in data that was not always the best.

Anchorage Waterways Council CEMP Water Quality Data Sheet Revised April 1, 2011						Office Use Only							Page 1 of 2					
						Entry Date			Edit		Comments							
Sample Information																		
Date					Site ID					Collection Time								
Monitoring Kit Number							Kit Condition											
Volunteer Information																		
Print Name					Signature					Mileage		Hours						
Volunteer 1																		
Volunteer 2																		
Exp. Dates		pH 4				Mang. Sulf.				Starch Ind.								
		pH 7				Alk. Pot.				Thiosulf.								
WR Ind.		Cond.				Sulf. Acid				Turb. Rgt.								
Hanna Meter Calibration						Weather			Wind			Sample Location						
Meter #				Date				Clear Partly Cloudy Cloudy Precipitation Fog or Haze			<u>Mph</u>		<u>Direction</u>		<u>Depth</u>		<u>Bottom</u>	
pH 7 (Initial)		Temp		pH 4 (Cal)		Temp					73 or greater		Variable		Pool		Rifle	
Cond. (initial)		Temp		Cond. (Cal)		Temp												
Precipitation						Type (circle one)			# of Days									
Last 24 hr.		Yes		Rain		Hail		Similar										
		No		Snow		Sleet		Temp. °F										
Comments						Sketch												
Photos																		
Photo #		Photo Description								Camera #								
#																		
#																		
#																		

5

Please Return ASAP to: Anchorage Waterways Council P.O. Box 241774 Anchorage, Alaska 99524-1774				Date 		Page 2 of 2													
Water Temperature Repeat if not within 2.0 °C <div style="display: flex; justify-content: space-between;"> Replicate 1 Replicate 2 Location (circle one) </div> <div style="display: flex; justify-content: space-between;"> <div style="width: 15%;"> Temp °C Time </div> <div style="width: 15%;"> <div style="border: 1px solid black; height: 20px; margin-bottom: 5px;"></div> <div style="border: 1px solid black; height: 20px;"></div> </div> <div style="width: 20%;"> Stream Bucket </div> </div> <p style="font-size: small; text-align: center;">Two replicates should be taken 5 minutes apart</p>				Color 2.5 Gallon 50 ml <div style="display: flex; justify-content: space-between;"> Apparent </div> <div style="display: flex; justify-content: space-between;"> Color </div> <div style="display: flex; justify-content: space-between;"> <div style="width: 15%;"></div> <div style="width: 15%;"></div> </div>															
Turbidity <div style="display: flex; justify-content: space-between;"> <div style="width: 30%;"> Sample Size (circle one) <div style="display: flex; justify-content: space-around;"> <div style="border: 1px solid black; padding: 2px 10px;">25 ml</div> <div style="border: 1px solid black; padding: 2px 10px;">50 ml</div> </div> </div> <div style="width: 30%;"> # of Additions Water Temp °C </div> <div style="width: 30%;"> <div style="display: flex; justify-content: space-around;"> <div style="border: 1px solid black; padding: 2px 10px;">Replicate 1</div> <div style="border: 1px solid black; padding: 2px 10px;">Replicate 2</div> </div> </div> </div> <p style="font-size: small; text-align: center;">Repeat if replicates are not within 1 addition of each other</p>						Colorimetric pH <div style="display: flex; justify-content: space-around;"> <div style="border: 1px solid black; padding: 2px 10px;">Replicate 1</div> <div style="border: 1px solid black; padding: 2px 10px;">Replicate 2</div> </div> <p style="font-size: small; text-align: center;">Record to nearest 0.25 pH units Repeat if replicates not within 0.25 units</p>													
Hanna Meter Wait for Hanna meter to stabilize before recording measurement. Record time when Hanna meter is placed in bucket/stream. Allow 15 seconds to stabilize between 3 replicate readings. Minimum of 2 replicates must be in the following ranges of each other, repeat outlier: pH 0.05 units, Conductivity 4µs, TDS 4 ppm.																			
Meter # 		<div style="display: flex; justify-content: space-around;"> <div style="width: 45%;"> <div style="display: flex; justify-content: space-around;"> <div style="border: 1px solid black; padding: 2px 10px;">start</div> <div style="border: 1px solid black; padding: 2px 10px;">stop</div> </div> <div style="display: flex; justify-content: space-around;"> <div style="border: 1px solid black; padding: 2px 10px;">Temp °C</div> <div style="border: 1px solid black; padding: 2px 10px;">Time</div> </div> </div> <div style="width: 5%; text-align: center;"> pH Cond TDS </div> <div style="width: 45%;"> <div style="display: flex; justify-content: space-around;"> <div style="width: 30%;"> Replicates <table border="1" style="width: 100%; border-collapse: collapse;"> <tr> <th style="font-size: small;">Rep. 1a</th> <th style="font-size: small;">Rep. 2a</th> <th style="font-size: small;">Rep. 3a</th> </tr> <tr><td style="height: 20px;"></td></tr> <tr><td style="height: 20px;"></td></tr> <tr><td style="height: 20px;"></td></tr> </table> </div> <div style="width: 30%;"> Replicates <table border="1" style="width: 100%; border-collapse: collapse;"> <tr> <th style="font-size: small;">Rep. 1b</th> <th style="font-size: small;">Rep. 2b</th> <th style="font-size: small;">Rep. 3b</th> </tr> <tr><td style="height: 20px;"></td></tr> <tr><td style="height: 20px;"></td></tr> <tr><td style="height: 20px;"></td></tr> </table> </div> </div> </div> </div>						Rep. 1a	Rep. 2a	Rep. 3a				Rep. 1b	Rep. 2b	Rep. 3b			
Rep. 1a	Rep. 2a	Rep. 3a																	
Rep. 1b	Rep. 2b	Rep. 3b																	
Comments: 																			
Dissolved Oxygen Minimum of 2 replicates must be within 0.6 mg/L of each other, repeat outlier.																			
Fix Time 		Fix Temperature °C 		Titration Date 		Titration Time 													
Replicate 1a 		Replicate 2a 		Replicate 3a 		Replicate 1b 													
Replicate 2b 		Replicate 3b 		Replicate 1c 		Replicate 2c 													
Comments: 																			
Coliform Bacteria																			
Time mixed 		Date counted 		Total E. coli Colonies		<div style="display: flex; justify-content: space-around;"> <div style="border: 1px solid black; padding: 2px 10px;">1 ml</div> <div style="border: 1px solid black; padding: 2px 10px;">5 ml</div> </div> <div style="text-align: right; font-size: small;">(blue/purple)</div>													
Time plated 		Time counted 		Total pink/red colonies		<div style="border: 1px solid black; display: inline-block; width: 50px; height: 20px;"></div>													
Easygel Exp. 		Plate Exp. 		Total teal Colonies		<div style="border: 1px solid black; display: inline-block; width: 50px; height: 20px;"></div>													

Figure 2. Page 2 of data sheet used until 2016

Anchorage Waterways Council CEMP Water Quality Data Sheet Revised April 1, 2016				Page 1 of 2		
				Office Use Only		
				Entry Date	Edit	Comments
Sample Information						
Date		Site ID	Ma	Collection Time		
Monitoring Kit Number			Kit Condition			
Volunteer Information						
Print Name		Signature		Mileage	Hours	
Volunteer 1						
Volunteer 2						
Weather	Precipitation		Type	# of days similar		
Clear	Last 24 hr.	Yes	Rain			
Partly Cloudy		No	Snow			
Cloudy			Hail			
Precipitation			Sleet			
Fog or Haze	Air Temp °C					
Sample Location				Comments		
Depth						
0-6"						
6-12"						
12-36"						
Bottom						
Silty						
Sandy						
Gravel						
Cobble						
Description						
Pool						
Riffle						
Photos						
	Photo #	Photo Description				
	#					
	#					
	#					
	#					

Figure 3. Page 1 of data sheet used in 2016

Please Return ASAP to: Anchorage Waterways Council P.O. Box 241774 Anchorage, Alaska 99524-1774				Date 		Page 2 of 2	
				Site ID Ma			
Water Temperature Temp °C <input type="text"/> Time <input type="text"/>				Location (circle one) Stream <input type="checkbox"/> Bucket <input type="checkbox"/>		Color Apparent Color 2.5 Gallon <input type="text"/> 50 ml <input type="text"/>	
Turbidity						pH Strips	
Sample Size (circle one) 25 ml <input type="checkbox"/> 50 ml <input type="checkbox"/>		Reagent exp <input type="text"/>	Replicate 1 <input type="text"/>	Replicate 2 <input type="text"/>	Replicate 3 <input type="text"/>	Replicate 1 <input type="text"/>	Replicate 2 <input type="text"/>
		# of Additions <input type="text"/>	<input type="text"/>	<input type="text"/>	<input type="text"/>	<input type="text"/>	<input type="text"/>
Remember to SHAKE the turbidity reagent well beforehand.						Do 2 readings, and record to nearest 0.5 pH units. Do a 3rd replicate if values are not within 0.5 units	
Repeat if first 2 replicates are not within 1 addition of each other							
Dissolved Oxygen							
DO # (PPM, mg/L) <input type="text"/>							
Bucket Temperature °C <input type="text"/>							
Coliform Bacteria		5 ml sample					
Time mixed <input type="text"/>	Date counted <input type="text"/>	Total E. coli Colonies <input type="text"/>		(blue/purple)			
Time plated <input type="text"/>	Time counted <input type="text"/>	Total pink/red colonies <input type="text"/>					
Easygel Exp. <input type="text"/>	Plate Exp. <input type="text"/>	Total teal Colonies <input type="text"/>					
Site Sketch							

Figure 4. Page 2 of data sheet used in 2016

QA/QC of Monitoring Data Entry

AWC has close to 20 years of water monitoring data for the Chester and Little Campbell Creek watersheds, however the only records that were used for this ACWA grant were those from 2010 to 2017. The reason is that the original Access database that had monitoring data entered between 1999 and 2009 was found to be extremely cumbersome with too much information entered, much of it unnecessary, and a considerable amount of what had been entered was fraught with errors.

In 2010 when Dr. Cherie Northon became the Executive Director and Dr. Thom Eley became the monitoring coordinator, a new data entry system was created using Google Docs. It was much less complicated yet covered all the parameters needed. Interns entering the data were given training and clear instructions on how to do so.

Even so, errors were still introduced. AWC's UAA intern, Veronica Campbell, was with us for 3 semesters and she was tasked with reviewing all the hard copy data sheets from 2010 to 2017 to the new database. After she completed checking the records, Dr. Northon reviewed all of her comments for sites in Little Campbell Creek and Chester Creek—and then made the corrections in the database herself. Our confidence level in the data accuracy is quite high for these two watersheds.

Screening or Analytical Water Quality Data

As discussed above, AWC is confident that the water quality data collected from 2010-2017 is useful for screening potential water quality problem areas. There is no doubt that using a handheld meter for pH is more accurate than test strips, but a sampling result of concern that did not meet Alaska Water Quality Standards (WQS) would most likely be identified and followed up on by additional more advanced water testing techniques. As mentioned, the DO testing would also capture any outliers (<6.5 or >8.5) that are detrimental to the Growth and Propagation of Fish, Shellfish, Other Aquatic Life, and Wildlife from the WQS.

One of the most important correlations is that of FC testing. SGS Laboratories in Anchorage has provided AWC with some lab-quality FC testing at no cost for some situations. We have done some side-by-side testing with SGS and Micrology's Coliscan® and found them to be highly comparable. Again, if there was a concern about a high FC result using the Coliscan screening methodology, there is always an opportunity to have follow up samples analyzed at a laboratory provided there is funding.

GIS Maps

Tasks 4 and 5 of ACWA 18-07 required a GIS base map with hydrology, AWC monitoring sites for Chester Creek and Little Campbell Creek, as well land use, pet waste stations (previous ACWA grant), and animal facilities and events created for the MOA's Watershed Management Service's APDES permit AKS-052558. AWC is providing the following shapefiles that we have created:

1. AWC-2018-DogParks.shp
2. AWC-MonitoringSites.shp
3. AWC-PetWasteStations.shp
4. AWC_Stables – Zoo.shp
5. AWC-AnimalFacilities.shp

Additionally, there are shapefiles from MOA WMS that have been altered to include only things that we need. An example would be MOA-ChesterCreekWatershed. It was originally a shapefile with all of Anchorage's watersheds. These will all be provided to DEC.

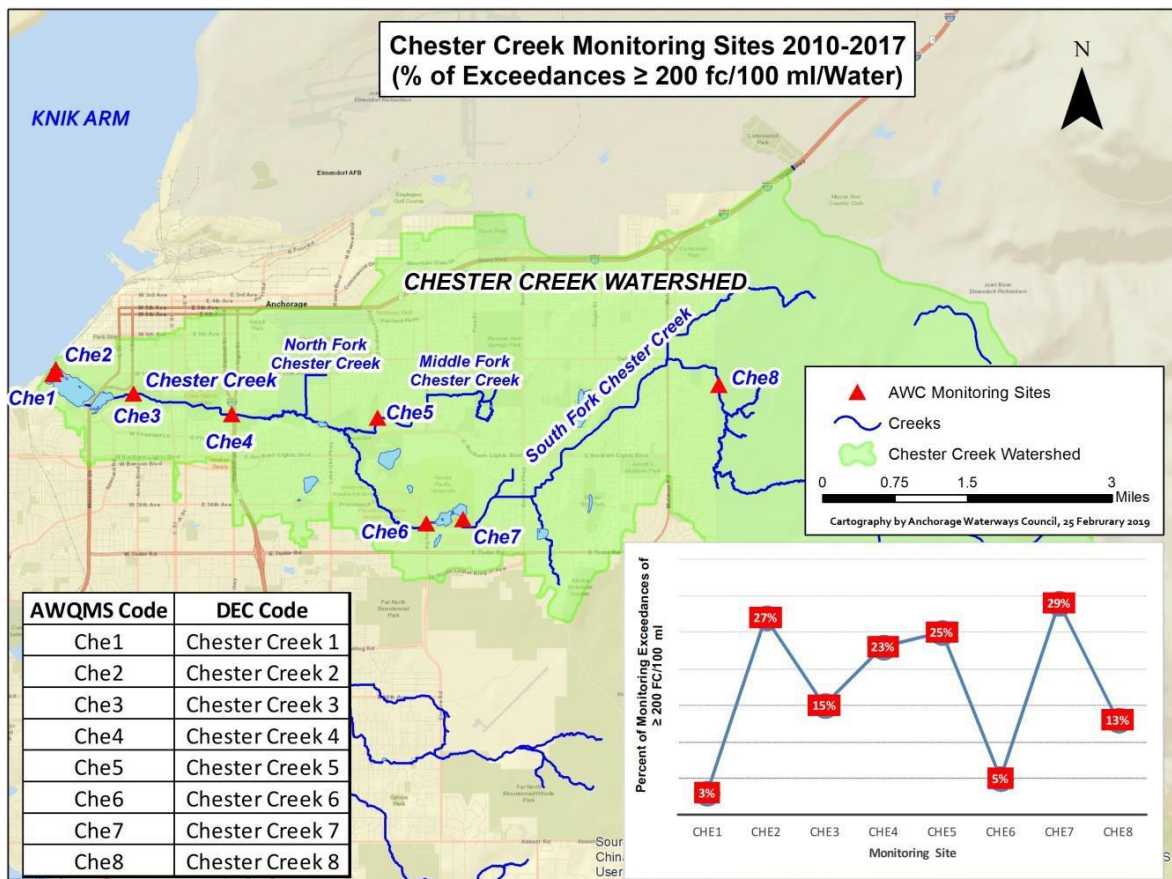
The following GIS maps are in the Appendix.

1. Commercial Stables, the Alaska Zoo, Trails and LCC Monitoring Sites (This map is specific to Little Campbell Creek because there are no commercial stables in the Chester Creek watershed.)
2. Indoor Animal Facilities, Pet Stores, Kennels and Monitoring Sites
3. Dog Parks, Pet Waste Stations, and Monitoring Sites
4. Landuse and Monitoring Sites in the Chester Creek and Little Campbell Creek Watersheds

Discussion

Chester Creek

Fecal Coliform (FC) bacteria data from 2010 to 2017 using the Coliscan method for Chester Creek watershed covers eight monitoring sites. These sites are shown in Map 1 along with a graph of the percentage of times that the FC sample was equal to or greater than WQS's 200 FC/100 ml/water for secondary recreation and not more than 10% of the total samples may exceed 400 fecal coliform/100 ml. Because the number of monitoring events varies by site, a percentage puts them on a more even basis (Table 1).



Map 1. Chester Creek Monitoring Sites 2010-2017 (% of Exceedances ≥ 200 fc/100 ml/Water)

Table 1. Chester Creek Monitoring Sites 2010-2017 (% of Exceedances ≥ 200 fc/100 ml/water)

Monitoring Site	Total # Monitoring Events	Exceedances ≥ 200 FC/100 ml	% Exceeding
Che1	39	1	3%
Che2	11	3	27%
Che3	26	4	15%
Che4	22	5	23%
Che5	4	1	25%
Che6	58	4	7%
Che7	14	4	29%
Che8	24	3	13%

This is an interesting exercise when these values are placed in their creek locations. The two lowest exceedances are at the outfall of Westchester Lagoon and University Lake. Site Che3 is at Arctic and 17th just below Valley of the Moon Park. It typically has

relatively high FC numbers, yet when the water exits the lagoon at Che1—the numbers have dropped dramatically. The same is true for Che5 which is the inflow into University Lake and Che6 which is the outflow from the lake at Elmore. This is even more remarkable because the lake is surrounded by a popular dog park and waterfowl are present.

When looking at the data for the highest readings by site, one of the most striking anomalies is the reading for Che8 at 2,000 FC in August 2010. This is the highest reading of all monitoring events that have been taken by AWC on Chester Creek in that time frame, and it is just downstream of military land which has no housing or other forms of development except for a few roads, trails, and buildings.

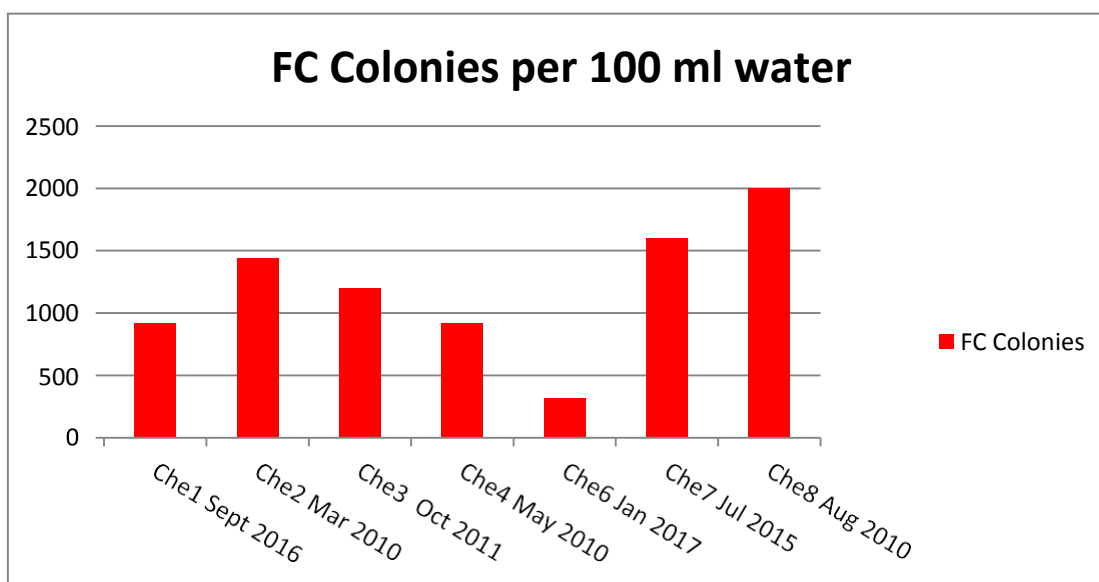
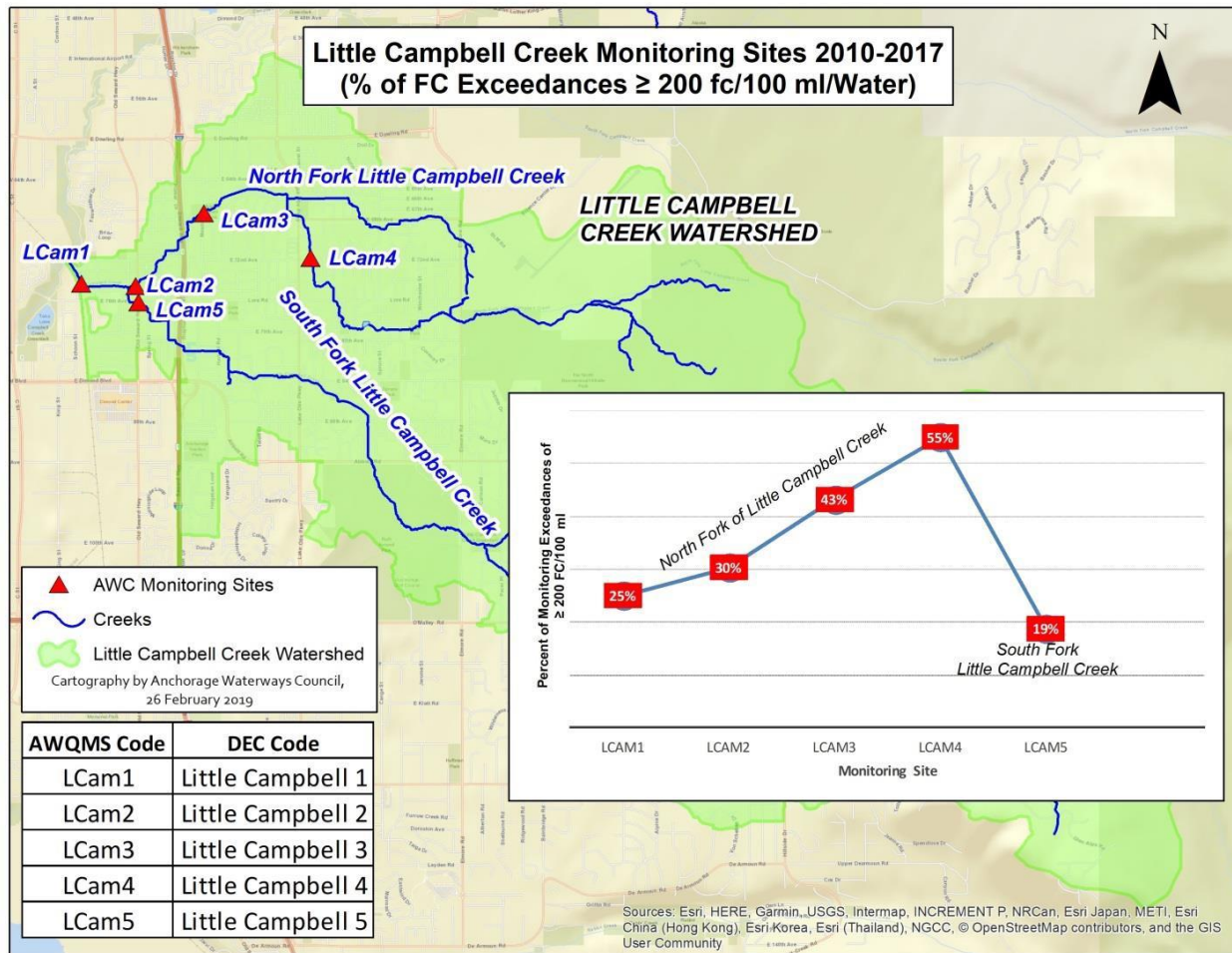


Figure 5. Highest recorded site readings by date for FC monitoring from 2010 to 2017 – Chester Creek

Little Campbell Creek

Fecal coliform bacteria data from 2010 to 2017 for Little Campbell Creek watershed covers five monitoring sites. These sites are shown in Map 2 along with a graph of the percentage of times that the FC sample was equal to or greater than WQS's 200 FC/100 ml/water for secondary recreation and not more than 10% of the total samples may exceed 400 fecal coliform/100 ml. Because the number of monitoring events varies by site, a percentage (Table 2) puts them on a more even basis.



Map 2. Little Campbell Creek Sites 2010-2017 (% of Exceedances ≥ 200 fc/100 ml/water)

Table 2. Little Campbell Creek Monitoring Sites 2010-2017 (% of Exceedances ≥ 200 fc/100 ml/water)

Monitoring Site	Total # Monitoring Events	Exceedances ≥ 200 FC/100 ml	% Exceeding
LCam1	16	4	25%
LCam2	33	10	30%
LCam3	37	16	43%
LCam4	29	16	55%
LCam5	16	3	19%

Figure 6 is also an interesting graph. When highest recorded values are placed in their creek locations, rather than showing an increase as the North Fork Little Campbell Creek runs downstream to Campbell Creek, the number of FC colonies actually decreases. The

highest exceedance is on the South Branch of the North Fork which is about 1.4 miles from Elmore Road an area bounded by BLM at Campbell Creek, Abbott Loop Community Park, and Far North Bicentennial Park. Once the creek enters at Elmore Road, it runs primarily through a residential area except for a 10 acre parcel known as the F Bar J Ranch which boards horses (Figure 7). A report by Davis and Davis for DEC suggests that this stable could be contributing bacteria to the North Fork Little Campbell Creek (2010:21).

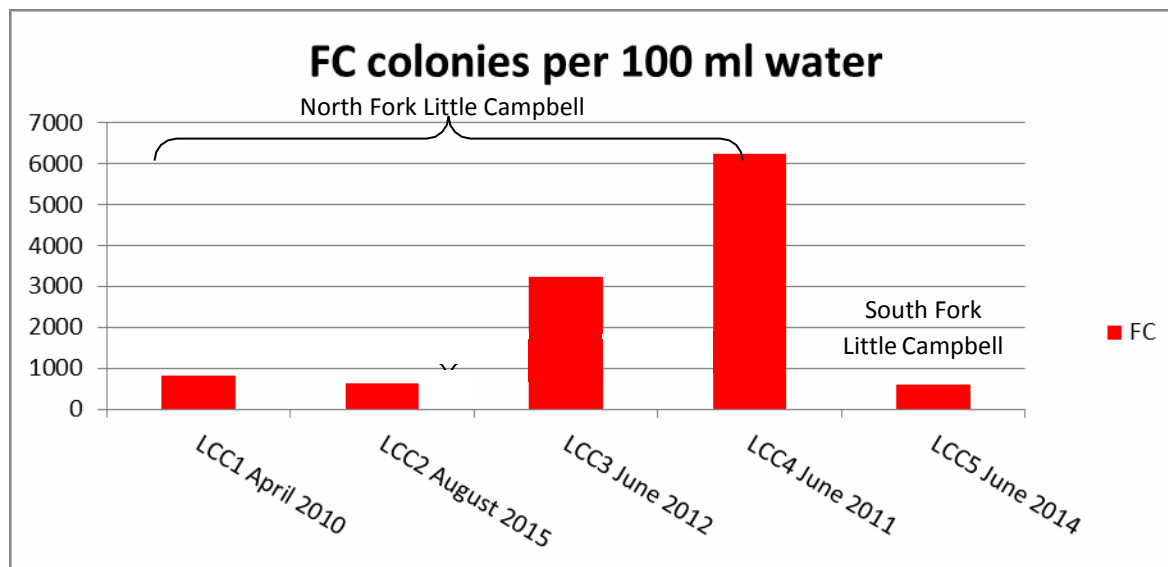


Figure 6. Highest recorded site readings by date for FC monitoring from 2010 to 2017 – Little Campbell Creek

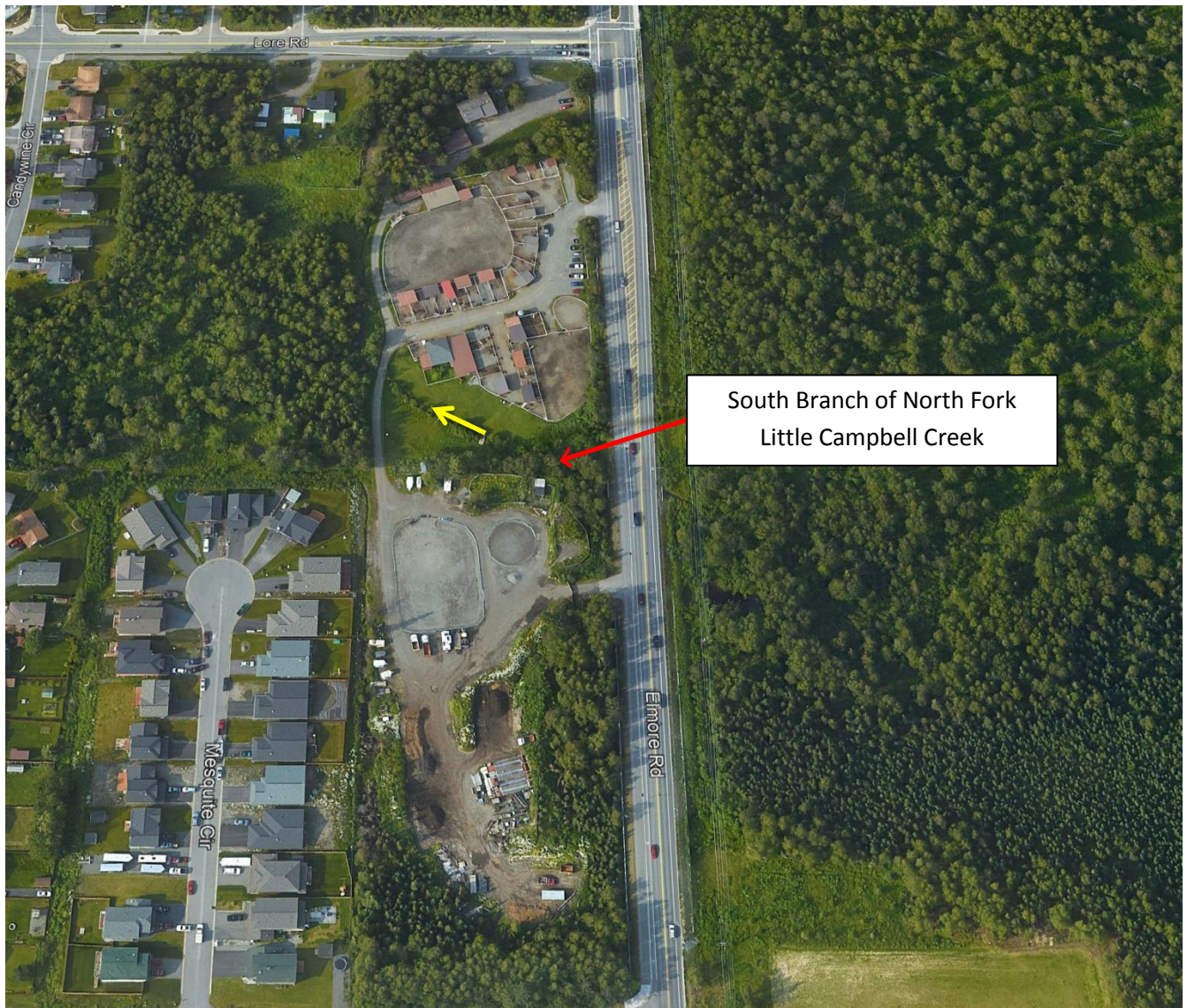


Figure 7. F Bar J Ranch with the South Branch of the North Fork Little Campbell Creek. The yellow arrow shows the direction of creek flow. Monitoring station LCam4 is 1.4 miles downstream.

Analysis

Chester Creek

Trying to account for the FC readings that don't seem to "fit" logically is difficult. The exceedances were checked against precipitation. In about half the cases there was no precipitation on the monitoring day or the previous 3 days for high FC readings, and the rest of them there was. A quick literature review did show that precipitation is not always a factor for high fecal counts.

Another way to look at the area involves the stormwater drainage maps by the Municipality from their Municipal Drainage Viewer. This is an online GIS that the Municipality uses to show stormwater drainage areas. It can be found at anchoragestormwater.com/maps.html Municipal Drainage Viewer. One feature that it can portray is the polygon boundary of a drainage area into an outlet. The following maps were created with this program.

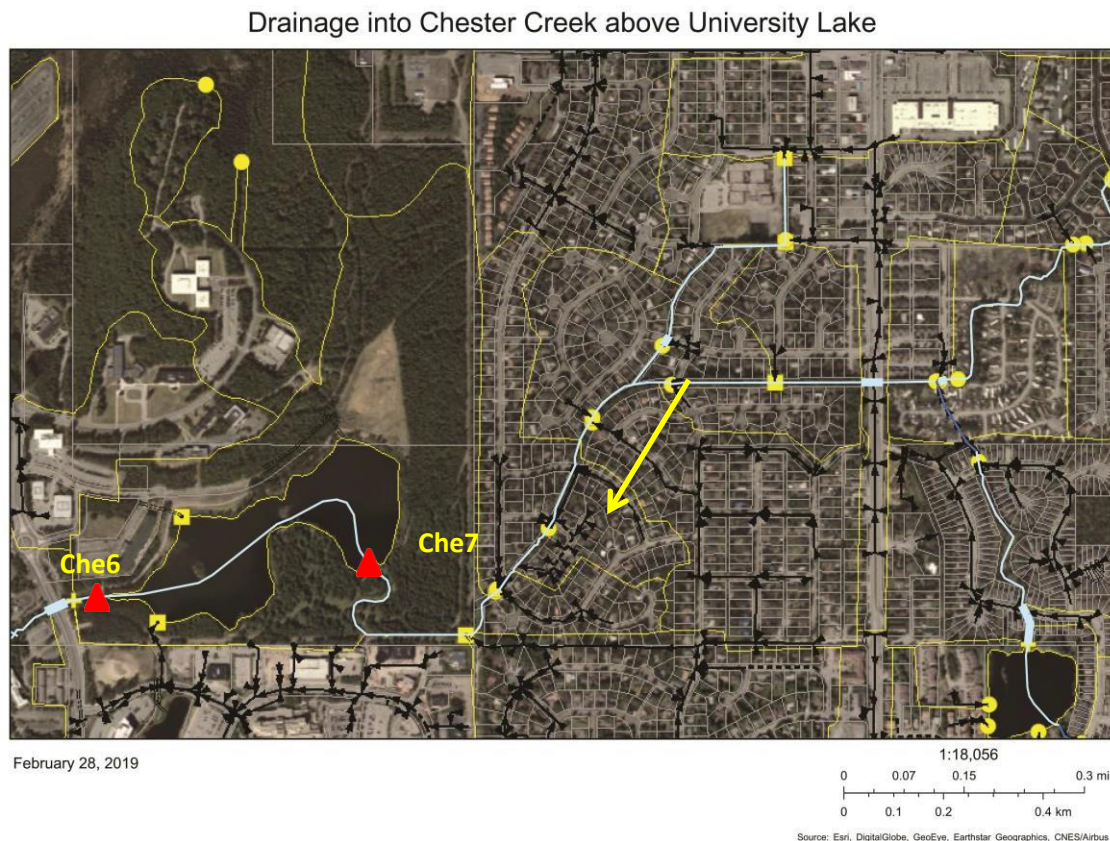


Figure 8. Monitoring sites above and below University Lake in the lower left of this image (MOA Drainage Viewer). The yellow arrow shows the direction of stream flow.

Figure 8 shows monitoring sites Che6 and Che7 in relation to University Lake. The FC readings in Che7 (upstream) have always been considerably higher than those at Che6—below the University Lake Dog Park. It is important to see what might be contributing to the high FC readings at this location. Land use shows primarily residential as well as the Alaska Native Medical Center campus. South Fork Chester Creek leaves military lands and enters the residential area of Anchorage at Early View Dr. above Windsong Park by site Che8. It then runs about 3.5 miles until the Che7 monitoring site where it enters University Lake on the east. The surrounding area that it runs through is primarily residential including Riviera Terrace Trailer Park and Begich Middle School, both of which appear to be on AWWU sewers.

There is one tributary, the North Branch of the South Fork Chester Creek that joins the South Fork Chester Creek just south of Debarr Road and Muldoon Road. It runs along half of the eastern boundary and all of the southern boundary of the Rangeview Mobile Home Park—which is entirely on septic according to Anchorage Water and Wastewater Utility's (AWWU) GIS data set "Parcels_No_AWWU_Sewer". From a literature search it appears that the trailer park was in use at least as early as 1966. Recommendation one: test for FC above and below Rangeview Mobile Home Park.

Another tributary, North Fork Chester Creek, flows south under Merrill Field in a culvert, exits at 15th and Lake Otis, and runs in a ditch due west to Sitka St. and then due south until it joins Chester Creek. The former Merrill Field landfill is located just north of 15th Ave., and it was used from the 1940s until 1987 when it was closed. It covers about 200 acres with a depth of 30'. According to a 1990 USGS report by Brunett, there has been concern about leachate which was tested for in 1989. That report concludes that while there are minor amounts of contaminants reaching much of the wetlands between 15th and Chester Creek, their concentration in the groundwater is generally less than U.S. EPA standards for drinking water (1990:1). Today this area holds Sitka Street Park which is classified as Class A wetlands and a municipal snow dump.

Although there would be no FC issues from the leachate, the snow dump and park area could be contributing FC to the North Fork Chester Creek before it converges with Chester Creek. Recommendation two: the areas above the snow dump and sites below it should be tested for FC.



Map 3. Chester Creek by Merrill Field and Former Landfill

The other anomaly on Chester Creek at Che8, which is at the boundary of military land and a residential area, remains unidentified as to the source of two high readings: 2000 FC on August 29, 2010 and 1200 FC on September 26, 2010. There are no houses, parks, stables, or anything that might cause readings this high. The only activity is on the military land which mostly has wildlife on it, although some civilians do access it for recreation. Just below the monitoring site at Windsong Park and by its outlet in South Fork Chester Creek there have been some large beaver colonies beginning around 2000 and which are no longer there. Figure 9 shows a lodge in the pond in June 2006, and Fig. 10 shows the location of the monitoring site and a beaver lodge. Perhaps there is/was a beaver lodge

upstream, but this is not an area that is open to civilians for exploration. Even so, it is obvious that this area is used by kids playing, bicyclists, hikers, and others.



Figure 9. Beaver lodge in the Windsong sedimentation basin June 6, 2006 (Photo by C. Northon)

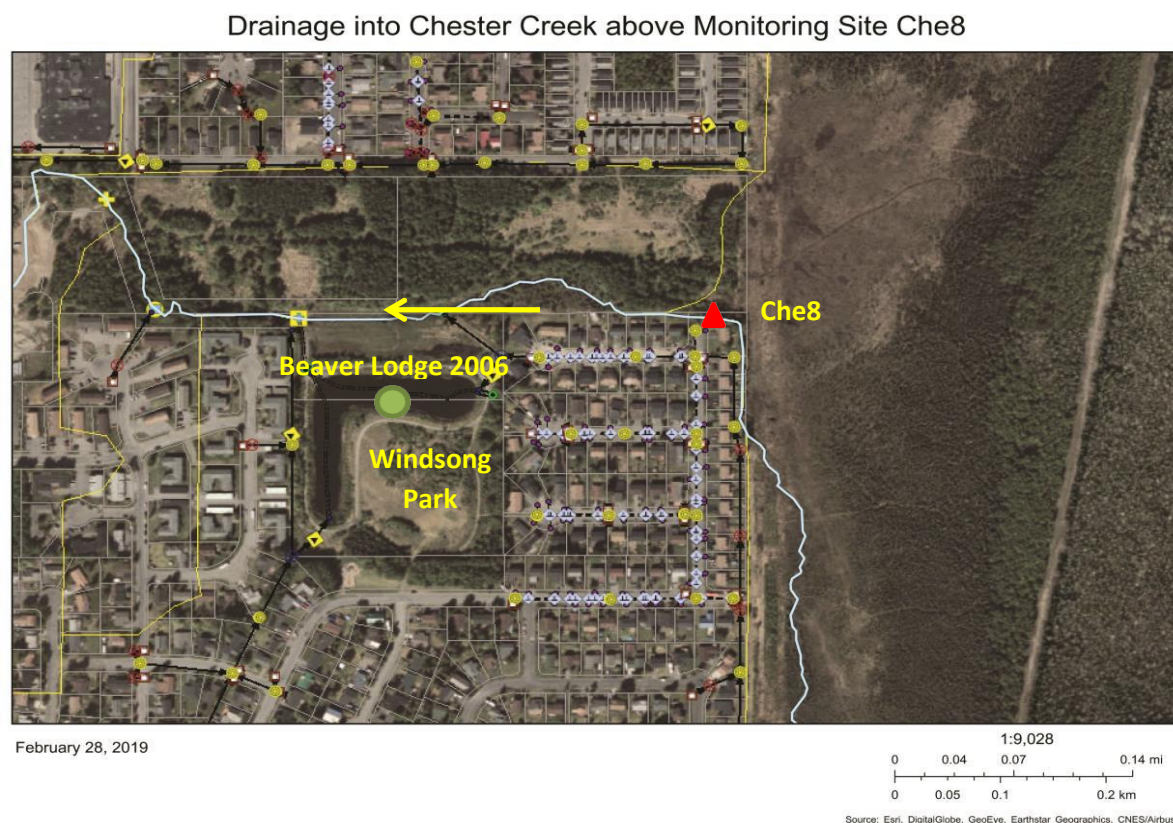


Figure 10. Drainage into Chester Creek above AWC monitoring site Che8 (MOA Drainage Viewer). The yellow arrow shows the direction of stream flow.

Little Campbell Creek

Little Campbell Creek also has a data anomaly not easily explained by weather. In Fig. 11, LCam 4 is the farthest east monitoring site on the North Fork Little Campbell Creek. It's actually on the South Branch North Fork, and is about 1.4 miles below Elmore Road which is bounded by BLM at Campbell Creek, Abbott Loop Community Park, and Far North Bicentennial Park. As noted above, the creek then runs through the F Bar J Ranch. Other than the park areas east of Elmore Road—there isn't much that could explain the high FC counts.

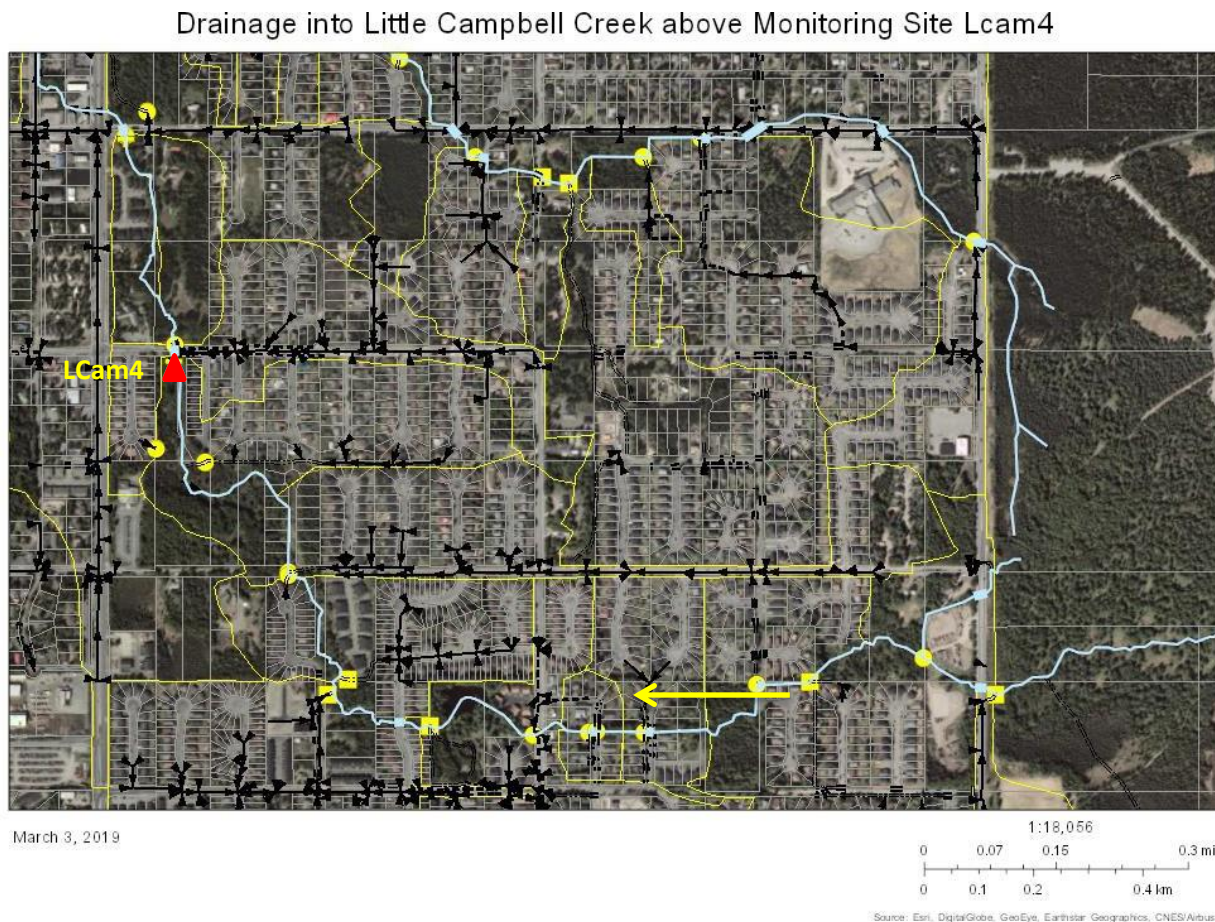


Figure 11. Drainage into Little Campbell Creek above monitoring site LCam4 (MOA Drainage Viewer). The yellow arrow shows the direction of flow.

Conclusions and Recommendations

One conclusion that can be drawn from this study is that what seems to be the obvious—is not always the case. High FC counts aren't necessarily found downstream of dog parks, and yet they can be found downstream of unoccupied land, and FC counts can be high during precipitation and as well as non- precipitation events. This suggests that other possible bacteria sources should be considered.

Accordingly, such things as storm drains, septic sewer systems, wildlife, and land use are beneficial for postulating the reason for high FC counts. Based on this study, AWC believes that there are many factors contributing to sporadic high FC counts. In the Chester and Little Campbell Creek watersheds, it would be useful to test above and below: stables, those animal facilities (Map 2 in Appendix) that are adjacent to or near streams, mobile home communities using onsite septic systems, the Alaska Zoo, snow dumps, and single family residential areas with extensive septic systems. To make these tests even more useful, tests using Microbial Source Tracking (MST) in some cases, e.g. stables and areas with extensive and older septic systems, could be performed.

AWC was tentatively awarded an ACWA grant for FY19-21, “Increase Knowledge on Anchorage Bowl Watersheds”, to collect additional bacteria information to answer some of the questions posed in this data analysis report. Funding for this proposed work is currently on hold due to state budget cuts.

AWC is putting forward the following specific recommendations:

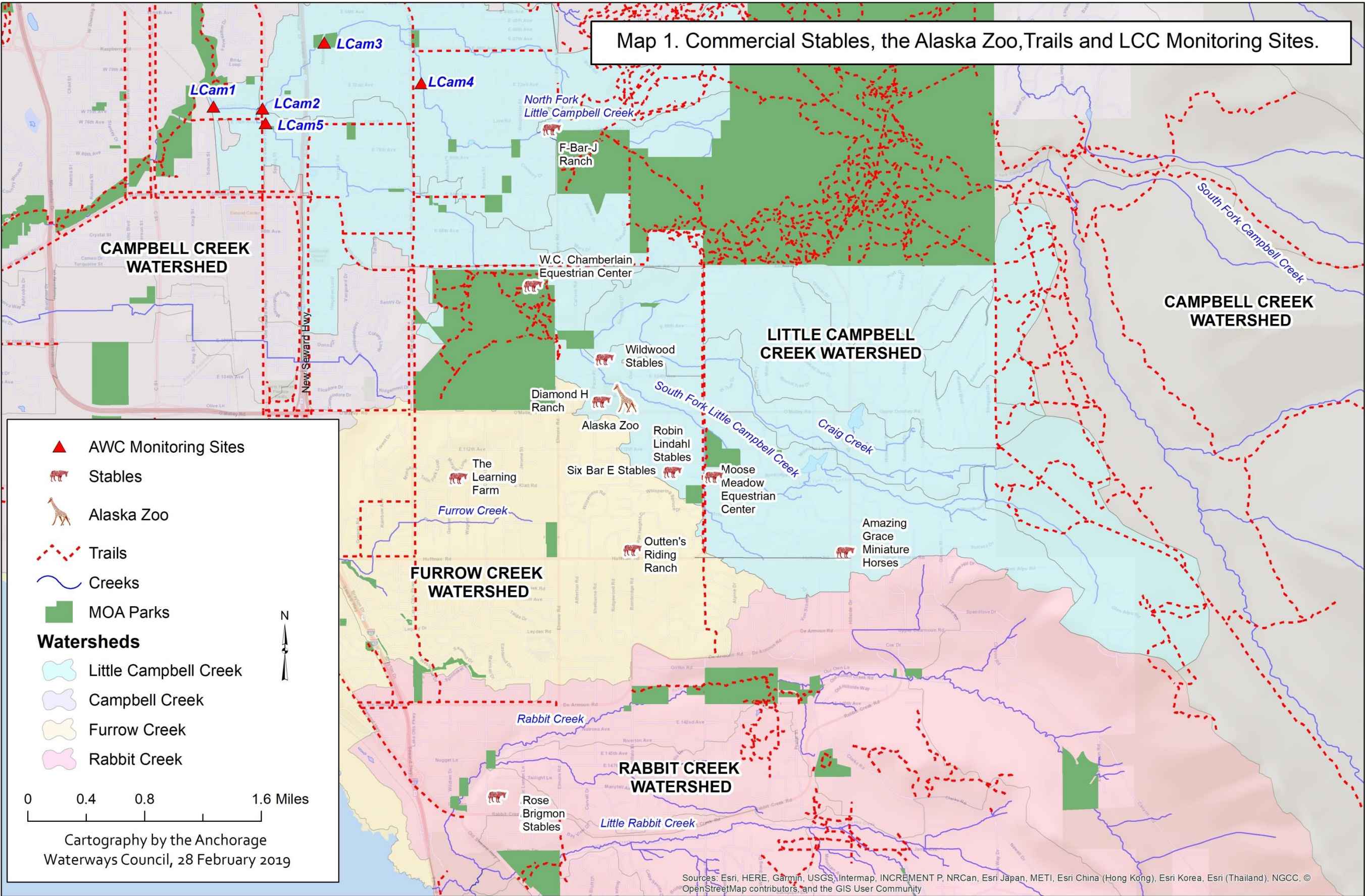
1. Test for FC above and below Rangeview Mobile Home Park along Chester Creek.
2. The areas above the snow dump and sites below it on Middle Fork Chester Creek should be tested for FC.
3. The area below Che8 should be tested using MST.
4. On Little Campbell Creek, the area above and below the F Bar J Ranch on Elmore Road should be tested for FC, and if it is higher at the downstream side—then test for MST to confirm if it is because of the stable.
5. Test for FC above and below the Alaska Zoo¹ on the South Fork Little Campbell Creek.

¹ Sites above and below the Alaska Zoo were monitored for FC on the South Fork Little Campbell Creek by AWC in 2017 after noticing that during heavy precipitation the creek was running through a portion of the brown bear pen. In addition, several of the surrounding large animal exhibits sit above the creek. It was monitored 5 times between May 30 and September 11 and analyzed by SGS Labs. Three of the five monitoring events had higher FC counts below the zoo, and two here higher above the zoo. Again, an

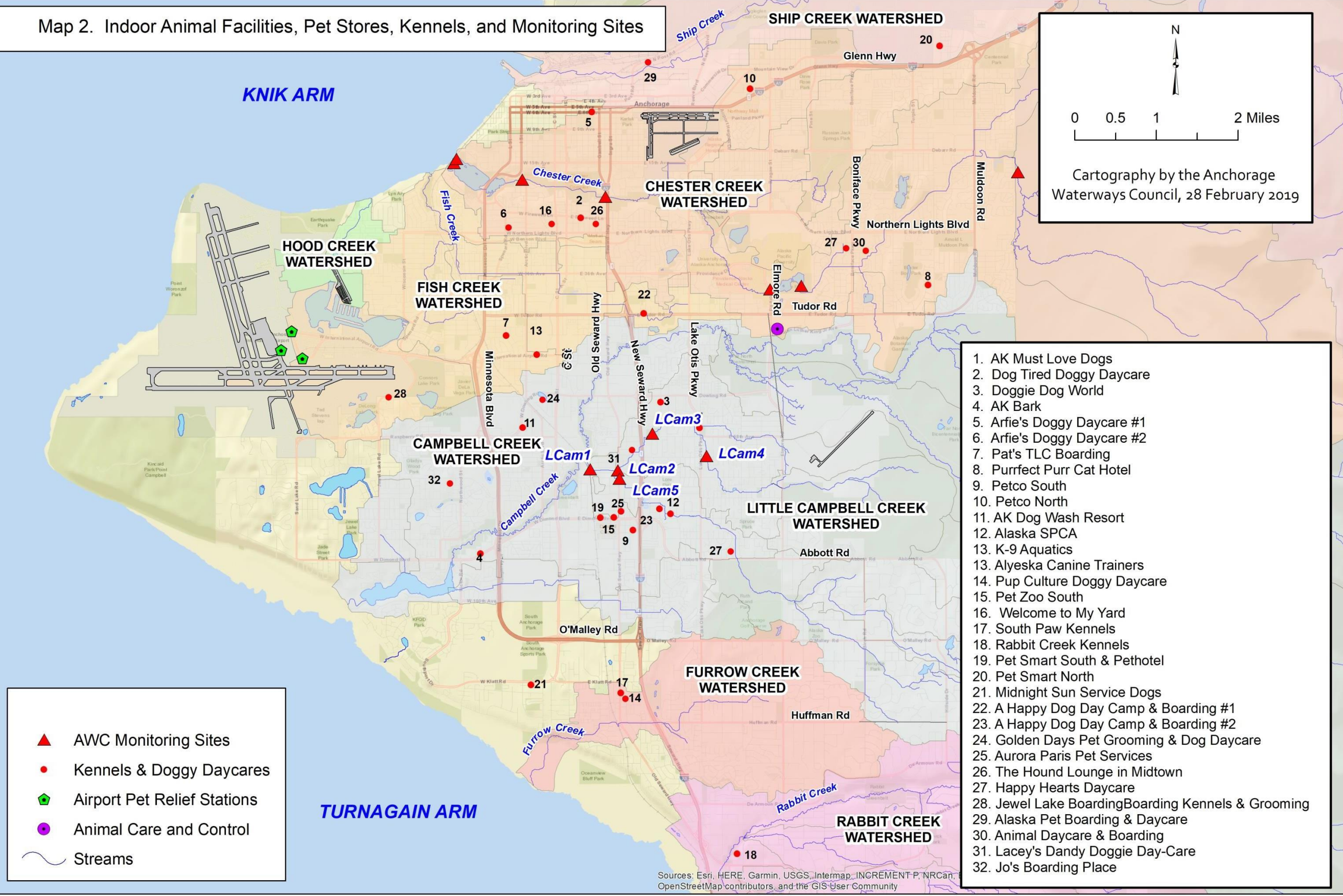
anomaly except when you take into account that this area of Anchorage is high density septic.
Davis and Davis again suggest that the Alaska Zoo could be a source of microbial contamination as well as the human markers above the zoo (2010:30).

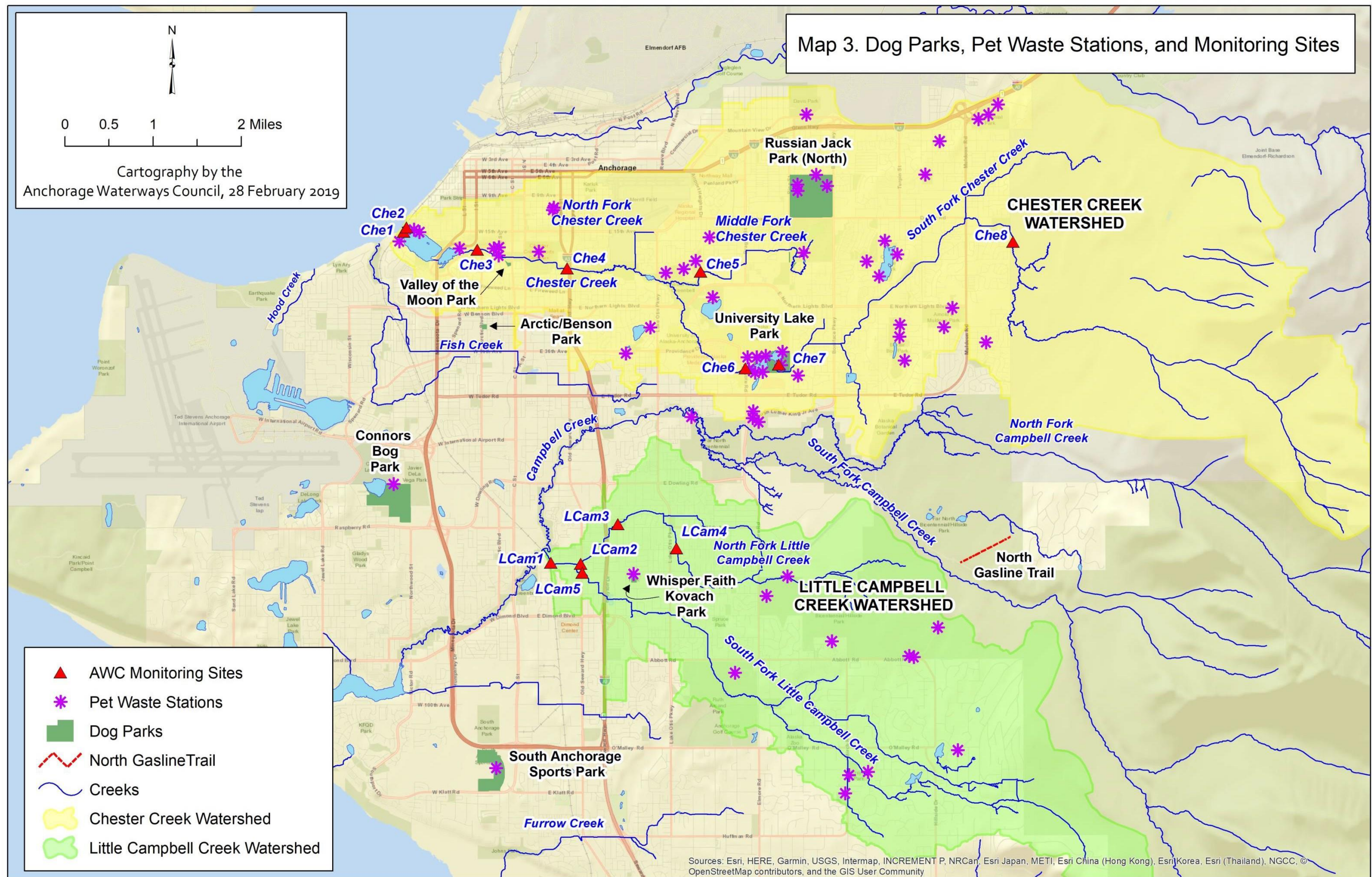
APPENDIX

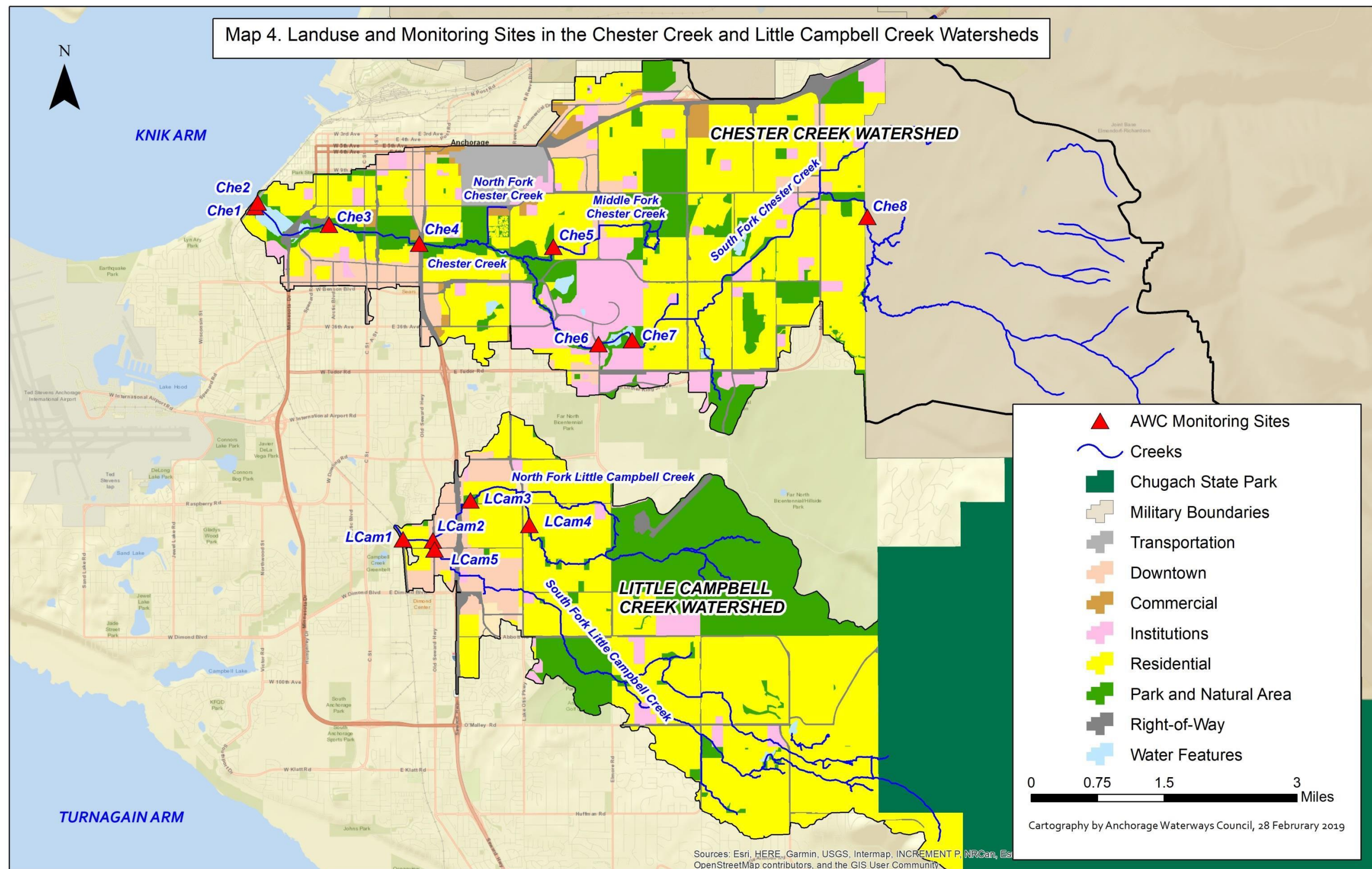
Map 1. Commercial Stables, the Alaska Zoo, Trails and LCC Monitoring Sites.



Map 2. Indoor Animal Facilities, Pet Stores, Kennels, and Monitoring Sites







BIBLIOGRAPHY

Brunett, J. (1990) *Lateral Movement of Contaminated Ground Water from Merrill Field Landfill, Anchorage, Alaska*, U.S. Geological Survey Open File Report 89-624. Retrieved from pubs.er.usgs.gov/publication/ofr89624.

Davis, J. and Gay Davis. (2010) *Fecal Coliform Bacteria Source Assessment in the Waters of Cottonwood Creek, Wasilla, and Little Campbell Creek, Anchorage*. Talkeetna, AK:Alaska Restoration and Research Institute.